

APPLICATION FOR UNITED STATES LETTERS PATENT
FOR
BROAD-SPECTRUM δ -ENDOTOXINS

BY

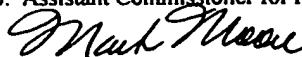
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1. BACKGROUND OF THE INVENTION.

The present application is a continuation-in-part of U. S. Patent Application Serial Number 08/754,490, filed November 20, 1996, the entire content of which is incorporated herein by reference.

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1.1 FIELD OF THE INVENTION

The present invention provides new proteins for combatting insects, and particularly, coleopteran, dipteran, and lepidopteran insects sensitive to the disclosed δ -endotoxins derived from *Bacillus thuringiensis*. The invention provides novel chimeric crystal proteins and the chimeric *cry* gene segments which encode them, as well as methods for making and using these DNA segments, methods of producing the encoded proteins, methods for making synthetically-modified chimeric crystal proteins, and methods of making and using the synthetic crystal proteins.

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1.2 DESCRIPTION OF RELATED ART

1.2.1 *B. THURINGIENSIS* CRYSTAL PROTEINS

The Gram-positive soil bacterium *B. thuringiensis* is well known for its production of proteinaceous parasporal crystals, or δ -endotoxins, that are toxic to a variety of lepidopteran, coleopteran, and dipteran larvae. *B. thuringiensis* produces crystal proteins during sporulation which are specifically toxic to certain species of insects. Many different strains of *B. thuringiensis* have been shown to produce insecticidal crystal proteins, and compositions comprising *B. thuringiensis* strains which produce proteins having insecticidal activity have been used commercially as environmentally-acceptable insecticides because of their toxicity to the specific target insect, and non-toxicity to plants and other non-targeted organisms.

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Commercial formulations of naturally occurring *B. thuringiensis* isolates have long been used for the biological control of agricultural insect pests. In commercial

production, the spores and crystals obtained from the fermentation process are concentrated and formulated for foliar application according to conventional agricultural practices.

5 1.2.2 NOMENCLATURE OF CRYSTAL PROTEINS

A review by Höfte *et al.*, (1989) describes the general state of the art with respect to the majority of insecticidal *B. thuringiensis* strains that have been identified which are active against insects of the Order Lepidoptera, *i.e.*, caterpillar insects. This treatise also describes *B. thuringiensis* strains having insecticidal activity against insects of the Orders
10 Diptera (*i.e.* flies and mosquitoes) and Coleoptera (*i.e.* beetles). A number of genes encoding crystal proteins have been cloned from several strains of *B. thuringiensis*. Höfte *et al.* (1989) discusses the genes and proteins that were identified in *B. thuringiensis* prior to 1990, and sets forth the nomenclature and classification scheme which has traditionally been applied to *B. thuringiensis* genes and proteins. *cry1* genes
15 encode lepidopteran-toxic Cry1 proteins. *cry2* genes encode Cry2 proteins that are toxic to both lepidopterans and dipterans. *cry3* genes encode coleopteran-toxic Cry3 proteins, while *cry4* genes encode dipteran-toxic Cry4 proteins, *etc.*

Recently a new nomenclature has been proposed which systematically classifies the Cry proteins based upon amino acid sequence homology rather than upon insect target
20 specificities. This classification scheme is summarized in Table 1.

TABLE 1
REVISED *B. THURINGIENSIS* δ -ENDOTOXIN NOMENCLATURE^A

New	Old	GenBank Accession #
CryIAa	CryIA(a)	M11250
CryIAb	CryIA(b)	M13898
CryIAc	CryIA(c)	M11068
CryIAd	CryIA(d)	M73250
CryIAe	CryIA(e)	M65252
CryIBa	CryIB	X06711
CryIBb	ET5	L32020
CryIBc	PEG5	Z46442
CryIBd	CryE1	U70726
CryICa	CryIC	X07518
CryICb	CryIC(b)	M97880
CryIDa	CryID	X54160
CryIDb	PrtB	Z22511
CryIEa	CryIE	X53985
CryIEb	CryIE(b)	M73253
CryIFa	CryIF	M63897
CryIFb	PrtD	Z22512
CryIGa	PrtA	Z22510
CryIGb	CryH2	U70725
CryIHa	PrtC	Z22513
CryIHb		U35780
CryIIa	CryV	X62821
CryIIb	CryV	U07642
CryIJa	ET4	L32019
CryIJb	ET1	U31527
CryIK		U28801
Cry2Aa	CryIIA	M31738
Cry2Ab	CryIIB	M23724
Cry2Ac	CryIIC	X57252
Cry3A	CryIIIA	M22472
Cry3Ba	CryIIIB	X17123
Cry3Bb	CryIIIB2	M89794
Cry3C	CryIIID	X59797
Cry4A	CryIVA	Y00423
Cry4B	CryIVB	X07423
Cry5Aa	CryVA(a)	L07025
Cry5Ab	CryVA(b)	L07026
Cry5B		U19725
Cry6A	CryVIA	L07022

New	Old	GenBank Accession #
Cry6B	CryVIB	L07024
Cry7Aa	CryIIIC	M64478
Cry7Ab	CryIIICb	U04367
Cry8A	CryIIIE	U04364
Cry8B	CryIIIG	U04365
Cry8C	CryIIIF	U04366
Cry9A	CryIG	X58120
Cry9B	CryIX	X75019
Cry9C	CryIH	Z37527
Cry10A	CryIVC	M12662
Cry11A	CryIVD	M31737
Cry11B	Jeg80	X86902
Cry12A	CryVB	L07027
Cry13A	CryVC	L07023
Cry14A	CryVD	U13955
Cry15A	34kDa	M76442
Cry16A	cbm71	X94146
Cry17A	cbm71	X99478
Cry18A	CryBP1	X99049
Cry19A	Jeg65	Y08920
Cyt1Aa	CytA	X03182
Cyt1Ab	CytM	X98793
Cyt1B		U37196
Cyt2A	CytB	Z14147
Cyt2B	CytB	U52043

^aAdapted from: http://epunix.biols.susx.ac.uk/Home/Neil_Crickmore/Bt/index.html

1.2.3 MODE OF CRYSTAL PROTEIN TOXICITY

All δ -endotoxin crystals are toxic to insect larvae by ingestion. Solubilization of the crystal in the midgut of the insect releases the protoxin form of the δ -endotoxin which, in most instances, is subsequently processed to an active toxin by midgut protease. The activated toxins recognize and bind to the brush-border of the insect midgut epithelium through receptor proteins. Several putative crystal protein receptors have been isolated from certain insect larvae (Knight *et al.*, 1995; Gill *et al.*, 1995; Masson *et al.*, 1995). The binding of active toxins is followed by intercalation and aggregation of toxin

molecules to form pores within the midgut epithelium. This process leads to osmotic imbalance, swelling, lysis of the cells lining the midgut epithelium, and eventual larvae mortality.

5 1.2.4 MOLECULAR BIOLOGY OF δ -ENDOTOXINS

With the advent of molecular genetic techniques, various δ -endotoxin genes have been isolated and their DNA sequences determined. These genes have been used to construct certain genetically engineered *B. thuringiensis* products that have been approved for commercial use. Recent developments have seen new δ -endotoxin delivery
10 systems developed, including plants that contain and express genetically engineered δ -endotoxin genes.

The cloning and sequencing of a number of δ -endotoxin genes from a variety of *Bacillus thuringiensis* strains have been described and are summarized by Höfte and Whiteley, 1989. Plasmid shuttle vectors designed for the cloning and expression of
15 δ -endotoxin genes in *E. coli* or *B. thuringiensis* are described by Gawron-Burke and Baum (1991). U. S. Patent No. 5,441,884 discloses a site-specific recombination system for constructing recombinant *B. thuringiensis* strains containing δ -endotoxin genes that are free of DNA not native to *B. thuringiensis*.

The CryI family of crystal proteins, which are primarily active against
20 lepidopteran pests, are the best studied class of δ -endotoxins. The pro-toxin form of CryI δ -endotoxins consist of two approximately equal sized segments. The carboxyl-half, or pro-toxin segment, is not toxic and is thought to be important for crystal formation (Arvidson *et al.*, 1989). The amino-half of the protoxin comprises the active-toxin segment of the CryI molecule and may be further divided into three structural domains as
25 determined by the recently described crystallographic structure for the active toxin segment of the CryIAa δ -endotoxin (Grochulski *et al.*, 1995). Domain 1 occupies the first third of the active toxin and is essential for channel formation (Thompson *et al.*, 1995). Domain 2 and domain 3 occupy the middle and last third of the active toxin,

respectively. Both domains 2 and 3 have been implicated in receptor binding and insect specificity, depending on the insect and δ -endotoxin being examined (Thompson *et al.*, 1995).

5 1.2.5 CHIMERIC CRYSTAL PROTEINS

In recent years, researchers have focused effort on the construction of hybrid δ -endotoxins with the hope of producing proteins with enhanced activity or improved properties. Advances in the art of molecular genetics over the past decade have facilitated a logical and orderly approach to engineering proteins with improved properties. Site-specific and random mutagenesis methods, the advent of polymerase chain reaction (PCRTM) methodologies, and the development of recombinant methods for generating gene fusions and constructing chimeric proteins have facilitated an assortment of methods for changing amino acid sequences of proteins, fusing portions of two or more proteins together in a single recombinant protein, and altering genetic sequences that encode proteins of commercial interest.

Unfortunately, for crystal proteins, these techniques have only been exploited in limited fashion. The likelihood of arbitrarily creating a chimeric protein with enhanced properties from portions of the numerous native proteins which have been identified is remote given the complex nature of protein structure, folding, oligomerization, activation, and correct processing of the chimeric protoxin to an active moiety. Only by careful selection of specific target regions within each protein, and subsequent protein engineering can toxins be synthesized which have improved insecticidal activity.

Some success in the area, however, has been reported in the literature. For example, the construction of a few hybrid δ -endotoxins is reported in the following related art: Intl. Pat. Appl. Publ. No. WO 95/30753 discloses the construction of hybrid *B. thuringiensis* δ -endotoxins for production in *Pseudomonas fluorescens* in which the non-toxic protoxin fragment of Cry1F has been replaced by the non-toxic protoxin fragment from the Cry1Ac/Cry1Ab that is disclosed in U. S. Patent 5,128,130.

U. S. Patent 5,128,130 discloses the construction of hybrid *B. thuringiensis* δ -endotoxins for production in *P. fluorescens* in which a portion of the non-toxic protoxin

segment of Cry1Ac is replaced with the corresponding non-toxic protoxin fragment of Cry1Ab. U. S. Patent 5,055,294 discloses the construction of a specific hybrid δ -endotoxin between Cry1Ac (amino acid residues 1-466) and Cry1Ab (amino acid residues 466-1155) for production in *P. fluorescens*. Although the aforementioned patent
5 discloses the construction of a hybrid toxin within the active toxin segment, no specifics are presented in regard to the hybrid toxin's insecticidal activity. Intl. Pat. Appl. Publ. No. WO 95/30752 discloses the construction of hybrid *B. thuringiensis* δ -endotoxins for production in *P. fluorescens* in which the non-toxic protoxin segment of Cry1C is replaced by the non-toxic protoxin segment from Cry1Ab. The aforementioned
10 application further discloses that the activity against *Spodoptera exigua* for the hybrid δ -endotoxin is improved over that of the parent active toxin, Cry1C.

Intl. Pat. Appl. Publ. No. WO 95/06730 discloses the construction of a hybrid *B. thuringiensis* δ -endotoxin consisting of domains 1 and 2 of Cry1E coupled to domain 3 and the non-toxic protoxin segment of Cry1C. Insect bioassays performed against
15 *Manduca sexta* (sensitive to Cry1C and Cry1E), *Spodoptera exigua* (sensitive to Cry1C), and *Mamestra brassicae* (sensitive to Cry1C) show that the hybrid Cry1E/Cry1C hybrid toxin is active against *M. sexta*, *S. exigua*, and *M. brassicae*. The bioassay results were expressed as EC₅₀ values (toxin concentration giving a 50% growth reduction) rather than LC₅₀ values (toxin concentration giving 50% mortality). Although the δ -endotoxins used
20 for bioassay were produced in *B. thuringiensis*, only artificially-generated active segments of the δ -endotoxins were used, not the naturally-produced crystals typically produced by *B. thuringiensis* that are present in commercial *B. thuringiensis* formulations. Bioassay results indicated that the LC₅₀ values for the hybrid Cry1E/Cry1C crystal against *S. frugiperda* were 1.5 to 1.7 fold lower (more active) than
25 for native Cry1C. This art also discloses the construction of a hybrid *B. thuringiensis* δ -endotoxin between Cry1Ab (domains 1 and 2) and Cry1C (domain 3 and the non-toxic protoxin segment), although no data are given regarding the hybrid toxin's activity or usefulness.

Lee *et al.* (1995) report the construction of hybrid *B. thuringiensis* δ -endotoxins between CryIAc and CryIAa within the active toxin segment. Artificially generated active segments of the hybrid toxins were used to examine protein interactions in susceptible insect brush border membranes vesicles (BBMV). The bioactivity of the hybrid toxins was not reported.

Honee *et al.* (1991) report the construction of hybrid δ -endotoxins between CryIC (domain 1) and CryIAb (domains 2 and 3) and the reciprocal hybrid between CryIAb (domain 1) and CryIC (domains 2 and 3). These hybrids failed to show any significant increase in activity against susceptible insects. Furthermore, the CryIC (domain 1)/CryIAb (domains 2 and 3) hybrid toxin was found to be hypersensitive to protease degradation. A report by Schnepf *et al.* (1990) discloses the construction of CryIAc hybrid toxin in which a small portion of domain 2 was replaced by the corresponding region of CryIAa, although no significant increase in activity against susceptible insect larvae was observed.

1.3 DEFICIENCIES IN THE PRIOR ART

The limited successes in producing chimeric crystal proteins which have improved activity have negatively impacted the field by thwarting efforts to produce recombinantly-engineered crystal protein for commercial development, and to extend the toxic properties and host specificities of the known endotoxins. Therefore, what is lacking in the prior art are reliable methods and compositions comprising recombinantly-engineered crystal proteins which have improved insecticidal activity, broad-host-range specificities, and which are suitable for commercial production in *B. thuringiensis*.

2. SUMMARY OF THE INVENTION

The present invention overcomes these and other limitations in the prior art by providing novel chimeric δ -endotoxins which have improved insecticidal properties, and broad-range specificities.

Disclosed are methods for the construction of *B. thuringiensis* hybrid δ -endotoxins comprising amino acid sequences from native CryIAc and CryIF crystal

proteins. These hybrid proteins, in which all or a portion of Cry1Ac domain 2, all or a portion of Cry1Ac domain 3, and all or a portion of the Cry1Ac protoxin segment is replaced by the corresponding portions of Cry1F, possess not only the insecticidal characteristics of the parent δ -endotoxins, but also have the unexpected and remarkable properties of enhanced broad-range specificity which is not proficiently displayed by either of the native δ -endotoxins from which the chimeric proteins were engineered.

Specifically, the present invention discloses and claims genetically-engineered hybrid δ -endotoxins which comprise a portion of a Cry1Ac crystal protein fused to a portion of a Cry1F crystal protein. These chimeric endotoxins have broad-range specificity for the insect pests described herein.

In a further embodiment, the present invention also discloses and claims recombinant *B. thuringiensis* hybrid δ -endotoxins which comprise a portion of Cry1Ab, Cry1F, and Cry1Ac in which all or a portion of Cry1Ab domain 2 or all or a portion of Cry1Ab domain 3 is replaced by the corresponding portions of Cry1F and all or a portion of the Cry1Ab protoxin segment is replaced by the corresponding portions of Cry1Ac. Exemplary hybrid δ -endotoxins between Cry1Ab and Cry1F are identified in SEQ ID NO:13 and SEQ ID NO:14.

One aspect of the present invention demonstrates the unexpected result that certain hybrid δ -endotoxins derived from Cry1Ac and Cry1F proteins exhibit not only the insecticidal characteristics of the parent δ -endotoxins, but also possess insecticidal activity which is not proficiently displayed by either of the parent δ -endotoxins.

Another aspect of the invention further demonstrates the unexpected result that certain chimeric Cry1Ab/Cry1F proteins maintain not only the insecticidal characteristics of the parent δ -endotoxins, but also exhibit insecticidal activity which is not displayed by either the native Cry1Ab or Cry1F endotoxins.

The present invention also encompasses Cry1Ac/Cry1F and Cry1Ab/Cry1F hybrid δ -endotoxins that maintain the desirable characteristics needed for commercial production in *B. thuringiensis*. Specifically, the hybrid δ -endotoxins identified in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, and SEQ ID NO:34 can efficiently form proteinaceous parasporal inclusions

in *B. thuringiensis* and have the favorable characteristics of solubility, protease susceptibility, and insecticidal activity of the parent δ -endotoxins.

5 In a further embodiment, the present invention also discloses and claims recombinant *B. thuringiensis* hybrid δ -endotoxins which comprise a portion of Cry1Ac and Cry1C in which all or a portion of Cry1Ac domain 3 is replaced by the corresponding portions of Cry1C and all or a portion of the Cry1Ac protoxin segment is replaced by the corresponding portion of Cry1C. Exemplary hybrid δ -endotoxins between Cry1Ac and Cry1C are identified in SEQ ID NO:29 and SEQ ID NO:30.

10 One aspect of the present invention demonstrates the unexpected result that, although neither Cry1Ac nor Cry1C possess *S. frugiperda* activity, the Cry1Ac/Cry1C hybrid δ -endotoxin identified by SEQ ID NO:29 and SEQ ID NO:30 has significant activity against *S. frugiperda*. Furthermore, the Cry1Ac/Cry1C hybrid δ -endotoxin identified by SEQ ID NO:29 and SEQ ID NO:30 has significantly better activity against *S. exigua* than the Cry1C parental δ -endotoxin.

15 The present invention further pertains to the recombinant nucleic acid sequences which encode the novel crystal proteins disclosed herein. Specifically, the invention discloses and claims the nucleic acid sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, and SEQ ID NO:33; nucleic acid sequences which are complementary to the nucleic acid sequences of
20 SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29; and SEQ ID NO:33, and nucleic acid sequences which hybridize to the sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, and SEQ ID NO:33.

25 The novel hybrid δ -endotoxins disclosed herein are useful in the control of a broad range of insect pests. These hybrid δ -endotoxins are described in FIG. 1 and FIG. 4 and are disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, and SEQ ID NO:34. The nucleic acid segments encoding these proteins are disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, and SEQ ID NO:33. The insecticidal
30 and biochemical properties of the hybrid δ -endotoxins are described in FIG. 2, FIG. 3,

and Table 4, Table 5, Table 6, and Table 7. The broad host range of the improved δ -endotoxins specified in the present invention is useful in circumventing dilution effects caused by expressing multiple δ -endotoxin genes within a single *B. thuringiensis* strain. Expression of such a broad host range δ -endotoxin in plants is expected to impart protection against a wider variety of insect pests.

The impetus for constructing these and other hybrid δ -endotoxins is to create novel toxins with improved insecticidal activity, increased host-range specificity, and improved production characteristics. The DNA sequences listed in Table 7 define the exchange points for the hybrid δ -endotoxins pertinent to the present invention and as oligonucleotide primers, may be used to identify like or similar hybrid δ -endotoxins by Southern or colony hybridization under conditions of moderate to high stringency. Researchers skilled in the art will recognize the importance of the exchange site chosen between two or more δ -endotoxins can be achieved using a number of *in vivo* or *in vitro* molecular genetic techniques. Small variations in the exchange region between two or more δ -endotoxins may yield similar results or, as demonstrated for EG11062 and EG11063, adversely affect desirable traits. Similarly, large variations in the exchange region between two or more δ -endotoxins may have no effect on desired traits, as demonstrated by EG11063 and EG11074, or may adversely affect desirable traits, as demonstrated by EG11060 and EG11063.

Favorable traits with regard to improved insecticidal activity, increased host range, and improved production characteristics may be achieved by other such hybrid δ -endotoxins including, but not limited to, the *cry1*, *cry2*, *cry3*, *cry4*, *cry5*, *cry6*, *cry7*, *cry8*, *cry9*, *cry10*, *cry11*, *cry12*, *cry13*, *cry14*, *cry15* class of δ -endotoxin genes and the *B. thuringiensis* cytolytic *cyt1* and *cyt2* genes. Members of these classes of *B. thuringiensis* insecticidal proteins include, but are not limited to Cry1Aa, Cry1Ab, Cry1Ac, Cry1Ad, Cry1Ae, Cry1Ba, Cry1Bb, Cry1Ca, Cry1Cb, Cry1Da, Cry1Db, Cry1Ea, Cry1Eb, Cry1Fa, Cry1Fb, Cry1Ga, Cry1Ha, Cry2a, Cry2b, Cry1Ja, Cry1Ka, Cry11Aa, Cry11Ab, Cry12Aa, Cry3Ba, Cry3Bb, Cry3C, Cry4a, Cry4Ba, Cry5a, Cry5Ab, Cry6Aa, Cry6Ba, Cry7Aa, Cry7Ab, Cry8Aa, Cry8Ba, Cry8Ca, Cry9Aa, Cry9Ba, Cry9Ca, Cry10Aa, Cry11Aa, Cry12Aa, Cry13Aa, Cry14Aa, Cry15Aa, Cyt1Aa, and Cyt2Aa. Related hybrid

5 δ -endotoxins would consist of the amino portion of one of the aforementioned δ -endotoxins, including all or part of domain 1 or domain 2, fused to all or part of domain 3 from another of the aforementioned δ -endotoxins. The non-active protoxin fragment of such hybrid δ -endotoxins may consist of the protoxin fragment from any of the
10 aforementioned δ -endotoxins which may act to stabilize the hybrid δ -endotoxin as demonstrated by EG11087 and EG11091 (see *e.g.*, Table 4). Hybrid δ -endotoxins possessing similar traits as those described in the present invention could be constructed by conservative, or "similar" replacements of amino acids within hybrid δ -endotoxins. Such substitutions would mimic the biochemical and biophysical properties of the native
15 amino acid at any position in the protein. Amino acids considered similar include for example, but are not limited to:

Ala, Ser, and Thr;
Asp and Glu;
Asn and Gln;
15 Lys and Arg;
Ile, Leu, Met, and Val; and
Phe, Tyr, and Trp.

20 Researchers skilled in the art will recognize that improved insecticidal activity, increased host range, and improved production characteristics imparted upon hybrid δ -endotoxins may be further improved by altering the genetic code for one or more amino acid positions in the hybrid δ -endotoxin such that the position, or positions, is replaced by any other amino acid. This may be accomplished by targeting a region or regions of the protein for mutagenesis by any number of established mutagenic techniques, including those procedures relevant to the present invention. Such techniques include site-specific
25 mutagenesis (Kunkle, 1985; Kunkle *et al.*, 1987), DNA shuffling (Stemmer, 1994), and PCRTM overlap extension (Horton *et al.*, 1989). Since amino acids situated at or near the surface of a protein are likely responsible for its interaction with other proteinaceous or non-proteinaceous moieties, they may serve as "target" regions for mutagenesis. Such surface exposed regions may consist of, but not be limited to, surface exposed amino acid
30 residues within the active toxin fragment of the protein and include the inter- α -helical or

inter- β -strand "loop" -regions of δ -endotoxins that separate α -helices within domain 1 and β -strands within domain 2 and domain 3. Such procedures may favorably change the protein's biochemical and biophysical characteristics or its mode of action as outlined in the Section 1. These include, but are not limited to: 1) improved crystal formation, 2) improved protein stability or reduced protease degradation, 3) improved insect membrane receptor recognition and binding, 4) improved oligomerization or channel formation in the insect midgut endothelium, and 5) improved insecticidal activity or insecticidal specificity due to any or all of the reasons stated above.

10 2.1 CRYSTAL PROTEIN TRANSGENES AND TRANSGENIC PLANTS

In yet another aspect, the present invention provides methods for producing a transgenic plant which expresses a nucleic acid segment encoding the novel chimeric crystal proteins of the present invention. The process of producing transgenic plants is well-known in the art. In general, the method comprises transforming a suitable host cell with a DNA segment which contains a promoter operatively linked to a coding region that encodes a *B. thuringiensis* Cry1Ac-1F or Cry1Ab-1F, Cry1Ac-1C, or a Cry1Ab-1Ac-1F chimeric crystal protein. Such a coding region is generally operatively linked to a transcription-terminating region, whereby the promoter is capable of driving the transcription of the coding region in the cell, and hence providing the cell the ability to produce the recombinant protein *in vivo*. Alternatively, in instances where it is desirable to control, regulate, or decrease the amount of a particular recombinant crystal protein expressed in a particular transgenic cell, the invention also provides for the expression of crystal protein antisense mRNA. The use of antisense mRNA as a means of controlling or decreasing the amount of a given protein of interest in a cell is well-known in the art.

25 Another aspect of the invention comprises a transgenic plant which express a gene or gene segment encoding one or more of the novel polypeptide compositions disclosed herein. As used herein, the term "transgenic plant" is intended to refer to a plant that has incorporated DNA sequences, including but not limited to genes which are perhaps not normally present, DNA sequences not normally transcribed into RNA or translated into a protein ("expressed"), or any other genes or DNA sequences which one desires to

introduce into the non-transformed plant, such as genes which may normally be present in the non-transformed plant but which one desires to either genetically engineer or to have altered expression. The construction and expression of synthetic *B. thuringiensis* genes in plants has been described in detail in U. S. Patents 5,500,365 and 5,380,831 (each specifically incorporated herein by reference).

It is contemplated that in some instances the genome of a transgenic plant of the present invention will have been augmented through the stable introduction of one or more *cryIAc-IF*, *cryIAb-IF*, *cryIAc-IC*, or *cryIAb-IaC-IF* transgenes, either native, synthetically-modified, or further mutated. In some instances, more than one transgene will be incorporated into the genome of the transformed host plant cell. Such is the case when more than one crystal protein-encoding DNA segment is incorporated into the genome of such a plant. In certain situations, it may be desirable to have one, two, three, four, or even more *B. thuringiensis* crystal proteins (either native or recombinantly-engineered) incorporated and stably expressed in the transformed transgenic plant.

A preferred gene, such as those disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, and SEQ ID NO:33 which may be introduced includes, for example, a crystal protein-encoding a DNA sequence from bacterial origin, and particularly one or more of those described herein which are obtained from *Bacillus* spp. Highly preferred nucleic acid sequences are those obtained from *B. thuringiensis*, or any of those sequences which have been genetically engineered to decrease or increase the insecticidal activity of the crystal protein in such a transformed host cell.

Means for transforming a plant cell and the preparation of a transgenic cell line are well-known in the art, and are discussed herein. Vectors, plasmids, cosmids, yeast artificial chromosomes (YACs) and nucleic acid segments for use in transforming such cells will, of course, generally comprise either the operons, genes, or gene-derived sequences of the present invention, either native, or synthetically-derived, and particularly those encoding the disclosed crystal proteins. These DNA constructs can further include structures such as promoters, enhancers, polylinkers, or even gene sequences which have positively- or negatively-regulating activity upon the particular

genes of interest as desired. The DNA segment or gene may encode either a native or modified crystal protein, which will be expressed in the resultant recombinant cells, and/or which will impart an improved phenotype to the regenerated plant. Nucleic acid sequences optimized for expression in plants have been disclosed in Intl. Pat. Appl. Publ. No. WO 93/07278 (specifically incorporated herein by reference).

Such transgenic plants may be desirable for increasing the insecticidal resistance of a monocotyledonous or dicotyledonous plant, by incorporating into such a plant, a transgenic DNA segment encoding Cry1Ac-1F and/or Cry1Ac-1C, and/or Cry1Ab-1F and/or Cry1Ab-1Ac-1F, crystal protein(s) which possess broad-insect specificity. Particularly preferred plants such as grains, including but not limited to corn, wheat, oats, rice, maize, and barley; cotton; soybeans and other legumes; trees, including but not limited to ornamentals, shrubs, fruits, nuts; vegetables, turf and pasture grasses, berries, citrus, and other crops of commercial interest; such as garden crops and/or houseplants, succulents, cacti, and flowering species.

In a related aspect, the present invention also encompasses a seed produced by the transformed plant, a progeny from such seed, and a seed produced by the progeny of the original transgenic plant, produced in accordance with the above process. Such progeny and seeds will have a stably crystal protein transgene stably incorporated into its genome, and such progeny plants will inherit the traits afforded by the introduction of a stable transgene in Mendelian fashion. All such transgenic plants having incorporated into their genome transgenic DNA segments encoding one or more chimeric crystal proteins or polypeptides are aspects of this invention.

2.2 CRYSTAL PROTEIN SCREENING AND IMMUNODETECTION KITS

The present invention contemplates methods and kits for screening samples suspected of containing crystal protein polypeptides or crystal protein-related polypeptides, or cells producing such polypeptides. Exemplary proteins include those disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, and SEQ ID NO:34. Said kit can contain a nucleic acid segment or an antibody of the present invention. The kit can contain reagents for

detecting an interaction between a sample and a nucleic acid or antibody of the present invention. The provided reagent can be radio-, fluorescently- or enzymatically-labeled. The kit can contain a known radiolabeled agent capable of binding or interacting with a nucleic acid or antibody of the present invention.

5 The reagent of the kit can be provided as a liquid solution, attached to a solid support or as a dried powder. Preferably, when the reagent is provided in a liquid solution, the liquid solution is an aqueous solution. Preferably, when the reagent provided is attached to a solid support, the solid support can be chromatograph media, a test plate having a plurality of wells, or a microscope slide. When the reagent provided is
10 a dry powder, the powder can be reconstituted by the addition of a suitable solvent, that may be provided.

 In still further embodiments, the present invention concerns immunodetection methods and associated kits. It is proposed that the crystal proteins or peptides of the present invention may be employed to detect antibodies having reactivity therewith, or,
15 alternatively, antibodies prepared in accordance with the present invention, may be employed to detect crystal proteins or crystal protein-related epitope-containing peptides. In general, these methods will include first obtaining a sample suspected of containing such a protein, peptide or antibody, contacting the sample with an antibody or peptide in accordance with the present invention, as the case may be, under conditions effective to
20 allow the formation of an immunocomplex, and then detecting the presence of the immunocomplex.

 In general, the detection of immunocomplex formation is quite well known in the art and may be achieved through the application of numerous approaches. For example, the present invention contemplates the application of ELISA, RIA, immunoblot (*e.g.*, dot
25 blot), indirect immunofluorescence techniques and the like. Generally, immunocomplex formation will be detected through the use of a label, such as a radiolabel or an enzyme tag (such as alkaline phosphatase, horseradish peroxidase, or the like). Of course, one may find additional advantages through the use of a secondary binding ligand such as a second antibody or a biotin/avidin ligand binding arrangement, as is known in the art.

For assaying purposes, it is proposed that virtually any sample suspected of comprising either a crystal protein or peptide or a crystal protein-related peptide or antibody sought to be detected, as the case may be, may be employed. It is contemplated that such embodiments may have application in the titering of antigen or antibody samples, in the selection of hybridomas, and the like. In related embodiments, the present invention contemplates the preparation of kits that may be employed to detect the presence of crystal proteins or related peptides and/or antibodies in a sample. Samples may include cells, cell supernatants, cell suspensions, cell extracts, enzyme fractions, protein extracts, or other cell-free compositions suspected of containing crystal proteins or peptides. Generally speaking, kits in accordance with the present invention will include a suitable crystal protein, peptide or an antibody directed against such a protein or peptide, together with an immunodetection reagent and a means for containing the antibody or antigen and reagent. The immunodetection reagent will typically comprise a label associated with the antibody or antigen, or associated with a secondary binding ligand. Exemplary ligands might include a secondary antibody directed against the first antibody or antigen or a biotin or avidin (or streptavidin) ligand having an associated label. Of course, as noted above, a number of exemplary labels are known in the art and all such labels may be employed in connection with the present invention.

The container will generally include a vial into which the antibody, antigen or detection reagent may be placed, and preferably suitably aliquotted. The kits of the present invention will also typically include a means for containing the antibody, antigen, and reagent containers in close confinement for commercial sale. Such containers may include injection or blow-molded plastic containers into which the desired vials are retained.

2.3 ELISAS AND IMMUNOPRECIPITATION

ELISAs may be used in conjunction with the invention. In an ELISA assay, proteins or peptides incorporating crystal protein antigen sequences are immobilized onto a selected surface, preferably a surface exhibiting a protein affinity such as the wells of a polystyrene microtiter plate. After washing to remove incompletely adsorbed material, it

is desirable to bind or coat the assay plate wells with a nonspecific protein that is known to be antigenically neutral with regard to the test antisera such as bovine serum albumin (BSA), casein or solutions of milk powder. This allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific binding of antisera onto the surface.

After binding of antigenic material to the well, coating with a non-reactive material to reduce background, and washing to remove unbound material, the immobilizing surface is contacted with the antisera or clinical or biological extract to be tested in a manner conducive to immune complex (antigen/antibody) formation. Such conditions preferably include diluting the antisera with diluents such as BSA, bovine gamma globulin (BGG) and phosphate buffered saline (PBS)/Tween[®]. These added agents also tend to assist in the reduction of nonspecific background. The layered antisera is then allowed to incubate for from about 2 to about 4 hours, at temperatures preferably on the order of about 25° to about 27°C. Following incubation, the antisera-contacted surface is washed so as to remove non-immunocomplexed material. A preferred washing procedure includes washing with a solution such as PBS/Tween[®], or borate buffer.

Following formation of specific immunocomplexes between the test sample and the bound antigen, and subsequent washing, the occurrence and even amount of immunocomplex formation may be determined by subjecting same to a second antibody having specificity for the first. To provide a detecting means, the second antibody will preferably have an associated enzyme that will generate a color development upon incubating with an appropriate chromogenic substrate. Thus, for example, one will desire to contact and incubate the antisera-bound surface with a urease or peroxidase-conjugated anti-human IgG for a period of time and under conditions which favor the development of immunocomplex formation (e.g., incubation for 2 hours at room temperature in a PBS-containing solution such as PBS-Tween[®]).

After incubation with the second enzyme-tagged antibody, and subsequent to washing to remove unbound material, the amount of label is quantified by incubation with a chromogenic substrate such as urea and bromocresol purple or 2, 2'-azino-di-(3-

ethyl-benzthiazoline)-6-sulfonic acid (ABTS) and H_2O_2 , in the case of peroxidase as the enzyme label. Quantitation is then achieved by measuring the degree of color generation, *e.g.*, using a visible spectra spectrophotometer.

5 The anti-crystal protein antibodies of the present invention are particularly useful for the isolation of other crystal protein antigens by immunoprecipitation. Immunoprecipitation involves the separation of the target antigen component from a complex mixture, and is used to discriminate or isolate minute amounts of protein. For the isolation of membrane proteins cells must be solubilized into detergent micelles. Nonionic salts are preferred, since other agents such as bile salts, precipitate at acid pH or
10 in the presence of bivalent cations.

 In an alternative embodiment the antibodies of the present invention are useful for the close juxtaposition of two antigens. This is particularly useful for increasing the localized concentration of antigens, *e.g.* enzyme-substrate pairs.

15 2.4 WESTERN BLOTS

 The compositions of the present invention will find great use in immunoblot or western blot analysis. The anti-peptide antibodies may be used as high-affinity primary reagents for the identification of proteins immobilized onto a solid support matrix, such as nitrocellulose, nylon or combinations thereof. In conjunction with
20 immunoprecipitation, followed by gel electrophoresis, these may be used as a single step reagent for use in detecting antigens against which secondary reagents used in the detection of the antigen cause an adverse background. This is especially useful when the antigens studied are immunoglobulins (precluding the use of immunoglobulins binding bacterial cell wall components), the antigens studied cross-react with the detecting agent,
25 or they migrate at the same relative molecular weight as a cross-reacting signal.

 Immunologically-based detection methods for use in conjunction with Western blotting include enzymatically-, radiolabel-, or fluorescently-tagged secondary antibodies against the toxin moiety are considered to be of particular use in this regard.

2.5 EPITOPIC CORE SEQUENCES

The present invention is also directed to protein or peptide compositions, free from total cells and other peptides, which comprise a purified protein or peptide which incorporates an epitope that is immunologically cross-reactive with one or more anti-crystal protein antibodies. In particular, the invention concerns epitopic core sequences derived from Cry proteins or peptides.

As used herein, the term "incorporating an epitope(s) that is immunologically cross-reactive with one or more anti-crystal protein antibodies" is intended to refer to a peptide or protein antigen which includes a primary, secondary or tertiary structure similar to an epitope located within a crystal protein or polypeptide. The level of similarity will generally be to such a degree that monoclonal or polyclonal antibodies directed against the crystal protein or polypeptide will also bind to, react with, or otherwise recognize, the cross-reactive peptide or protein antigen. Various immunoassay methods may be employed in conjunction with such antibodies, such as, for example, Western blotting, ELISA, RIA, and the like, all of which are known to those of skill in the art.

The identification of Cry immunodominant epitopes, and/or their functional equivalents, suitable for use in vaccines is a relatively straightforward matter. For example, one may employ the methods of Hopp, as taught in U. S. Patent 4,554,101, incorporated herein by reference, which teaches the identification and preparation of epitopes from amino acid sequences on the basis of hydrophilicity. The methods described in several other papers, and software programs based thereon, can also be used to identify epitopic core sequences (see, for example, Jameson and Wolf, 1988; Wolf *et al.*, 1988; U. S. Patent. 4,554,101). The amino acid sequence of these "epitopic core sequences" may then be readily incorporated into peptides, either through the application of peptide synthesis or recombinant technology.

Preferred peptides for use in accordance with the present invention will generally be on the order of about 8 to about 20 amino acids in length, and more preferably about 8 to about 15 amino acids in length. It is proposed that shorter antigenic crystal protein-derived peptides will provide advantages in certain circumstances, for example, in the

preparation of immunologic detection assays. Exemplary advantages include the ease of preparation and purification, the relatively low cost and improved reproducibility of production, and advantageous biodistribution.

5 It is proposed that particular advantages of the present invention may be realized through the preparation of synthetic peptides which include modified and/or extended epitopic/immunogenic core sequences which result in a "universal" epitopic peptide directed to crystal proteins, and in particular Cry and Cry-related sequences. These epitopic core sequences are identified herein in particular aspects as hydrophilic regions of the particular polypeptide antigen. It is proposed that these regions represent those
10 which are most likely to promote T-cell or B-cell stimulation, and, hence, elicit specific antibody production.

An epitopic core sequence, as used herein, is a relatively short stretch of amino acids that is "complementary" to, and therefore will bind, antigen binding sites on the crystal protein-directed antibodies disclosed herein. Additionally or alternatively, an
15 epitopic core sequence is one that will elicit antibodies that are cross-reactive with antibodies directed against the peptide compositions of the present invention. It will be understood that in the context of the present disclosure, the term "complementary" refers to amino acids or peptides that exhibit an attractive force towards each other. Thus, certain epitope core sequences of the present invention may be operationally defined in
20 terms of their ability to compete with or perhaps displace the binding of the desired protein antigen with the corresponding protein-directed antisera.

In general, the size of the polypeptide antigen is not believed to be particularly crucial, so long as it is at least large enough to carry the identified core sequence or sequences. The smallest useful core sequence anticipated by the present disclosure would
25 generally be on the order of about 8 amino acids in length, with sequences on the order of 10 to 20 being more preferred. Thus, this size will generally correspond to the smallest peptide antigens prepared in accordance with the invention. However, the size of the antigen may be larger where desired, so long as it contains a basic epitopic core sequence.

The identification of epitopic core sequences is known to those of skill in the art,
30 for example, as described in U. S. Patent 4,554,101, incorporated herein by reference,

which teaches the identification and preparation of epitopes from amino acid sequences on the basis of hydrophilicity. Moreover, numerous computer programs are available for use in predicting antigenic portions of proteins (see *e.g.*, Jameson and Wolf, 1988; Wolf *et al.*, 1988). Computerized peptide sequence analysis programs (*e.g.*, DNASTar[®] software, DNASTar, Inc., Madison, WI) may also be useful in designing synthetic peptides in accordance with the present disclosure.

Syntheses of epitopic sequences, or peptides which include an antigenic epitope within their sequence, are readily achieved using conventional synthetic techniques such as the solid phase method (*e.g.*, through the use of commercially available peptide synthesizer such as an Applied Biosystems Model 430A Peptide Synthesizer). Peptide antigens synthesized in this manner may then be aliquotted in predetermined amounts and stored in conventional manners, such as in aqueous solutions or, even more preferably, in a powder or lyophilized state pending use.

In general, due to the relative stability of peptides, they may be readily stored in aqueous solutions for fairly long periods of time if desired, *e.g.*, up to six months or more, in virtually any aqueous solution without appreciable degradation or loss of antigenic activity. However, where extended aqueous storage is contemplated it will generally be desirable to include agents including buffers such as Tris or phosphate buffers to maintain a pH of about 7.0 to about 7.5. Moreover, it may be desirable to include agents which will inhibit microbial growth, such as sodium azide or Merthiolate. For extended storage in an aqueous state it will be desirable to store the solutions at about 4°C, or more preferably, frozen. Of course, where the peptides are stored in a lyophilized or powdered state, they may be stored virtually indefinitely, *e.g.*, in metered aliquots that may be rehydrated with a predetermined amount of water (preferably distilled) or buffer prior to use.

2.6 NUCLEIC ACID SEGMENTS ENCODING CRYSTAL PROTEIN CHIMERAS

The present invention also concerns DNA segments, both native, synthetic, and mutagenized, that can be synthesized, or isolated from virtually any source, that are free from total genomic DNA and that encode the novel chimeric peptides disclosed herein.

DNA segments encoding these peptide species may prove to encode proteins, polypeptides, subunits, functional domains, and the like of crystal protein-related or other non-related gene products. In addition these DNA segments may be synthesized entirely *in vitro* using methods that are well-known to those of skill in the art.

5 As used herein, the term "DNA segment" refers to a DNA molecule that has been isolated free of total genomic DNA of a particular species. Therefore, a DNA segment encoding a crystal protein or peptide refers to a DNA segment that contains crystal protein coding sequences yet is isolated away from, or purified free from, total genomic DNA of the species from which the DNA segment is obtained, which in the instant case
10 is the genome of the Gram-positive bacterial genus, *Bacillus*, and in particular, the species of *Bacillus* known as *B. thuringiensis*. Included within the term "DNA segment", are DNA segments and smaller fragments of such segments, and also recombinant vectors, including, for example, plasmids, cosmids, phagemids, phage, viruses, and the like.

15 Similarly, a DNA segment comprising an isolated or purified crystal protein-encoding gene refers to a DNA segment which may include in addition to peptide encoding sequences, certain other elements such as, regulatory sequences, isolated substantially away from other naturally occurring genes or protein-encoding sequences. In this respect, the term "gene" is used for simplicity to refer to a functional protein-,
20 polypeptide- or peptide-encoding unit. As will be understood by those in the art, this functional term includes both genomic sequences, operon sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides or peptides.

 "Isolated substantially away from other coding sequences" means that the gene of
25 interest, in this case, a gene encoding a bacterial crystal protein, forms the significant part of the coding region of the DNA segment, and that the DNA segment does not contain large portions of naturally-occurring coding DNA, such as large chromosomal fragments or other functional genes or operon coding regions. Of course, this refers to the DNA segment as originally isolated, and does not exclude genes, recombinant genes, synthetic
30 linkers, or coding regions later added to the segment by the hand of man.

In particular embodiments, the invention concerns isolated DNA segments and recombinant vectors incorporating DNA sequences that encode a Cry peptide species that includes within its amino acid sequence an amino acid sequence essentially as set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:34.

The term "a sequence essentially as set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34" means that the sequence substantially corresponds to a portion of the sequence of either SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34 and has relatively few amino acids that are not identical to, or a biologically functional equivalent of, the amino acids of any of these sequences. The term "biologically functional equivalent" is well understood in the art and is further defined in detail herein (*e.g.*, see Illustrative Embodiments). Accordingly, sequences that have between about 70% and about 80%, or more preferably between about 81% and about 90%, or even more preferably between about 91% and about 99% amino acid sequence identity or functional equivalence to the amino acids of SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34 will be sequences that are "essentially as set forth in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34."

It will also be understood that amino acid and nucleic acid sequences may include additional residues, such as additional N- or C-terminal amino acids or 5' or 3' sequences, and yet still be essentially as set forth in one of the sequences disclosed herein, so long as the sequence meets the criteria set forth above, including the maintenance of biological protein activity where protein expression is concerned. The addition of terminal sequences particularly applies to nucleic acid sequences that may, for example, include various non-coding sequences flanking either of the 5' or 3' portions of the coding region or may include various internal sequences, *i.e.*, introns, which are known to occur within genes.

The nucleic acid segments of the present invention, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is
5 therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For example, nucleic acid fragments may be prepared that include a short contiguous stretch encoding either of the peptide sequences disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26,
10 SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34, or that are identical to or complementary to DNA sequences which encode any of the peptides disclosed in SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34, and particularly those DNA segments disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27,
15 SEQ ID NO:29, or SEQ ID NO:33. For example, DNA sequences such as about 14 nucleotides, and that are up to about 10,000, about 5,000, about 3,000, about 2,000, about 1,000, about 500, about 200, about 100, about 50, and about 14 base pairs in length (including all intermediate lengths) are also contemplated to be useful.

It will be readily understood that "intermediate lengths", in these contexts, means
20 any length between the quoted ranges, such as 14, 15, 16, 17, 18, 19, 20, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103, *etc.*; 150, 151, 152, 153, *etc.*; including all integers through the 200-500; 500-1,000; 1,000-2,000; 2,000-3,000; 3,000-5,000; and up to and including sequences of about 10,000 nucleotides and the like.

It will also be understood that this invention is not limited to the particular nucleic
25 acid sequences which encode peptides of the present invention, or which encode the amino acid sequences of SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34, including those DNA sequences which are particularly disclosed in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, or SEQ ID NO:33.
30 Recombinant vectors and isolated DNA segments may therefore variously include the

peptide-coding regions themselves, coding regions bearing selected alterations or modifications in the basic coding region, or they may encode larger polypeptides that nevertheless include these peptide-coding regions or may encode biologically functional equivalent proteins or peptides that have variant amino acids sequences.

5 The DNA segments of the present invention encompass biologically-functional, equivalent peptides. Such sequences may arise as a consequence of codon redundancy and functional equivalency that are known to occur naturally within nucleic acid sequences and the proteins thus encoded. Alternatively, functionally-equivalent proteins or peptides may be created via the application of recombinant DNA technology, in which
10 changes in the protein structure may be engineered, based on considerations of the properties of the amino acids being exchanged. Changes designed by man may be introduced through the application of site-directed mutagenesis techniques, *e.g.*, to introduce improvements to the antigenicity of the protein or to test mutants in order to examine activity at the molecular level.

15 If desired, one may also prepare fusion proteins and peptides, *e.g.*, where the peptide-coding regions are aligned within the same expression unit with other proteins or peptides having desired functions, such as for purification or immunodetection purposes (*e.g.*, proteins that may be purified by affinity chromatography and enzyme label coding regions, respectively).

20 Recombinant vectors form further aspects of the present invention. Particularly useful vectors are contemplated to be those vectors in which the coding portion of the DNA segment, whether encoding a full length protein or smaller peptide, is positioned under the control of a promoter. The promoter may be in the form of the promoter that is naturally associated with a gene encoding peptides of the present invention, as may be
25 obtained by isolating the 5' non-coding sequences located upstream of the coding segment or exon, for example, using recombinant cloning and/or PCR™ technology, in connection with the compositions disclosed herein.

2.7 RECOMBINANT VECTORS AND PROTEIN EXPRESSION

In other embodiments, it is contemplated that certain advantages will be gained by positioning the coding DNA segment under the control of a recombinant, or heterologous, promoter. As used herein, a recombinant or heterologous promoter is intended to refer to a promoter that is not normally associated with a DNA segment encoding a crystal protein or peptide in its natural environment. Such promoters may include promoters normally associated with other genes, and/or promoters isolated from any bacterial, viral, eukaryotic, or plant cell. Naturally, it will be important to employ a promoter that effectively directs the expression of the DNA segment in the cell type, organism, or even animal, chosen for expression. The use of promoter and cell type combinations for protein expression is generally known to those of skill in the art of molecular biology, for example, see Sambrook *et al.*, 1989. The promoters employed may be constitutive, or inducible, and can be used under the appropriate conditions to direct high level expression of the introduced DNA segment, such as is advantageous in the large-scale production of recombinant proteins or peptides. Appropriate promoter systems contemplated for use in high-level expression include, but are not limited to, the *Pichia* expression vector system (Pharmacia LKB Biotechnology).

In connection with expression embodiments to prepare recombinant proteins and peptides, it is contemplated that longer DNA segments will most often be used, with DNA segments encoding the entire peptide sequence being most preferred. However, it will be appreciated that the use of shorter DNA segments to direct the expression of crystal peptides or epitopic core regions, such as may be used to generate anti-crystal protein antibodies, also falls within the scope of the invention. DNA segments that encode peptide antigens from about 8 to about 50 amino acids in length, or more preferably, from about 8 to about 30 amino acids in length, or even more preferably, from about 8 to about 20 amino acids in length are contemplated to be particularly useful. Such peptide epitopes may be amino acid sequences which comprise contiguous amino acid sequences from SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34; or any peptide epitope encoded by

the nucleic acid sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, SEQ ID NO:33.

Methods for the recombinant expression of crystal proteins and vectors useful in the expression of DNA constructs encoding crystal proteins are described in Intl. Pat. Appl. Publ. No. WO 95/02058, specifically incorporated herein by reference.

2.8 RECOMBINANT HOST CELLS

TABLE 2

STRAINS DEPOSITED WITH NRRL

STRAIN	PLASMID	ACCESSION NUMBER	DEPOSIT DATE
EG 11063	pEG1068	B-21579	June 26, 1996
EG11074	pEG1077	B-21580	June 26, 1996
EG11091	pEG1092	B-21780	May XX, 1997
EG11092	pEG1093	B-21635	November 14, 1996
EG11735	pEG365	B-21581	June 26, 1996
EG11751	pEG378	B-21636	November 14, 1996
EG11768	pEG381	B-21781	May XX, 1997

2.9 DNA SEGMENTS AS HYBRIDIZATION PROBES AND PRIMERS

In addition to their use in directing the expression of crystal proteins or peptides of the present invention, the nucleic acid sequences contemplated herein also have a variety of other uses. For example, they also have utility as probes or primers in nucleic acid hybridization embodiments. As such, it is contemplated that nucleic acid segments that comprise a sequence region that consists of at least a 14 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 14 nucleotide long contiguous DNA segment of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, or SEQ ID NO:33 will find particular utility. Also, nucleic acid segments which encode at least a 6 amino acid contiguous sequence from SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34, are also preferred. Longer

contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000, 2000, 5000, 10000 *etc.* (including all intermediate lengths and up to and including full-length sequences will also be of use in certain embodiments.

5 The ability of such nucleic acid probes to specifically hybridize to crystal protein-encoding sequences will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are envisioned, including the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

10 Nucleic acid molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so, identical or complementary to DNA sequences of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, or SEQ ID NO:33, are particularly contemplated as hybridization probes for use in, *e.g.*, Southern and Northern blotting. Smaller fragments will generally find use in hybridization embodiments,
15 wherein the length of the contiguous complementary region may be varied, such as between about 10-14 and about 100 or 200 nucleotides, but larger contiguous complementarity stretches may be used, according to the length complementary sequences one wishes to detect.

Of course, fragments may also be obtained by other techniques such as, *e.g.*, by
20 mechanical shearing or by restriction enzyme digestion. Small nucleic acid segments or fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patents 4,683,195 and 4,683,202
25 (each specifically incorporated herein by reference), by introducing selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular biology.

Accordingly, the nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of DNA
30 fragments. Depending on the application envisioned, one will desire to employ varying

conditions of hybridization to achieve varying degrees of selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, e.g., one will select relatively low salt and/or high temperature conditions, such as provided by about 0.02 M to about 0.15 M NaCl at temperatures of about 50°C to about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating crystal protein-encoding DNA segments. Detection of DNA segments via hybridization is well-known to those of skill in the art, and the teachings of U. S. Patents 4,965,188 and 5,176,995 (each specifically incorporated herein by reference) are exemplary of the methods of hybridization analyses. Teachings such as those found in the texts of Maloy *et al.*, 1994; Segal 1976; Prokop, 1991; and Kuby, 1994, are particularly relevant.

Of course, for some applications, for example, where one desires to prepare mutants employing a mutant primer strand hybridized to an underlying template or where one seeks to isolate crystal protein-encoding sequences from related species, functional equivalents, or the like, less stringent hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ conditions such as about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

In certain embodiments, it will be advantageous to employ nucleic acid sequences of the present invention in combination with an appropriate means, such as a label, for determining hybridization. A wide variety of appropriate indicator means are known in the art, including fluorescent, radioactive, enzymatic or other ligands, such as avidin/biotin, which are capable of giving a detectable signal. In preferred embodiments,

one will likely desire to employ a fluorescent label or an enzyme tag, such as urease, alkaline phosphatase or peroxidase, instead of radioactive or other environmental undesirable reagents. In the case of enzyme tags, colorimetric indicator substrates are known that can be employed to provide a means visible to the human eye or spectrophotometrically, to identify specific hybridization with complementary nucleic acid-containing samples.

In general, it is envisioned that the hybridization probes described herein will be useful both as reagents in solution hybridization as well as in embodiments employing a solid phase. In embodiments involving a solid phase, the test DNA (or RNA) is adsorbed or otherwise affixed to a selected matrix or surface. This fixed, single-stranded nucleic acid is then subjected to specific hybridization with selected probes under desired conditions. The selected conditions will depend on the particular circumstances based on the particular criteria required (depending, for example, on the G+C content, type of target nucleic acid, source of nucleic acid, size of hybridization probe, *etc.*). Following washing of the hybridized surface so as to remove nonspecifically bound probe molecules, specific hybridization is detected, or even quantitated, by means of the label.

2.10 BIOLOGICAL FUNCTIONAL EQUIVALENTS

Modification and changes may be made in the structure of the peptides of the present invention and DNA segments which encode them and still obtain a functional molecule that encodes a protein or peptide with desirable characteristics. The following is a discussion based upon changing the amino acids of a protein to create an equivalent, or even an improved, second-generation molecule. In particular embodiments of the invention, mutated crystal proteins are contemplated to be useful for increasing the insecticidal activity of the protein, and consequently increasing the insecticidal activity and/or expression of the recombinant transgene in a plant cell. The amino acid changes may be achieved by changing the codons of the DNA sequence, according to the codons given in Table 3.

TABLE 3

Amino Acid			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites on substrate

5

molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated by the inventors
5 that various changes may be made in the peptide sequences of the disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic
10 function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporate herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like.

Each amino acid has been assigned a hydropathic index on the basis of their
15 hydrophobicity and charge characteristics (Kyte and Doolittle, 1982), these are: isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (-0.4); threonine (-0.7); serine (-0.8); tryptophan (-0.9); tyrosine (-1.3); proline (-1.6); histidine (-3.2); glutamate (-3.5);
20 glutamine (-3.5); aspartate (-3.5); asparagine (-3.5); lysine (-3.9); and arginine (-4.5).

It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.*, still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are
25 within ± 2 is preferred, those which are within ± 1 are particularly preferred, and those within ± 0.5 are even more particularly preferred.

It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 4,554,101, incorporated herein by reference, states that the greatest local average hydrophilicity of a protein, as

governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0 \pm 1);
5 glutamate (+3.0 \pm 1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0);
threonine (-0.4); proline (-0.5 \pm 1); alanine (-0.5); histidine (-0.5); cysteine (-1.0);
methionine (-1.3); valine (-1.5); leucine (-1.8); isoleucine (-1.8); tyrosine (-2.3);
phenylalanine (-2.5); tryptophan (-3.4).

It is understood that an amino acid can be substituted for another having a similar
10 hydrophilicity value and still obtain a biologically equivalent, and in particular, an
immunologically equivalent protein. In such changes, the substitution of amino acids
whose hydrophilicity values are within ± 2 is preferred, those which are within ± 1 are
particularly preferred, and those within ± 0.5 are even more particularly preferred.

As outlined above, amino acid substitutions are generally therefore based on the
15 relative similarity of the amino acid side-chain substituents, for example, their
hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions which
take various of the foregoing characteristics into consideration are well known to those of
skill in the art and include: arginine and lysine; glutamate and aspartate; serine and
threonine; glutamine and asparagine; and valine, leucine and isoleucine.

20

2.11 SITE-SPECIFIC MUTAGENESIS

Site-specific mutagenesis is a technique useful in the preparation of individual
peptides, or biologically functional equivalent proteins or peptides, through specific
mutagenesis of the underlying DNA. The technique further provides a ready ability to
25 prepare and test sequence variants, for example, incorporating one or more of the
foregoing considerations, by introducing one or more nucleotide sequence changes into
the DNA. Site-specific mutagenesis allows the production of mutants through the use of
specific oligonucleotide sequences which encode the DNA sequence of the desired
mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer
30 sequence of sufficient size and sequence complexity to form a stable duplex on both sides

of the deletion junction being traversed. Typically, a primer of about 17 to 25 nucleotides in length is preferred, with about 5 to 10 residues on both sides of the junction of the sequence being altered.

5 In general, the technique of site-specific mutagenesis is well known in the art, as exemplified by various publications. As will be appreciated, the technique typically employs a phage vector which exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially available and their use is generally well known to those skilled in the art. Double stranded plasmids are also routinely employed
10 in site directed mutagenesis which eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double stranded vector which includes within its sequence a DNA sequence which encodes the desired
15 peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second
20 strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis is provided as a means of producing potentially
25 useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence variants.

2.12 CRYSTAL PROTEIN COMPOSITIONS AS INSECTICIDES AND METHODS OF USE

The inventors contemplate that the chimeric crystal protein compositions disclosed herein will find particular utility as insecticides for topical and/or systemic application to field crops, grasses, fruits and vegetables, and ornamental plants. In a preferred embodiment, the bioinsecticide composition comprises an oil flowable suspension of bacterial cells which expresses a novel crystal protein disclosed herein. Preferably the cells are *B. thuringiensis* cells, however, any such bacterial host cell expressing the novel nucleic acid segments disclosed herein and producing a crystal protein is contemplated to be useful, such as *B. megaterium*, *B. subtilis*, *E. coli*, or *Pseudomonas* spp.

In another important embodiment, the bioinsecticide composition comprises a water dispersible granule. This granule comprises bacterial cells which expresses a novel crystal protein disclosed herein. Preferred bacterial cells are *B. thuringiensis* cells, however, bacteria such as *B. megaterium*, *B. subtilis*, *E. coli*, or *Pseudomonas* spp. cells transformed with a DNA segment disclosed herein and expressing the crystal protein are also contemplated to be useful.

In a third important embodiment, the bioinsecticide composition comprises a wettable powder, dust, pellet, or colloidal concentrate. This powder comprises bacterial cells which expresses a novel crystal protein disclosed herein. Preferred bacterial cells are *B. thuringiensis* cells, however, bacteria such as *B. megaterium*, *B. subtilis*, *E. coli*, or *Pseudomonas* spp. cells transformed with a DNA segment disclosed herein and expressing the crystal protein are also contemplated to be useful. Such dry forms of the insecticidal compositions may be formulated to dissolve immediately upon wetting, or alternatively, dissolve in a controlled-release, sustained-release, or other time-dependent manner.

In a fourth important embodiment, the bioinsecticide composition comprises an aqueous suspension of bacterial cells such as those described above which express the crystal protein. Such aqueous suspensions may be provided as a concentrated stock solution which is diluted prior to application, or alternatively, as a diluted solution ready-to-apply.

For these methods involving application of bacterial cells, the cellular host containing the crystal protein gene(s) may be grown in any convenient nutrient medium, where the DNA construct provides a selective advantage, providing for a selective medium so that substantially all or all of the cells retain the *B. thuringiensis* gene. These cells may then be harvested in accordance with conventional ways. Alternatively, the cells can be treated prior to harvesting.

When the insecticidal compositions comprise intact *B. thuringiensis* cells expressing the protein of interest, such bacteria may be formulated in a variety of ways. They may be employed as wettable powders, granules or dusts, by mixing with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, and the like) or botanical materials (powdered corncobs, rice hulls, walnut shells, and the like). The formulations may include spreader-sticker adjuvants, stabilizing agents, other pesticidal additives, or surfactants. Liquid formulations may be aqueous-based or non-aqueous and employed as foams, suspensions, emulsifiable concentrates, or the like. The ingredients may include rheological agents, surfactants, emulsifiers, dispersants, or polymers.

Alternatively, the novel chimeric Cry proteins may be prepared by recombinant bacterial expression systems *in vitro* and isolated for subsequent field application. Such protein may be either in crude cell lysates, suspensions, colloids, etc., or alternatively may be purified, refined, buffered, and/or further processed, before formulating in an active biocidal formulation. Likewise, under certain circumstances, it may be desirable to isolate crystals and/or spores from bacterial cultures expressing the crystal protein and apply solutions, suspensions, or colloidal preparations of such crystals and/or spores as the active bioinsecticidal composition.

Regardless of the method of application, the amount of the active component(s) are applied at an insecticidally-effective amount, which will vary depending on such factors as, for example, the specific coleopteran insects to be controlled, the specific plant or crop to be treated, the environmental conditions, and the method, rate, and quantity of application of the insecticidally-active composition.

The insecticide compositions described may be made by formulating either the bacterial cell, crystal and/or spore suspension, or isolated protein component with the desired agriculturally-acceptable carrier. The compositions may be formulated prior to administration in an appropriate means such as lyophilized, freeze-dried, dessicated, or in an aqueous carrier, medium or suitable diluent, such as saline or other buffer. The formulated compositions may be in the form of a dust or granular material, or a suspension in oil (vegetable or mineral), or water or oil/water emulsions, or as a wettable powder, or in combination with any other carrier material suitable for agricultural application. Suitable agricultural carriers can be solid or liquid and are well known in the art. The term "agriculturally-acceptable carrier" covers all adjuvants, *e.g.*, inert components, dispersants, surfactants, tackifiers, binders, *etc.* that are ordinarily used in insecticide formulation technology; these are well known to those skilled in insecticide formulation. The formulations may be mixed with one or more solid or liquid adjuvants and prepared by various means, *e.g.*, by homogeneously mixing, blending and/or grinding the insecticidal composition with suitable adjuvants using conventional formulation techniques.

The insecticidal compositions of this invention are applied to the environment of the target coleopteran insect, typically onto the foliage of the plant or crop to be protected, by conventional methods, preferably by spraying. The strength and duration of insecticidal application will be set with regard to conditions specific to the particular pest(s), crop(s) to be treated and particular environmental conditions. The proportional ratio of active ingredient to carrier will naturally depend on the chemical nature, solubility, and stability of the insecticidal composition, as well as the particular formulation contemplated.

Other application techniques, *e.g.*, dusting, sprinkling, soaking, soil injection, seed coating, seedling coating, spraying, aerating, misting, atomizing, and the like, are also feasible and may be required under certain circumstances such as *e.g.*, insects that cause root or stalk infestation, or for application to delicate vegetation or ornamental plants. These application procedures are also well-known to those of skill in the art.

The insecticidal composition of the invention may be employed in the method of the invention singly or in combination with other compounds, including and not limited to other pesticides. The method of the invention may also be used in conjunction with other treatments such as surfactants, detergents, polymers or time-release formulations.

5 The insecticidal compositions of the present invention may be formulated for either systemic or topical use.

The concentration of insecticidal composition which is used for environmental, systemic, or foliar application will vary widely depending upon the nature of the particular formulation, means of application, environmental conditions, and degree of
10 biocidal activity. Typically, the bioinsecticidal composition will be present in the applied formulation at a concentration of at least about 0.5% by weight and may be up to and including about 99% by weight. Dry formulations of the compositions may be from about 0.5% to about 99% or more by weight of the composition, while liquid formulations may generally comprise from about 0.5% to about 99% or more of the
15 active ingredient by weight. Formulations which comprise intact bacterial cells will generally contain from about 10^4 to about 10^{12} cells/mg.

The insecticidal formulation may be administered to a particular plant or target area in one or more applications as needed, with a typical field application rate per hectare ranging on the order of from about 50 g to about 500 g of active ingredient, or of
20 from about 500 g to about 1000 g, or of from about 1000 g to about 5000 g or more of active ingredient.

2.13 ANTIBODY COMPOSITIONS AND METHODS FOR PRODUCING

In particular embodiments, the inventors contemplate the use of antibodies, either
25 monoclonal or polyclonal which bind to the crystal proteins disclosed herein. Means for preparing and characterizing antibodies are well known in the art (See, *e.g.*, Harlow and Lane, 1988; incorporated herein by reference). The methods for generating monoclonal antibodies (mAbs) generally begin along the same lines as those for preparing polyclonal antibodies. Briefly, a polyclonal antibody is prepared by immunizing an animal with an
30 immunogenic composition in accordance with the present invention and collecting

antisera from that immunized animal. A wide range of animal species can be used for the production of antisera. Typically the animal used for production of anti-antisera is a rabbit, a mouse, a rat, a hamster, a guinea pig or a goat. Because of the relatively large blood volume of rabbits, a rabbit is a preferred choice for production of polyclonal antibodies.

As is well known in the art, a given composition may vary in its immunogenicity. It is often necessary therefore to boost the host immune system, as may be achieved by coupling a peptide or polypeptide immunogen to a carrier. Exemplary and preferred carriers are keyhole limpet hemocyanin (KLH) and bovine serum albumin (BSA). Other albumins such as ovalbumin, mouse serum albumin or rabbit serum albumin can also be used as carriers. Means for conjugating a polypeptide to a carrier protein are well known in the art and include glutaraldehyde, *m*-maleimidobencoyl-*N*-hydroxysuccinimide ester, carbodiimide and bis-biazotized benzidine.

As is also well known in the art, the immunogenicity of a particular immunogen composition can be enhanced by the use of non-specific stimulators of the immune response, known as adjuvants. Exemplary and preferred adjuvants include complete Freund's adjuvant (a non-specific stimulator of the immune response containing killed *Mycobacterium tuberculosis*), incomplete Freund's adjuvants and aluminum hydroxide adjuvant.

The amount of immunogen composition used in the production of polyclonal antibodies varies upon the nature of the immunogen as well as the animal used for immunization. A variety of routes can be used to administer the immunogen (subcutaneous, intramuscular, intradermal, intravenous and intraperitoneal). The production of polyclonal antibodies may be monitored by sampling blood of the immunized animal at various points following immunization. A second, booster, injection may also be given. The process of boosting and titering is repeated until a suitable titer is achieved. When a desired level of immunogenicity is obtained, the immunized animal can be bled and the serum isolated and stored, and/or the animal can be used to generate mAbs.

mAbs may be readily prepared through use of well-known techniques, such as those exemplified in U. S. Patent 4,196,265 (specifically incorporated herein by reference). Typically, this technique involves immunizing a suitable animal with a selected immunogen composition, e.g., a purified or partially purified crystal protein, polypeptide or peptide. The immunizing composition is administered in a manner effective to stimulate antibody producing cells. Rodents such as mice and rats are preferred animals, however, the use of rabbit, sheep frog cells is also possible. The use of rats may provide certain advantages (Goding, 1986, pp. 60-61), but mice are preferred, with the BALB/c mouse being most preferred as this is most routinely used and generally gives a higher percentage of stable fusions.

Following immunization, somatic cells with the potential for producing antibodies, specifically B lymphocytes (B cells), are selected for use in the mAb generating protocol. These cells may be obtained from biopsied spleens, tonsils or lymph nodes, or from a peripheral blood sample. Spleen cells and peripheral blood cells are preferred, the former because they are a rich source of antibody-producing cells that are in the dividing plasmablast stage, and the latter because peripheral blood is easily accessible. Often, a panel of animals will have been immunized and the spleen of animal with the highest antibody titer will be removed and the spleen lymphocytes obtained by homogenizing the spleen with a syringe. Typically, a spleen from an immunized mouse contains approximately 5×10^7 to 2×10^8 lymphocytes.

The antibody-producing B lymphocytes from the immunized animal are then fused with cells of an immortal myeloma cell, generally one of the same species as the animal that was immunized. Myeloma cell lines suited for use in hybridoma-producing fusion procedures preferably are non-antibody-producing, have high fusion efficiency, and enzyme deficiencies that render them incapable of growing in certain selective media which support the growth of only the desired fused cells (hybridomas).

Any one of a number of myeloma cells may be used, as are known to those of skill in the art (Goding, pp. 65-66, 1986; Campbell, pp. 75-83, 1984). For example, where the immunized animal is a mouse, one may use P3-X63/Ag8, X63-Ag8.653, NS1/1.Ag 4 1, Sp210-Ag14, FO, NSO/U, MPC-11, MPC11-X45-GTG 1.7 and

S194/5XX0 Bul; for rats, one may use R210.RCY3, Y3-Ag 1.2.3, IR983F and 4B210; and U-266, GM1500-GRG2, LICR-LON-HMy2 and UC729-6 are all useful in connection with human cell fusions.

5 One preferred murine myeloma cell is the NS-1 myeloma cell line (also termed P3-NS-1-Ag4-1), which is readily available from the NIGMS Human Genetic Mutant Cell Repository by requesting cell line repository number GM3573. Another mouse myeloma cell line that may be used is the 8-azaguanine-resistant mouse murine myeloma SP2/0 non-producer cell line.

10 Methods for generating hybrids of antibody-producing spleen or lymph node cells and myeloma cells usually comprise mixing somatic cells with myeloma cells in a 2:1 ratio, though the ratio may vary from about 20:1 to about 1:1, respectively, in the presence of an agent or agents (chemical or electrical) that promote the fusion of cell membranes. Fusion methods using Sendai virus have been described (Kohler and Milstein, 1975; 1976), and those using polyethylene glycol (PEG), such as 37% (vol./vol.) PEG, (Geftter *et al.*, 1977). The use of electrically induced fusion methods is
15 also appropriate (Goding, 1986, pp. 71-74).

Fusion procedures usually produce viable hybrids at low frequencies, about 1×10^{-6} to 1×10^{-8} . However, this does not pose a problem, as the viable, fused hybrids are differentiated from the parental, unfused cells (particularly the unfused myeloma cells
20 that would normally continue to divide indefinitely) by culturing in a selective medium. The selective medium is generally one that contains an agent that blocks the *de novo* synthesis of nucleotides in the tissue culture media. Exemplary and preferred agents are aminopterin, methotrexate, and azaserine. Aminopterin and methotrexate block *de novo* synthesis of both purines and pyrimidines, whereas azaserine blocks only purine
25 synthesis. Where aminopterin or methotrexate is used, the media is supplemented with hypoxanthine and thymidine as a source of nucleotides (HAT medium). Where azaserine is used, the media is supplemented with hypoxanthine.

The preferred selection medium is HAT. Only cells capable of operating nucleotide salvage pathways are able to survive in HAT medium. The myeloma cells are
30 defective in key enzymes of the salvage pathway, *e.g.*, hypoxanthine phosphoribosyl

transferase (HPRT), and they cannot survive. The B-cells can operate this pathway, but they have a limited life span in culture and generally die within about two weeks. Therefore, the only cells that can survive in the selective media are those hybrids formed from myeloma and B-cells.

5 This culturing provides a population of hybridomas from which specific hybridomas are selected. Typically, selection of hybridomas is performed by culturing the cells by single-clone dilution in microtiter plates, followed by testing the individual clonal supernatants (after about two to three weeks) for the desired reactivity. The assay should be sensitive, simple and rapid, such as radioimmunoassays, enzyme
10 immunoassays, cytotoxicity assays, plaque assays, dot immunobinding assays, and the like.

 The selected hybridomas would then be serially diluted and cloned into individual antibody-producing cell lines, which clones can then be propagated indefinitely to provide mAbs. The cell lines may be exploited for mAb production in two basic ways.
15 A sample of the hybridoma can be injected (often into the peritoneal cavity) into a histocompatible animal of the type that was used to provide the somatic and myeloma cells for the original fusion. The injected animal develops tumors secreting the specific monoclonal antibody produced by the fused cell hybrid. The body fluids of the animal, such as serum or ascites fluid, can then be tapped to provide mAbs in high concentration.
20 The individual cell lines could also be cultured *in vitro*, where the mAbs are naturally secreted into the culture medium from which they can be readily obtained in high concentrations. mAbs produced by either means may be further purified, if desired, using filtration, centrifugation and various chromatographic methods such as HPLC or affinity chromatography.

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3. **BRIEF DESCRIPTION OF THE DRAWINGS**

 The following drawings form part of the present specification and are included to further demonstrate certain aspects of the present invention. The invention may be better understood by reference to one or more of these drawings in combination with the
30 detailed description of specific embodiments presented herein.

FIG. 1. The wild-type δ -endotoxins and the relevant restriction sites that were used to construct the hybrid δ -endotoxins pertinent to the invention are diagrammed in FIG. 1A. Only the DNA encoding the δ -endotoxin that is contained on the indicated plasmid (identified by the "pEG" prefix) is shown. The *B. thuringiensis* strains containing the indicated plasmids are identified by the "EG" prefix. The hybrid δ -endotoxins described in the invention are diagrammed in FIG. 1B and are aligned with the wild-type δ -endotoxins in FIG. 1A.

FIG. 2. An equal amount of each washed sporulated *B. thuringiensis* culture was analyzed by SDS-PAGE. Lane a: control Cry1Ac producing *B. thuringiensis* strain EG11070, b: EG11060, c: EG11062, d: EG11063, e: EG11065, f: EG11067, g: EG11071, h: EG11073, i: EG11074, j: EG11088, k: EG11090, and l: EG11091.

FIG. 3. Solubilized hybrid δ -endotoxins were exposed to trypsin for 0, 15, 30, 60, and 120 minutes. The resulting material was analyzed by SDS-PAGE. The amount of active δ -endotoxin fragment remaining was quantitated by scanning densitometry using a Molecular Dynamics model 300A densitometer. The percent active toxin remaining was plotted versus time. Wild-type Cry1Ac δ -endotoxin (open box) served as the control.

FIG. 4. Schematic diagrams of the wild-type toxins and the relevant restriction sites that were used to construct the hybrid δ -endotoxin encoded by pEG381 and expressed in EG11768. Only the DNA encoding the δ -endotoxin that is contained on the indicated plasmid (identified by the "pEG" prefix) is shown.

4. BRIEF DESCRIPTION OF THE SEQUENCES

SEQ ID NO:1 is oligonucleotide primer A.

SEQ ID NO:2 is oligonucleotide primer B.

SEQ ID NO:3 is oligonucleotide primer C.

SEQ ID NO:4 is oligonucleotide primer D.

SEQ ID NO:5 is oligonucleotide primer E.

SEQ ID NO:6 is oligonucleotide primer F.

SEQ ID NO:7 is oligonucleotide primer G.

SEQ ID NO:8 is oligonucleotide primer H.

5 **SEQ ID NO:9** is the nucleotide and deduced amino acid sequences of the
EG11063 hybrid δ -endotoxin.

SEQ ID NO:10 denotes in the three-letter abbreviation form, the amino acid
sequence for the hybrid δ -endotoxin specified in **SEQ ID NO:9**.

SEQ ID NO:11 is the nucleotide and deduced amino acid sequences of the
EG11074 hybrid δ -endotoxin.

10 **SEQ ID NO:12** denotes in the three-letter abbreviation form, the amino acid
sequence for the hybrid δ -endotoxin specified in **SEQ ID NO:11**.

SEQ ID NO:13 is the nucleotide and deduced amino acid sequences of the
EG11735 hybrid δ -endotoxin.

15 **SEQ ID NO:14** denotes in the three-letter abbreviation form, the amino acid
sequence for the hybrid δ -endotoxin specified in **SEQ ID NO:13**.

SEQ ID NO:15 is the 5' exchange site for pEG1065, pEG1070, and pEG1074.

SEQ ID NO:16 is the 5' exchange site for pEG1067, pEG1072, and pEG1076.

SEQ ID NO:17 is the 5' exchange site for pEG1068, pEG1077, and pEG365.

SEQ ID NO:18 is the 5' exchange site for pEG1088 and pEG1092.

20 **SEQ ID NO:19** is the 5' exchange site for pEG1089 and the 3' exchange site for
pEG1070 and pEG1072.

SEQ ID NO:20 is the 5' exchange site for pEG1091.

SEQ ID NO:21 is the 3' exchange site for pEG1065, pEG1067, pEG1068,
pEG1093, pEG378, and pEG 365.

25 **SEQ ID NO:22** is the 3' exchange site for pEG1088.

SEQ ID NO:23 is oligonucleotide Primer I.

SEQ ID NO:24 is oligonucleotide Primer J.

SEQ ID NO:25 is the nucleic acid sequence and deduced amino acid sequence of
the hybrid crystal protein-encoding gene of EG11092.

SEQ ID NO:26 is the three-letter abbreviation form of the amino acid sequence of the hybrid crystal protein produced by strain EG11092 encoded by SEQ ID NO:25.

SEQ ID NO:27 is the nucleic acid sequence and the deduced amino acid sequence of the hybrid crystal protein-encoding gene of EG11751.

5 SEQ ID NO:28 is the three-letter abbreviation form of the amino acid sequence of the hybrid crystal protein produced by strain EG11751 encoded by SEQ ID NO:27.

SEQ ID NO:29 is the nucleic acid sequence and the deduced amino acid sequence of the hybrid crystal protein-encoding gene of EG11091.

10 SEQ ID NO:30 is the three-letter abbreviation form of the amino acid sequence of the hybrid crystal protein produced by strain EG11091 encoded by SEQ ID NO:29.

SEQ ID NO:31 is oligonucleotide primer K.

SEQ ID NO:32 is the 5' exchange site for pEG378 and pEG381.

SEQ ID NO:33 is the nucleic acid sequence and the deduced amino acid sequence of the hybrid crystal protein-encoding gene of EG11768.

15 SEQ ID NO:34 denotes in the three-letter abbreviation form, the amino acid sequence of the hybrid crystal protein produced by strain EG11768 encoded by SEQ ID NO:33.

SEQ ID NO:35 is the 3' exchange site for pEG1074, pEG1076, pEG1077 and pEG381.

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5. DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

5.1 METHODS FOR CULTURING *B. THURINGIENSIS* TO PRODUCE CRY PROTEINS

The *B. thuringiensis* strains described herein may be cultured using standard known media and fermentation techniques. Upon completion of the fermentation cycle, the bacteria may be harvested by first separating the *B. thuringiensis* spores and crystals from the fermentation broth by means well known in the art. The recovered *B. thuringiensis* spores and crystals can be formulated into a wettable powder, a liquid concentrate, granules or other formulations by the addition of surfactants, dispersants, inert carriers and other components to facilitate handling and application for particular target pests. The formulation and application procedures are all well known in the art and

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are used with commercial strains of *B. thuringiensis* (HD-1) active against Lepidoptera, e.g., caterpillars.

5.2 RECOMBINANT HOST CELLS FOR EXPRESSION OF CRY GENES

5 The nucleotide sequences of the subject invention can be introduced into a wide variety of microbial hosts. Expression of the toxin gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. With suitable hosts, e.g., *Pseudomonas*, the microbes can be applied to the sites of lepidopteran insects where they will proliferate and be ingested by the insects. The results is a control of the unwanted insects. Alternatively, the microbe hosting the toxin gene can be treated under conditions that prolong the activity of the toxin produced in the cell. The treated cell then can be applied to the environment of target pest(s). The resulting product retains the toxicity of the *B. thuringiensis* toxin.

15 Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility or toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi. Illustrative prokaryotes, both Gram-negative and Gram-positive, include *Enterobacteriaceae*, such as *Escherichia*, *Erwinia*, *Shigella*, *Salmonella*, and *Proteus*; *Bacillaceae*; *Rhizobiceae*, such as *Rhizobium*; *Spirillaceae*, such as photobacterium, *Zymomonas*, *Serratia*, *Aeromonas*, *Vibrio*, *Desulfovibrio*, *Spirillum*; *Lactobacillaceae*; *Pseudomonadaceae*, such as *Pseudomonas* and *Acetobacter*; *Azotobacteraceae*, *Actinomycetales*, and *Nitrobacteraceae*. Among eukaryotes are fungi, such as *Phycomycetes* and *Ascomycetes*, which includes yeast, such as *Saccharomyces* and *Schizosaccharomyces*; and *Basidiomycetes* yeast, such as *Rhodotorula*, *Aureobasidium*, *Sporobolomyces*, and the like.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the *B. thuringiensis* gene into the host, availability of expression systems, efficiency of expression, stability of the pesticide in the host, and the presence of auxiliary genetic capabilities. Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; leaf affinity; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

Host organisms of particular interest include yeast, such as *Rhodotorula sp.*, *Aureobasidium sp.*, *Saccharomyces sp.*, and *Sporobolomyces sp.*; phylloplane organisms such as *Pseudomonas sp.*, *Erwinia sp.* and *Flavobacterium sp.*; or such other organisms as *Escherichia*, *Lactobacillus sp.*, *Bacillus sp.*, *Streptomyces sp.*, and the like. Specific organisms include *Pseudomonas aeruginosa*, *P. fluorescens*, *Saccharomyces cerevisiae*, *B. thuringiensis*, *B. subtilis*, *E. coli*, *Streptomyces lividans* and the like.

Treatment of the microbial cell, e.g., a microbe containing the *B. thuringiensis* toxin gene, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in protecting the toxin. Examples of chemical reagents are halogenating agents, particularly halogens of atomic no. 17-80. More particularly, iodine can be used under mild conditions and for sufficient time to achieve the desired results. Other suitable techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride and cetylpyridinium chloride; alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Lugol's iodine, Bouin's fixative, and Helly's fixatives, (see e.g., Humason, 1967); or a combination of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced in the cell when the cell is administered to a suitable host. Examples of physical means are short wavelength radiation such as γ -radiation and X-radiation, freezing, UV irradiation, lyophilization, and the like. The cells employed will usually be intact and be substantially in the

proliferative form when treated, rather than in a spore form, although in some instances spores may be employed.

Where the *B. thuringiensis* toxin gene is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is essential that certain host microbes be used. Microorganism hosts are selected which are known to occupy the "phytosphere" (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots) of a wide variety of important crops. These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera *Bacillus*, *Pseudomonas*, *Erwinia*, *Serratia*, *Klebsiella*, *Xanthomonas*, *Streptomyces*, *Rhizobium*, *Rhodopseudomonas*, *Methylophilus*, *Agrobacterium*, *Acetobacter*, *Lactobacillus*, *Arthrobacter*, *Azotobacter*, *Leuconostoc*, and *Alcaligenes*; fungi, particularly yeast, e.g., genera *Saccharomyces*, *Cryptococcus*, *Kluyveromyces*, *Sporobolomyces*, *Rhodotorula*, and *Aureobasidium*. Of particular interest are such phytosphere bacterial species as *Pseudomonas syringae*, *Pseudomonas fluorescens*, *Serratia marcescens*, *Acetobacter xylinum*, *Agrobacterium tumefaciens*, *Rhodobacter sphaeroides*, *Xanthomonas campestris*, *Rhizobium melioli*, *Alcaligenes eutrophus*, and *Azotobacter vinlandii*; and phytosphere yeast species such as *Rhodotorula rubra*, *R. glutinis*, *R. marina*, *R. aurantiaca*, *Cryptococcus albidus*, *C. diffluens*, *C. laurentii*, *Saccharomyces rosei*, *S. pretoriensis*, *S. cerevisiae*, *Sporobolomyces roseus*, *S. odoratus*, *Kluyveromyces veronae*, and *Aureobasidium pollulans*.

5.3 DEFINITIONS

The following words and phrases have the meanings set forth below.

Broad-Spectrum: refers to a wide range of insect species.

Broad-Spectrum Insecticidal Activity: toxicity towards a wide range of insect species.

5 **Expression:** The combination of intracellular processes, including transcription and translation undergone by a coding DNA molecule such as a structural gene to produce a polypeptide.

Insecticidal Activity: toxicity towards insects.

Insecticidal Specificity: the toxicity exhibited by a crystal protein towards multiple insect species.

10 **Intraorder Specificity:** the toxicity of a particular crystal protein towards insect species within an Order of insects (*e.g.*, Order Lepidoptera).

Interorder Specificity: the toxicity of a particular crystal protein towards insect species of different Orders (*e.g.*, Orders Lepidoptera and Diptera).

15 **LC₅₀:** the lethal concentration of crystal protein that causes 50% mortality of the insects treated.

LC₉₅: the lethal concentration of crystal protein that causes 95% mortality of the insects treated.

20 **Promoter:** A recognition site on a DNA sequence or group of DNA sequences that provide an expression control element for a structural gene and to which RNA polymerase specifically binds and initiates RNA synthesis (transcription) of that gene.

Regeneration: The process of growing a plant from a plant cell (*e.g.*, plant protoplast or explant).

Structural Gene: A gene that is expressed to produce a polypeptide.

25 **Transformation:** A process of introducing an exogenous DNA sequence (*e.g.*, a vector, a recombinant DNA molecule) into a cell or protoplast in which that exogenous DNA is incorporated into a chromosome or is capable of autonomous replication.

Transformed Cell: A cell whose DNA has been altered by the introduction of an exogenous DNA molecule into that cell.

30 **Transgene:** An exogenous gene which when introduced into the genome of a host cell through a process such as transformation, electroporation, particle

bombardment, and the like, is expressed by the host cell and integrated into the cells genome such that the trait or traits produced by the expression of the transgene is inherited by the progeny of the transformed cell.

5 **Transgenic Cell:** Any cell derived or regenerated from a transformed cell or derived from a transgenic cell. Exemplary transgenic cells include plant calli derived from a transformed plant cell and particular cells such as leaf, root, stem, *e.g.*, somatic cells, or reproductive (germ) cells obtained from a transgenic plant.

10 **Transgenic Plant:** A plant or progeny thereof derived from a transformed plant cell or protoplast, wherein the plant DNA contains an introduced exogenous DNA molecule not originally present in a native, non-transgenic plant of the same strain. The terms "transgenic plant" and "transformed plant" have sometimes been used in the art as synonymous terms to define a plant whose DNA contains an exogenous DNA molecule. However, it is thought more scientifically correct to refer to a regenerated plant or callus obtained from a transformed plant cell or protoplast as being a transgenic plant, and that
15 usage will be followed herein.

Vector: A DNA molecule capable of replication in a host cell and/or to which another DNA segment can be operatively linked so as to bring about replication of the attached segment. A plasmid is an exemplary vector.

20 5.4 PROBES AND PRIMERS

 In another aspect, DNA sequence information provided by the invention allows for the preparation of relatively short DNA (or RNA) sequences having the ability to specifically hybridize to gene sequences of the selected polynucleotides disclosed herein. In these aspects, nucleic acid probes of an appropriate length are prepared based on a
25 consideration of a selected crystal protein gene sequence, *e.g.*, a sequence such as that shown in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, or SEQ ID NO:33. The ability of such nucleic acid probes to specifically hybridize to a crystal protein-encoding gene sequence lends them particular utility in a variety of embodiments. Most importantly, the probes may be used

in a variety of assays for detecting the presence of complementary sequences in a given sample.

In certain embodiments, it is advantageous to use oligonucleotide primers. The sequence of such primers is designed using a polynucleotide of the present invention for use in detecting, amplifying or mutating a defined segment of a crystal protein gene from *B. thuringiensis* using PCR™ technology. Segments of related crystal protein genes from other species may also be amplified by PCR™ using such primers.

To provide certain of the advantages in accordance with the present invention, a preferred nucleic acid sequence employed for hybridization studies or assays includes sequences that are complementary to at least a 14 to 30 or so long nucleotide stretch of a crystal protein-encoding sequence, such as that shown in SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, or SEQ ID NO:33. A size of at least 14 nucleotides in length helps to ensure that the fragment will be of sufficient length to form a duplex molecule that is both stable and selective. Molecules having complementary sequences over stretches greater than 14 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 14 to 20 nucleotides, or even longer where desired. Such fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patents 4,683,195, and 4,683,202 (each specifically incorporated herein by reference), or by excising selected DNA fragments from recombinant plasmids containing appropriate inserts and suitable restriction sites.

5.5 EXPRESSION VECTORS

The present invention contemplates an expression vector comprising a polynucleotide of the present invention. Thus, in one embodiment an expression vector is an isolated and purified DNA molecule comprising a promoter operatively linked to an coding region that encodes a polypeptide of the present invention, which coding region is

operatively linked to a transcription-terminating region, whereby the promoter drives the transcription of the coding region.

As used herein, the term "operatively linked" means that a promoter is connected to an coding region in such a way that the transcription of that coding region is controlled and regulated by that promoter. Means for operatively linking a promoter to a coding region are well known in the art.

Promoters that function in bacteria are well known in the art. Exemplary and preferred promoters for the *Bacillus* crystal proteins include the *sigA*, *sigE*, and *sigK* gene promoters. Alternatively, the native, mutagenized, or recombinant crystal protein-encoding gene promoters themselves can be used.

Where an expression vector of the present invention is to be used to transform a plant, a promoter is selected that has the ability to drive expression in plants. Promoters that function in plants are also well known in the art. Useful in expressing the polypeptide in plants are promoters that are inducible, viral, synthetic, constitutive as described (Poszkowski *et al.*, 1989; Odell *et al.*, 1985), and temporally regulated, spatially regulated, and spatio-temporally regulated (Chau *et al.*, 1989).

A promoter is also selected for its ability to direct the transformed plant cell's or transgenic plant's transcriptional activity to the coding region. Structural genes can be driven by a variety of promoters in plant tissues. Promoters can be near-constitutive, such as the CaMV 35S promoter, or tissue-specific or developmentally specific promoters affecting dicots or monocots.

Where the promoter is a near-constitutive promoter such as CaMV 35S, increases in polypeptide expression are found in a variety of transformed plant tissues (*e.g.*, callus, leaf, seed and root). Alternatively, the effects of transformation can be directed to specific plant tissues by using plant integrating vectors containing a tissue-specific promoter.

An exemplary tissue-specific promoter is the lectin promoter, which is specific for seed tissue. The Lectin protein in soybean seeds is encoded by a single gene (*Lel*) that is only expressed during seed maturation and accounts for about 2 to about 5% of total seed mRNA. The lectin gene and seed-specific promoter have been fully characterized and

used to direct seed specific expression in transgenic tobacco plants (Vodkin *et al.*, 1983; Lindstrom *et al.*, 1990.)

5 An expression vector containing a coding region that encodes a polypeptide of interest is engineered to be under control of the lectin promoter and that vector is introduced into plants using, for example, a protoplast transformation method (Dhir *et al.*, 1991). The expression of the polypeptide is directed specifically to the seeds of the transgenic plant.

10 A transgenic plant of the present invention produced from a plant cell transformed with a tissue specific promoter can be crossed with a second transgenic plant developed from a plant cell transformed with a different tissue specific promoter to produce a hybrid transgenic plant that shows the effects of transformation in more than one specific tissue.

15 Exemplary tissue-specific promoters are corn sucrose synthetase 1 (Yang *et al.*, 1990), corn alcohol dehydrogenase 1 (Vogel *et al.*, 1989), corn light harvesting complex (Simpson, 1986), corn heat shock protein (Odell *et al.*, 1985), pea small subunit RuBP carboxylase (Poulsen *et al.*, 1986; Cashmore *et al.*, 1983), Ti plasmid mannopirite synthase (Langridge *et al.*, 1989), Ti plasmid nopaline synthase (Langridge *et al.*, 1989), petunia chalcone isomerase (Van Tunen *et al.*, 1988), bean glycine rich protein 1 (Keller *et al.*, 1989), CaMV 35s transcript (Odell *et al.*, 1985) and Potato patatin (Wenzler *et al.*, 1989). Preferred promoters are the cauliflower mosaic virus (CaMV 35S) promoter and
20 the S-E9 small subunit RuBP carboxylase promoter.

The choice of which expression vector and ultimately to which promoter a polypeptide coding region is operatively linked depends directly on the functional properties desired, *e.g.*, the location and timing of protein expression, and the host cell to be transformed. These are well known limitations inherent in the art of constructing
25 recombinant DNA molecules. However, a vector useful in practicing the present invention is capable of directing the expression of the polypeptide coding region to which it is operatively linked.

30 Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of *Agrobacterium tumefaciens* described (Rogers *et al.*, 1987). However, several other plant

integrating vector systems are known to function in plants including pCaMVCN transfer control vector described (Fromm *et al.*, 1985). pCaMVCN (available from Pharmacia, Piscataway, NJ) includes the cauliflower mosaic virus CaMV 35S promoter.

In preferred embodiments, the vector used to express the polypeptide includes a selection marker that is effective in a plant cell, preferably a drug resistance selection marker. One preferred drug resistance marker is the gene whose expression results in kanamycin resistance; *i.e.*, the chimeric gene containing the nopaline synthase promoter, Tn5 neomycin phosphotransferase II (*nptII*) and nopaline synthase 3' non-translated region described (Rogers *et al.*, 1988).

RNA polymerase transcribes a coding DNA sequence through a site where polyadenylation occurs. Typically, DNA sequences located a few hundred base pairs downstream of the polyadenylation site serve to terminate transcription. Those DNA sequences are referred to herein as transcription-termination regions. Those regions are required for efficient polyadenylation of transcribed messenger RNA (mRNA).

Means for preparing expression vectors are well known in the art. Expression (transformation vectors) used to transform plants and methods of making those vectors are described in U. S. Patents 4,971,908, 4,940,835, 4,769,061 and 4,757,011 (each of which is specifically incorporated herein by reference). Those vectors can be modified to include a coding sequence in accordance with the present invention.

A variety of methods has been developed to operatively link DNA to vectors via complementary cohesive termini or blunt ends. For instance, complementary homopolymer tracts can be added to the DNA segment to be inserted and to the vector DNA. The vector and DNA segment are then joined by hydrogen bonding between the complementary homopolymeric tails to form recombinant DNA molecules.

A coding region that encodes a polypeptide having the ability to confer insecticidal activity to a cell is preferably a chimeric *B. thuringiensis* crystal protein-encoding gene. In preferred embodiments, such a polypeptide has the amino acid residue sequence of SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, or SEQ ID NO:34; or a functional equivalent of one or more of those sequences. In accordance with such embodiments, a coding region

comprising the DNA sequence of SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13, SEQ ID NO:25, SEQ ID NO:27, SEQ ID NO:29, or SEQ ID NO:33 is also preferred.

5.6 TRANSFORMED OR TRANSGENIC PLANT CELLS

5 A bacterium, a yeast cell, or a plant cell or a plant transformed with an expression vector of the present invention is also contemplated. A transgenic bacterium, yeast cell, plant cell or plant derived from such a transformed or transgenic cell is also contemplated. Means for transforming bacteria and yeast cells are well known in the art. Typically, means of transformation are similar to those well known means used to
10 transform other bacteria or yeast such as *E. coli* or *S. cerevisiae*.

Methods for DNA transformation of plant cells include *Agrobacterium*-mediated plant transformation, protoplast transformation, gene transfer into pollen, injection into reproductive organs, injection into immature embryos and particle bombardment. Each of these methods has distinct advantages and disadvantages. Thus, one particular method
15 of introducing genes into a particular plant strain may not necessarily be the most effective for another plant strain, but it is well known which methods are useful for a particular plant strain.

There are many methods for introducing transforming DNA segments into cells, but not all are suitable for delivering DNA to plant cells. Suitable methods are believed
20 to include virtually any method by which DNA can be introduced into a cell, such as infection by *A. tumefaciens* and related *Agrobacterium*, direct delivery of DNA such as, for example, by PEG-mediated transformation of protoplasts (Omirulleh *et al.*, 1993), by desiccation/inhibition-mediated DNA uptake, by electroporation, by agitation with silicon carbide fibers, by acceleration of DNA coated particles, *etc.* In certain embodiments,
25 acceleration methods are preferred and include, for example, microprojectile bombardment and the like.

Technology for introduction of DNA into cells is well-known to those of skill in the art. Four general methods for delivering a gene into cells have been described: (1) chemical methods (Graham and van der Eb, 1973); (2) physical methods such as
30 microinjection (Capecchi, 1980), electroporation (Wong and Neumann, 1982; Fromm

et al., 1985) and the gene gun (Johnston and Tang, 1994; Fynan *et al.*, 1993); (3) viral vectors (Clapp, 1993; Lu *et al.*, 1993; Eglitis and Anderson, 1988a; 1988b); and (4) receptor-mediated mechanisms (Curiel *et al.*, 1991; 1992; Wagner *et al.*, 1992).

5 5.6.1 ELECTROPORATION

The application of brief, high-voltage electric pulses to a variety of animal and plant cells leads to the formation of nanometer-sized pores in the plasma membrane. DNA is taken directly into the cell cytoplasm either through these pores or as a consequence of the redistribution of membrane components that accompanies closure of the pores. Electroporation can be extremely efficient and can be used both for transient expression of clones genes and for establishment of cell lines that carry integrated copies of the gene of interest. Electroporation, in contrast to calcium phosphate-mediated transfection and protoplast fusion, frequently gives rise to cell lines that carry one, or at most a few, integrated copies of the foreign DNA.

15 The introduction of DNA by means of electroporation, is well-known to those of skill in the art. In this method, certain cell wall-degrading enzymes, such as pectin-degrading enzymes, are employed to render the target recipient cells more susceptible to transformation by electroporation than untreated cells. Alternatively, recipient cells are made more susceptible to transformation, by mechanical wounding. To effect transformation by electroporation one may employ either friable tissues such as a suspension culture of cells, or embryogenic callus, or alternatively, one may transform immature embryos or other organized tissues directly. One would partially degrade the cell walls of the chosen cells by exposing them to pectin-degrading enzymes (pectolyases) or mechanically wounding in a controlled manner. Such cells would then be recipient to DNA transfer by electroporation, which may be carried out at this stage, and transformed cells then identified by a suitable selection or screening protocol dependent on the nature of the newly incorporated DNA.

5.6.2 MICROPROJECTILE BOMBARDMENT

A further advantageous method for delivering transforming DNA segments to plant cells is microprojectile bombardment. In this method, particles may be coated with nucleic acids and delivered into cells by a propelling force. Exemplary particles include those comprised of tungsten, gold, platinum, and the like.

An advantage of microprojectile bombardment, in addition to it being an effective means of reproducibly stably transforming monocots, is that neither the isolation of protoplasts (Cristou *et al.*, 1988) nor the susceptibility to *Agrobacterium* infection is required. An illustrative embodiment of a method for delivering DNA into maize cells by acceleration is a Biolistics Particle Delivery System, which can be used to propel particles coated with DNA or cells through a screen, such as a stainless steel or Nytex screen, onto a filter surface covered with corn cells cultured in suspension. The screen disperses the particles so that they are not delivered to the recipient cells in large aggregates. It is believed that a screen intervening between the projectile apparatus and the cells to be bombarded reduces the size of projectiles aggregate and may contribute to a higher frequency of transformation by reducing damage inflicted on the recipient cells by projectiles that are too large.

For the bombardment, cells in suspension are preferably concentrated on filters or solid culture medium. Alternatively, immature embryos or other target cells may be arranged on solid culture medium. The cells to be bombarded are positioned at an appropriate distance below the macroprojectile stopping plate. If desired, one or more screens are also positioned between the acceleration device and the cells to be bombarded. Through the use of techniques set forth herein one may obtain up to 1000 or more foci of cells transiently expressing a marker gene. The number of cells in a focus which express the exogenous gene product 48 hours post-bombardment often range from 1 to 10 and average 1 to 3.

In bombardment transformation, one may optimize the prebombardment culturing conditions and the bombardment parameters to yield the maximum numbers of stable transformants. Both the physical and biological parameters for bombardment are important in this technology. Physical factors are those that involve manipulating the

DNA/microprojectile precipitate or those that affect the flight and velocity of either the macro- or microprojectiles. Biological factors include all steps involved in manipulation of cells before and immediately after bombardment, the osmotic adjustment of target cells to help alleviate the trauma associated with bombardment, and also the nature of the transforming DNA, such as linearized DNA or intact supercoiled plasmids. It is believed that pre-bombardment manipulations are especially important for successful transformation of immature embryos.

Accordingly, it is contemplated that one may wish to adjust various of the bombardment parameters in small scale studies to fully optimize the conditions. One may particularly wish to adjust physical parameters such as gap distance, flight distance, tissue distance, and helium pressure. One may also minimize the trauma reduction factors (TRFs) by modifying conditions which influence the physiological state of the recipient cells and which may therefore influence transformation and integration efficiencies. For example, the osmotic state, tissue hydration and the subculture stage or cell cycle of the recipient cells may be adjusted for optimum transformation. The execution of other routine adjustments will be known to those of skill in the art in light of the present disclosure.

The methods of particle-mediated transformation is well-known to those of skill in the art. U. S. Patent 5,015,580 (specifically incorporated herein by reference) describes the transformation of soybeans using such a technique.

5.6.3 *AGROBACTERIUM*-MEDIATED TRANSFER

Agrobacterium-mediated transfer is a widely applicable system for introducing genes into plant cells because the DNA can be introduced into whole plant tissues, thereby bypassing the need for regeneration of an intact plant from a protoplast. The use of *Agrobacterium*-mediated plant integrating vectors to introduce DNA into plant cells is well known in the art. See, for example, the methods described (Fraley *et al.*, 1985; Rogers *et al.*, 1987). The genetic engineering of cotton plants using *Agrobacterium*-mediated transfer is described in U. S. Patent 5,004,863 (specifically incorporated herein by reference), while the transformation of lettuce plants is described in U. S. Patent

5,349,124 (specifically incorporated herein by reference). Further, the integration of the Ti-DNA is a relatively precise process resulting in few rearrangements. The region of DNA to be transferred is defined by the border sequences, and intervening DNA is usually inserted into the plant genome as described (Spielmann *et al.*, 1986; Jorgensen *et al.*, 1987).

Modern *Agrobacterium* transformation vectors are capable of replication in *E. coli* as well as *Agrobacterium*, allowing for convenient manipulations as described (Klee *et al.*, 1985). Moreover, recent technological advances in vectors for *Agrobacterium*-mediated gene transfer have improved the arrangement of genes and restriction sites in the vectors to facilitate construction of vectors capable of expressing various polypeptide coding genes. The vectors described (Rogers *et al.*, 1987), have convenient multi-linker regions flanked by a promoter and a polyadenylation site for direct expression of inserted polypeptide coding genes and are suitable for present purposes. In addition, *Agrobacterium* containing both armed and disarmed Ti genes can be used for the transformations. In those plant strains where *Agrobacterium*-mediated transformation is efficient, it is the method of choice because of the facile and defined nature of the gene transfer.

Agrobacterium-mediated transformation of leaf disks and other tissues such as cotyledons and hypocotyls appears to be limited to plants that *Agrobacterium* naturally infects. *Agrobacterium*-mediated transformation is most efficient in dicotyledonous plants. Few monocots appear to be natural hosts for *Agrobacterium*, although transgenic plants have been produced in asparagus using *Agrobacterium* vectors as described (Bytebier *et al.*, 1987). Therefore, commercially important cereal grains such as rice, corn, and wheat must usually be transformed using alternative methods. However, as mentioned above, the transformation of asparagus using *Agrobacterium* can also be achieved (see, *e.g.*, Bytebier *et al.*, 1987).

A transgenic plant formed using *Agrobacterium* transformation methods typically contains a single gene on one chromosome. Such transgenic plants can be referred to as being heterozygous for the added gene. However, inasmuch as use of the word "heterozygous" usually implies the presence of a complementary gene at the same locus

of the second chromosome of a pair of chromosomes, and there is no such gene in a plant containing one added gene as here, it is believed that a more accurate name for such a plant is an independent segregant, because the added, exogenous gene segregates independently during mitosis and meiosis.

5 More preferred is a transgenic plant that is homozygous for the added structural gene; *i.e.*, a transgenic plant that contains two added genes, one gene at the same locus on each chromosome of a chromosome pair. A homozygous transgenic plant can be obtained by sexually mating (selfing) an independent segregant transgenic plant that contains a single added gene, germinating some of the seed produced and analyzing the
10 resulting plants produced for enhanced carboxylase activity relative to a control (native, non-transgenic) or an independent segregant transgenic plant.

 It is to be understood that two different transgenic plants can also be mated to produce offspring that contain two independently segregating added, exogenous genes. Selfing of appropriate progeny can produce plants that are homozygous for both added,
15 exogenous genes that encode a polypeptide of interest. Back-crossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.

 Transformation of plant protoplasts can be achieved using methods based on calcium phosphate precipitation, polyethylene glycol treatment, electroporation, and combinations of these treatments (see, *e.g.*, Potrykus *et al.*, 1985; Lorz *et al.*, 1985;
20 Fromm *et al.*, 1985; Uchimiya *et al.*, 1986; Callis *et al.*, 1987; Marcotte *et al.*, 1988).

 Application of these systems to different plant strains depends upon the ability to regenerate that particular plant strain from protoplasts. Illustrative methods for the regeneration of cereals from protoplasts are described (see, *e.g.*, Fujimura *et al.*, 1985; Toriyama *et al.*, 1986; Yamada *et al.*, 1986; Abdullah *et al.*, 1986).

25 To transform plant strains that cannot be successfully regenerated from protoplasts, other ways to introduce DNA into intact cells or tissues can be utilized. For example, regeneration of cereals from immature embryos or explants can be effected as described (Vasil, 1988). In addition, "particle gun" or high-velocity microprojectile technology can be utilized (Vasil, 1992).

Using that latter technology, DNA is carried through the cell wall and into the cytoplasm on the surface of small metal particles as described (Klein *et al.*, 1987; Klein *et al.*, 1988; McCabe *et al.*, 1988). The metal particles penetrate through several layers of cells and thus allow the transformation of cells within tissue explants.

5

5.7 PRODUCTION OF INSECT-RESISTANT TRANSGENIC PLANTS

Thus, the amount of a gene coding for a polypeptide of interest (*i.e.*, a bacterial crystal protein or polypeptide having insecticidal activity against one or more insect species) can be increased in plant such as corn by transforming those plants using particle bombardment methods (Maddock *et al.*, 1991). By way of example, an expression vector containing a coding region for a *B. thuringiensis* crystal protein and an appropriate selectable marker is transformed into a suspension of embryonic maize (corn) cells using a particle gun to deliver the DNA coated on microprojectiles. Transgenic plants are regenerated from transformed embryonic calli that express the disclosed insecticidal crystal proteins. Particle bombardment has been used to successfully transform wheat (Vasil *et al.*, 1992).

DNA can also be introduced into plants by direct DNA transfer into pollen as described (Zhou *et al.*, 1983; Hess, 1987; Luo *et al.*, 1988). Expression of polypeptide coding genes can be obtained by injection of the DNA into reproductive organs of a plant as described (Pena *et al.*, 1987). DNA can also be injected directly into the cells of immature embryos and the rehydration of desiccated embryos as described (Neuhaus *et al.*, 1987; Benbrook *et al.*, 1986).

The development or regeneration of plants from either single plant protoplasts or various explants is well known in the art (Weissbach and Weissbach, 1988). This regeneration and growth process typically includes the steps of selection of transformed cells, culturing those individualized cells through the usual stages of embryonic development through the rooted plantlet stage. Transgenic embryos and seeds are similarly regenerated. The resulting transgenic rooted shoots are thereafter planted in an appropriate plant growth medium such as soil.

The development or regeneration of plants containing the foreign, exogenous gene that encodes a polypeptide of interest introduced by *Agrobacterium* from leaf explants can be achieved by methods well known in the art such as described (Horsch *et al.*, 1985). In this procedure, transformants are cultured in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant strain being transformed as described (Fraley *et al.*, 1983). In particular, U. S. Patent 5,349,124 (specification incorporated herein by reference) details the creation of genetically transformed lettuce cells and plants resulting therefrom which express hybrid crystal proteins conferring insecticidal activity against Lepidopteran larvae to such plants.

This procedure typically produces shoots within two to four months and those shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth. Shoots that rooted in the presence of the selective agent to form plantlets are then transplanted to soil or other media to allow the production of roots. These procedures vary depending upon the particular plant strain employed, such variations being well known in the art.

Preferably, the regenerated plants are self-pollinated to provide homozygous transgenic plants, as discussed before. Otherwise, pollen obtained from the regenerated plants is crossed to seed-grown plants of agronomically important, preferably inbred lines. Conversely, pollen from plants of those important lines is used to pollinate regenerated plants. A transgenic plant of the present invention containing a desired polypeptide is cultivated using methods well known to one skilled in the art.

A transgenic plant of this invention thus has an increased amount of a coding region (*e.g.*, a *cry* gene) that encodes one or more of the Chimeric Cry polypeptides disclosed herein. A preferred transgenic plant is an independent segregant and can transmit that gene and its activity to its progeny. A more preferred transgenic plant is homozygous for that gene, and transmits that gene to all of its offspring on sexual mating. Seed from a transgenic plant may be grown in the field or greenhouse, and resulting sexually mature transgenic plants are self-pollinated to generate true breeding plants. The progeny from these plants become true breeding lines that are evaluated for, by way of example, increased insecticidal capacity against Coleopteran insects, preferably in the

field, under a range of environmental conditions. The inventors contemplate that the present invention will find particular utility in the creation of transgenic corn, soybeans, cotton, wheat, oats, barley, other grains, vegetables, fruits, fruit trees, berries, turf grass, ornamentals, shrubs and trees.

5

6. EXAMPLES

The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

15

6.1 EXAMPLE 1 — CONSTRUCTION OF HYBRID *B. THURINGIENSIS* δ -ENDOTOXINS

The *B. thuringiensis* shuttle vectors pEG853, pEG854, and pEG857 which are used in the present invention have been described (Baum *et al.*, 1990). pEG857 contains the *CryIAc* gene cloned into pEG853 as an *SphI*-*BamHI* DNA fragment. pEG1064 was constructed in such a way that the *KpnI* site within the *cryIAc* gene was preserved and the *KpnI* site in the pEG857 multiple cloning site (MCS) was eliminated. This was accomplished by sequentially subjecting pEG857 DNA to limited *KpnI* digestion so that only one *KpnI* site is cut, filling in the *KpnI* 5' overhang by Klenow fragment of DNA polymerase I to create blunt DNA ends, and joining the blunt ends of DNA by T4 DNA ligase: pEG318 contains the *cryIF* gene (Chambers *et al.*, 1991) cloned into the *XhoI* site of pEG854 as an *XhoI*-*SalI* DNA fragment. pEG315 contains the *cryIC* gene from strain EG6346 (Chambers *et al.*, 1991) cloned into the *XhoI*-*BamHI* sites of pEG854 as a *SalI*-*BamHI* DNA fragment.

FIG. 1A shows a schematic representation of the DNA encoding the complete *cryIAc*, *cryIAb*, *cryIC*, and *cryIF* genes contained on pEG854/pEG1064, pEG20,

30

pEG315, and pEG318, respectively. Unique restriction sites that were used in constructing certain hybrid genes are also shown. FIG. 1B shows a schematic representation of hybrid genes pertaining to the present invention. In some cases standard PCRTM amplification with mutagenic oligonucleotide primers were used to incorporate appropriate restrictions sites into DNA fragments used for hybrid gene construction. Certain hybrid gene constructions could not be accomplished by restriction fragment subcloning. In those instances, PCRTM overlap extension (POE) was used to construct the desired hybrid gene (Horton *et al.*, 1989). The following oligonucleotide primers (purchased from Integrated DNA Technologies Inc., Coralville, IA) were used:

10

Primer A: 5'-GGATAGCACTCATCAAAGGTACC-3' (SEQ ID NO:1)
 Primer B: 5'-GAAGATATCCAATTCGAACAGTTTCCC-3' (SEQ ID NO:2)
 Primer C: 5'-CATATTCTGCCTCGAGTGTTCAGTAAC-3' (SEQ ID NO:3)
 Primer D: 5'-CCCGATCGGCCGCATGC-3' (SEQ ID NO:4)
 15 Primer E: 5'-CATTGGAGCTCTCCATG-3' (SEQ ID NO:5)
 Primer F: 5'-GCACTACGATGTATCC-3' (SEQ ID NO:6)
 Primer G: 5'-CATCGTAGTGCAACTCTTAC-3' (SEQ ID NO:7)
 Primer H: 5'-CCAAGAAAATACTAGAGCTCTTGTAAAAAAGGTGTTCC-3' (SEQ ID NO:8)
 Primer I: 5'-ATTTGAGTAATACTATCC-3' (SEQ ID NO:23)
 20 Primer J: 5'-ATTACTCAAATACCATTGG-3' (SEQ ID NO:24)
 Primer K: 5'-TCGTTGCTCTGTTCCCG-3' (SEQ ID NO:31)

The plasmids described in FIG. 1B containing the hybrid δ -endotoxin genes pertinent to this invention are described below. Isolation or purification of DNA fragments generated by restriction of plasmid DNA, PCRTM amplification, or POE refers to the sequential application of agarose-TAE gel electrophoresis and use of the GeneClean Kit (Bio 101) following the manufacturer's recommendation. pEG1065 was constructed by PCRTM amplification of the *cryIF* DNA fragment using primer pair A and B and pEG318 as the DNA template. The resulting PCRTM product was isolated, cut with *AsuII* and *KpnI*, and used to replace the corresponding *AsuII-KpnI* DNA fragment in pEG857. Plasmid pEG1067 was constructed using POE and DNA fragments *SauI-KpnI* of *cryIF*

and *AsuII*-*Clal* of *cryI*Ac that were isolated from pEG318 and pEG857, respectively. The resulting POE product was PCR[™] amplified with primer pair A and B, cut with *AsuII* and *KpnI*, and used to replace the corresponding *AsuII*-*KpnI* fragment in pEG857.

pEG1068 was constructed by replacing the *SacI*-*KpnI* DNA fragment of *cryI*Ac
5 isolated from pEG857 with the corresponding *SacI*-*KpnI* DNA fragment isolated from
cryIF (pEG318). pEG1070 was constructed by replacing the *SacI*-*KpnI* DNA fragment
isolated from pEG1065 with the corresponding *SacI*-*KpnI* DNA fragment isolated from
*cryI*Ac (pEG857). pEG1072 was constructed by replacing the *SacI*-*KpnI* DNA fragment
isolated from pEG1067 with the corresponding *SacI*-*KpnI* DNA fragment isolated from
10 *cryI*Ac (pEG857). pEG1074, pEG1076, and pEG1077 were constructed by replacing the
SphI-*XhoI* DNA fragment from pEG1064 with the PCR[™] amplified *SphI*-*XhoI* DNA
fragment from pEG1065, pEG1067, pEG1068, respectively, using primer pairs C and D.
pEG1089 was constructed by replacing the *SphI*-*SacI* DNA fragment of pEG1064 with
the isolated and *SphI* and *SacI* cut PCR[™] product of *cryIF* that was generated using
15 primer pair D and E and the template pEG318.

pEG1091 was constructed by replacing the *SphI*-*SacI* DNA fragment of pEG1064
with the isolated and *SphI* and *SacI* cut PCR[™] product of *cryIC* that was generated using
primer pair D and H and the template pEG315.

pEG1088 was constructed by POE using a *cryI*Ac DNA fragment generated using
20 primer pair B and F and a *cryIC* DNA fragment generated using primer pair A and G.
The *SacI*-*KpnI* fragment was isolated from the resulting POE product and used to replace
the corresponding *SacI*-*KpnI* fragment in pEG1064.

pEG365 was constructed by first replacing the *SphI*-*KpnI* DNA fragment from
pEG1065 with the corresponding *cryI*Ab DNA fragment isolated from pEG20 to give
25 pEG364. The *SacI*-*KpnI* DNA fragment from pEG364 was then replaced with the
corresponding *cryIF* DNA fragment isolated from pEG318.

pEG1092 was constructed by replacing the *KpnI*-*Bam*HI DNA fragment from
pEG1088 with the corresponding DNA fragment isolated from pEG315. pEG1092 is
distinct from the *cryI*Ab/*cryIC* hybrid δ -endotoxin gene disclosed in Intl. Pat. Appl.
30 Publ. No. WO 95/06730.

pEG1093 was constructed by replacing the *SphI*-*AsuII* DNA fragment from pEG1068 with the corresponding *SphI*-*AsuII* DNA fragment isolated from pEG20.

5 pEG378 was constructed by POE using a *cryI**Ac* DNA fragment generated using primer pair B and I using pEG857 as the template and a *cryI**F* DNA fragment generated using primer pair A and J using pEG318 as the template. The resulting POE product was cut with *AsuII* and *KpnI* and the resulting isolated DNA fragment used to replace the corresponding *AsuII*-*KpnI* DNA fragment in pEG1064.

10 pEG381 was constructed by replacing the *AsuII*-*XhoI* DNA fragment in pEG1064 with the corresponding *AsuII*-*XhoI* DNA fragment isolated from the PCR™ amplification of pEG378 using primer pair C and K.

6.2 EXAMPLE 2 -- PRODUCTION OF THE HYBRID TOXINS IN *B. THURINGIENSIS*

15 The plasmids encoding the hybrid toxins described in Example 1 were transformed into *B. thuringiensis* as described (Mettus and Macaluso, 1990). The resulting *B. thuringiensis* strains were grown in 50 ml of C-2 medium until the culture was fully sporulated and lysed (approximately 48 hr.). Since crystal formation is a prerequisite for efficient commercial production of δ -endotoxins in *B. thuringiensis*, microscopic analysis was used to identify crystals in the sporulated cultures (Table 4).

TABLE 4
CRYSTAL FORMATION BY THE HYBRID δ -ENDOTOXINS

Strain	Plasmid	Parent δ -Endotoxins	Crystal Formation
EG11060	pEG1065	Cry1Ac + Cry1F	+
EG11062	pEG1067	Cry1Ac + Cry1F	+
EG11063	pEG1068	Cry1Ac + Cry1F	+
EG11065	pEG1070	Cry1Ac + Cry1F	-
EG11067	pEG1072	Cry1Ac + Cry1F	-
EG11071	pEG1074	Cry1Ac + Cry1F	+
EG11073	pEG1076	Cry1Ac + Cry1F	+
EG11074	pEG1077	Cry1Ac + Cry1F	+
EG11087	pEG1088	Cry1Ac + Cry1C	-
EG11088	pEG1089	Cry1F + Cry1Ac	-
EG11090	pEG1091	Cry1C + Cry1Ac	-
EG11091	pEG1092	Cry1Ac + Cry1C	+
EG11092	pEG1093	Cry1Ab + Cry1Ac + Cry1F	+
EG11735	pEG365	Cry1Ab + Cry1F + Cry1Ac	+
EG11751	pEG378	Cry1Ac + Cry1F	+
EG11768	pEG381	Cry1Ac + Cry1F	+

5 The δ -endotoxin production for some of the *B. thuringiensis* strains specified in
Table 4 was examined by sodium dodecyl sulfate-polyacrylamide gel electrophoresis
(SDS-PAGE) as described by Baum *et al.*, 1990. Equal volume cultures of each *B.*
thuringiensis strain were grown in C-2 medium until fully sporulated and lysed. The
cultures were centrifuged and the spore/crystal pellet was washed twice with equal
volumes of distilled deionized water. The final pellet was suspended in half the culture
10 volume of 0.005% Triton X-100[®]. An equal volume of each washed culture was
analyzed by SDS-PAGE as shown in FIG. 2.

The majority of hybrids involving Cry1Ac and Cry1F formed stable crystals in *B. thuringiensis*. A notable exception is EG11088 in which the active toxin fragment would be the reciprocal exchange of EG11063. Two of the three hybrids involving Cry1Ac and Cry1C, EG11087 and EG11090, failed to produce crystal in *B. thuringiensis* even though these reciprocal hybrids mimic the activated toxin fragments of crystal-forming EG11063 and EG11074.

Every strain that was examined by SDS-PAGE produced some level of δ -endotoxin. As expected, however, those cultures identified as crystal negative produced very little protein (e.g., lane e: EG11065, lane f: EG11067, lane j: EG11088, and lane k: EG11090). For reference, typical yields from a crystal forming δ -endotoxin is shown for Cry1Ac (lane a). Several hybrid δ -endotoxins produce comparable levels of protein including EG11060 (lane b), EG11062 (lane c), EG11063 (lane d; SEQ ID NO:10), and EG11074 (lane i; SEQ ID NO:12). The data clearly show that efficient hybrid δ -endotoxin production in *B. thuringiensis* is unpredictable and varies depending on the parent δ -endotoxins used to construct the hybrid.

6.3 EXAMPLE 3 -- PROTEOLYTIC PROCESSING OF THE HYBRID δ -ENDOTOXINS

Proteolytic degradation of the protoxin form of the δ -endotoxin to a stable active toxin occurs once δ -endotoxin crystals are solubilized in the larval midgut. One measure of the potential activity of δ -endotoxins is the stability of the active δ -endotoxin in a proteolytic environment. To test the proteolytic sensitivity of the hybrid δ -endotoxins, solubilized toxin was subjected to trypsin digestion. The δ -endotoxins were purified from sporulated *B. thuringiensis* cultures and quantified as described (Chambers *et al.*, 1991). Exactly 250 μ g of each hybrid δ -endotoxin crystal was solubilized in 30 mM NaHCO₃, 10 mM DTT (total volume 0.5 ml). Trypsin was added to the solubilized toxin at a 1:10 ratio. At appropriate time points 50 μ l aliquots were removed to 50 μ l Laemmli buffer, heated to 100°C for 3 min., and frozen in a dry-ice ethanol bath for subsequent analysis. The trypsin digests of the solubilized toxins were analyzed by SDS-PAGE and the amount of active δ -endotoxin at each time point was quantified by densitometry. A graphic representation of the results from these studies are shown in FIG. 3.

The wild-type Cry1Ac is rapidly processed to the active δ -endotoxin fragment that is stable for the duration of the study. The hybrid δ -endotoxins from EG11063 and EG11074 are also processed to active δ -endotoxin fragments which are stable for the duration of the study. The processing of the EG11063 δ -endotoxin occurs at a slower rate and a higher percentage of this active δ -endotoxin fragment remains at each time point. Although the hybrid δ -endotoxins from EG11060 and EG11062 are process to active δ -endotoxin fragments, these fragments are more susceptible to further cleavage and degrade at various rates during the course of the study. The 5' exchange points between *cry1Ac* and *cry1F* for the EG11062 and EG11063 δ -endotoxins result in toxins that differ by only 21 amino acid residues (see FIG. 1). However, the importance of maintaining *Cry1Ac* sequences at these positions is evident by the more rapid degradation of the EG11062 δ -endotoxin. These data demonstrate that different hybrid δ -endotoxins constructed using the same parental δ -endotoxins can vary significantly in biochemical characteristics such as proteolytic stability.

6.4 EXAMPLE 4 -- BIOACTIVITY OF THE HYBRID δ -ENDOTOXINS

B. thuringiensis cultures expressing the desired δ -endotoxin were grown until fully sporulated and lysed and washed as described in Example 2. The δ -endotoxin levels for each culture were quantified by SDS-PAGE as described (Baum *et al.*, 1990). In the case of bioassay screens, a single appropriate concentration of each washed δ -endotoxin culture was topically applied to 32 wells containing 1.0 ml artificial diet per well (surface area of 175 mm²). A single neonate larvae was placed in each of the treated wells and the tray covered by a clear perforated mylar sheet. Larvae mortality was scored after 7 days of feeding and percent mortality expressed as the ratio of the number of dead larvae to the total number of larvae treated, 32.

In the case of LC₅₀ determinations (δ -endotoxin concentration giving 50% mortality), δ -endotoxins were purified from the *B. thuringiensis* cultures and quantified as described by Chambers *et al.* (1991). Eight concentrations of the δ -endotoxins were prepared by serial dilution in 0.005% Triton X-100[®] and each concentration was topically applied to wells containing 1.0 ml of artificial diet. Larvae mortality was scored after 7

days of feeding (32 larvae for each δ -endotoxin concentration). In all cases the diluent served as the control.

5 A comparison of the Cry1A/Cry1F hybrid toxins by bioassay screens is shown in Table 5. The hybrid δ -endotoxins from strains EG11063 and EG11074 maintain the activities of the parental Cry1Ac and Cry1F δ -endotoxins. Furthermore, the hybrid
10 δ -endotoxin from EG11735 maintains the activity of its parental Cry1Ab and Cry1F δ -endotoxins. The δ -endotoxins produce by strains EG11061, EG11062, EG11071, and EG11073 have no insecticidal activity on the insect larvae tested despite 1) being comprised of at least one parental δ -endotoxin that is active against the indicated larvae and 2) forming stable, well-defined crystals in *B. thuringiensis*. These results demonstrate the unpredictable nature of hybrid toxin constructions.

For the data in Table 5. All strains were tested as washed sporulated cultures. For each insect tested, equivalent amounts of δ -endotoxins were used and insecticidal activity was based on the strain showing the highest percent mortality (++++).

TABLE 5
BIOASSAY SCREENS OF HYBRID CRY1A/CRY1F δ -ENDOTOXINS

Strain	<i>S. frugiperda</i>	<i>S. exigua</i>	<i>H. virescens</i>	<i>H. zea</i>	<i>O. nubilalis</i>
Cry1Ac	-	-	++++	++++	+++
Cry1F	++++	++	++	++	++
Cry1Ab	++	+	+++	++	+++
EG11060	-	-	-	-	-
EG11062	-	-	-	-	-
EG11063	++++	++++	+++	+++	++++
EG11071	-	-	-	-	-
EG11073	-	-	-	-	-
EG11074	++++	++++	+++	+++	++++
EG11090	-	+++	-	-	-
EG11091	++++	++++	-	-	N.D.
EG11092	++++	++++	+++	+++	N.D.
EG11735	++++	++++	+++	+++	N.D.
EG11751	N.D. ^a	++++	N.D.	++++	N.D.

^aN.D. = not determined.

The δ -endotoxins described in FIG. 1 and that demonstrated insecticidal activity in bioassay screens were tested as purified crystals to determine their LC₅₀ (see Table 6). The δ -endotoxins purified from strains EG11063, EG11074, EG11091, and EG11735 all show increased armyworm (*S. frugiperda* and *S. exigua*) activity compared to any of the wild-type δ -endotoxins tested. The EG11063 and EG11074 δ -endotoxins would yield identical active toxin fragments (FIG. 1B) which is evident by their similar LC50 values on the insects examined. An unexpected result evident from these data is that a hybrid δ -endotoxin such as EG11063, EG11092, EG11074, EG11735, or EG11751 can retain the activity of their respective parental δ -endotoxins, and, against certain insects such as *S. exigua*, can have activity far better than either parental δ -endotoxin. This broad range of insecticidal activity at doses close to or lower than the parental δ -endotoxins, along

with the wild-type level of toxin production (Example 2), make these proteins particularly suitable for production in *B. thuringiensis*. Although the EG11091 derived δ -endotoxin has better activity against *S. frugiperda* and *S. exigua* than its parental δ -endotoxins, it has lost the *H. virescens* and *H. zea* activity attributable to its Cry1Ac parent. This restricted host range along with lower toxin yield observed for the EG11091 δ -endotoxin (Example 2) make it less amenable to production in *B. thuringiensis*.

TABLE 6
LC₅₀ VALUES FOR THE PURIFIED HYBRID δ -ENDOTOXIN^A

Toxin	<i>S. frugiperda</i>	<i>S. exigua</i>	<i>H. virescens</i>	<i>H. zea</i>	<i>O. nubilalis</i>
Cry1Ac	>10000	>10000	9	100	23
Cry1Ab	1435	4740	118	400	17
Cry1C	>10000	490	>10000	>10000	>10000
Cry1F	1027	3233	54	800	51
EG11063	550	114	33	80	7
(Cry1Ac/1F)					
EG11074	468	77	25	76	9
(Cry1Ac/1F)					
EG11091	21	21	219	>10000	N.D. ^a
(Cry1Ac/1C)					

^aN.D.=not determined.

In Table 6, the LC₅₀ values are expressed in nanograms of purified δ -endotoxin per well (175 mm²) and are the composite values for 2 to 6 replications. nd = not determined.

TABLE 7
DNA EXCHANGE SITES FOR CRY1 HYBRID δ -ENDOTOXINS

Plasmid	SEQ ID NO:	5' Exchange Site	SEQ ID NO:	3' Exchange Site
pEG1065	15	TATCCAATTTCGAACGTCATC	21	ACTACCAGGTACCTTTTGATG
pEG1067	16	TTTAGTCATCGATTAAATCA	21	ACTACCAGGTACCTTTTGATG
pEG1068	17	ATAATAAGAGCTCCAATGTT	21	ACTACCAGGTACCTTTTGATG
pEG1070	15	TATCCAATTTCGAACGTCATC	19	TCATGGAGAGCTCCTATGTT
pEG1072	16	TTTAGTCATCGATTAAATCA	19	TCATGGAGAGCTCCTATGTT
pEG1074	15	TATCCAATTTCGAACGTCATC	35	TGCAACACTCGAGGCTGAAT
pEG1076	16	TTTAGTCATCGATTAAATCA	35	TGCAACACTCGAGGCTGAAT
pEG1077	17	ATAATAAGAGCTCCAATGTT	35	TGCAACACTCGAGGCTGAAT
pEG1088	18	TACATCGTAGTGCAACTCTT	22	ACTACCGGGTACCTTTTGATA
pEG1089	19	TCATGGAGAGCTCCTATGTT	-	NA
pEG1091	20	TTAACAAGAGCTCCTATGTT	-	NA
pEG1092	18	TACATCGTAGTGCAACTCTT	-	NA
pEG1093	-	ND ^b	21	ACTACCAGGTACCTTTTGATG
pEG365	17	ATAATAAGAGCTCCAATGTT	21	ACTACCAGGTACCTTTTGATG
pEG378	32	TCAAATACCATTTGGTAAAG	21	ACTACCAGGTACCTTTTGATG
pEG381	32	TCAAATACCATTTGGTAAAG	35	TGCAACACTCGAGGCTGAAT

^aNA = Not Applicable. These hybrid toxins contain only one exchange site as shown in FIG. 1.

^bND = Not Distinguishable. The exchange site for these hybrid proteins are identified by DNA sequences that are not distinguishable from either of the parent toxins.

Table 7 describes the DNA surrounding the 5' and 3' exchange points for the hybrid δ -endotoxins which are pertinent to the present invention. As evident by the SEQ ID NO, certain hybrid δ -endotoxins share exchange sites.

To examine the effect of other small changes in the exchange site chosen for hybrid endotoxin construction, the activity of EG11751 and EG11063 on *S. exigua* and *H. zea* were compared (Table 8). The data clearly show that hybrid δ -endotoxin improvements can be made by altering the exchange site between the two parental δ -endotoxins. In this example, the exchange site in the EG11751 δ -endotoxin was moved 75 base pairs 3' compared to the EG11063 δ -endotoxin and results in improved insecticidal activity. Although no significant improvement in *S. exigua* activity is observed between EG11063 and EG11751, a significant improvement in *H. zea* activity of almost 4-fold is observed for EG11751. It is important to note that improvements in hybrid δ -endotoxin bioactivity by altering exchange sites is unpredictable. In the case of EG11062, moving the exchange site 63 base pairs 5' of the EG11063 exchange site abolishes insecticidal activity as shown in Table 7.

TABLE 8
BIOACTIVITY OF EG11063 AND EG11751

<i>B. thuringiensis</i> Strain	LC ₅₀ Values for Washed Sporulated Cultures	
	<i>S. exigua</i>	<i>H. zea</i>
EG11063	106	38
EG11751	90	10.

To further examine the effect of changes in the exchange site for hybrid δ -endotoxins, the hybrid δ -endotoxin encoded by pEG381 was compared to those encoded by pEG378 and pEG1068. In this example, the 3' exchange site for the pEG381 encoded hybrid δ -endotoxin was moved 340 base pairs 5' compared to the pEG378 hybrid δ -endotoxin. The data in Table 8 show that this change results in an increase in *S. frugiperda* activity compared to the pEG378 and pEG1066 encoded δ -endotoxins while maintaining the increased activity that was observed for the pEG378 encoded δ -endotoxin over the pEG1068 encoded δ -endotoxin (see Table 7). This result is

unexpected since the activated toxin resulting from the proteolysis of the encoded δ -endotoxins from pEG378 and pEG381 should be identical. This example further demonstrates that exchange sites within the protoxin fragment of δ -endotoxins can have a profound effect on insecticidal activity.

5

TABLE 9
BIOACTIVITY OF TOXINS ENCODED BY PEG378, PEG381 AND PEG1068

Plasmid	LC ₅₀ Values for Purified Crystals			
	<i>S. frugiperda</i>	<i>T. ni</i>	<i>H. zea</i>	<i>P. xylostella</i>
pEG378	464	57.7	37.5	3.02
pEG381	274	56.0	36.6	2.03
pEG1068	476	66.7	72.7	3.83

6.5 EXAMPLE 5 – ACTIVITY OF THE HYBRID TOXINS ON ADDITIONAL PESTS

10 The toxins of the present invention were also assayed against additional pests, including the southwestern corn borer and two pests active against soybean. Toxin proteins were solubilized, added to diet and bioassayed against target pests. The hybrid toxins showed very effective control of all three pests.

TABLE 10

LC₅₀ AND EC₅₀ RANGES OF HYBRID TOXINS ON SOUTHWESTERN CORN BORER^{1,2}

	EG11063	EG11074	EG11091	EG11751
LC ₅₀	20	10-20	10-20	10-20
EC ₅₀	0.2-2	0.2-2	0.2-2	0.2-2

¹All values are expressed in µg/ml of diet.²SWCB data ranges represent LC₅₀ and EC₅₀ ranges (as determined by % >1st

5 instar), respectively.

TABLE 11

LC₅₀ VALUES OF CHIMERIC CRYSTAL PROTEINS ON SOYBEAN PESTS¹

Pest	EG11063	EG11074	EG11091	EG11751	EG11768
Velvetbean caterpillar ¹	0.9	0.6	0.3	0.1	0.06
Soybean looper	0.9	0.8	0.6	0.7	0.2

¹All values are expressed in µg/ml of diet.²Velvetbean caterpillar (*Anticarsia gemmatilis*) and soybean looper (*Psuedoplusi*10 *inclusens*) are both members of the family *Noctuidae*.

6.6 EXAMPLE 6 -- AMINO ACID SEQUENCES OF THE NOVEL CRYSTAL PROTEINS

6.6.1 AMINO ACID SEQUENCE OF THE EG11063 CRYSTAL PROTEIN (SEQ ID NO:10)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
 15 GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
 TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
 ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 20 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
 LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaValArgTrpTyrAsnThrGlyLeuGlu
 ArgValTrpGlyProAspSerArgAspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrProIleArgThrValSerGlnLeuThr

ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
 TyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
 5 ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp
 SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpThr
 HisArgSerAlaThrProThrAsnThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
 10 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGlyGlyAspIleLeuArgArgThrSer
 GlyGlyProPheAlaTyrThrIleValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGluArgIlePheAlaGlyGlnPheAsn
 LysThrMetAspThrGlyAspProLeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSerSerGlyAsnGluValTyrIleAsp
 15 ArgPheGluLeuIleProValThrAlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspValThrAspTyrHisIleAspGlnVal
 SerAsnLeuValAspCysLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsnPheLysGlyIleAsnArgGlnLeu
 AspArgGlyTrpArgGlySerThrAspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 20 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLysIleAspGluSerLysLeuLys
 AlaPheThrArgTyrGlnLeuArgGlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerProIle
 GlyLysCysGlyGluProAsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIleAspValGlyCysThrAspLeuAsn
 25 GluAspLeuGlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 LysArgGluLysLeuGluTrpGluThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMetIleHisAlaAlaAspLysArgVal
 HisSerIleArgGluAlaTyrLeuProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 30 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsnValIleLysAsnGlyAspPheAsn
 AsnGlyLeuSerCysTrpAsnValLysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCysProGlyArgGlyTyrIleLeuArg
 ValThrAlaTyrLysGluGlyTyrGlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThrValThrCysAsnAspTyrThrVal
 35 AsnGlnGluGluTyrGlyGlyAlaTyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCysGluPheAsnArg

GlyTyrArgAspTyrThrProLeuProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGlu
Glu

5 6.6.2 AMINO ACID SEQUENCE OF THE EG11074 CRYSTAL PROTEIN (SEQ ID NO:12)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
10 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaValArgTrpTyrAsnThrGlyLeuGlu
15 ArgValTrpGlyProAspSerArgAspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrProIleArgThrValSerGlnLeuThr
ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
TyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
20 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp
SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpThr
25 HisArgSerAlaThrProThrAsnThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGlyGlyAspIleLeuArgArgThrSer
GlyGlyProPheAlaTyrThrIleValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGluArgIlePheAlaGlyGlnPheAsn
LysThrMetAspThrGlyAspProLeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
30 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSerSerGlyAsnGluValTyrIleAsp
ArgPheGluLeuIleProValThrAlaThrLeuGluAlaGluTyrAsnLeuGluArgAlaGlnLysAlaVal
AsnAlaLeuPheThrSerThrAsnGlnLeuGlyLeuLysThrAsnValThrAspTyrHisIleAspGlnVal
SerAsnLeuValThrTyrLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspSerAsnPheLysAspIleAsnArgGlnPro
35 GluArgGlyTrpGlyGlySerThrGlyIleThrIleGlnGlyGlyAspAspValPheLysGluAsnTyrVal
ThrLeuSerGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLysIleAspGluSerLysLeuLys

AlaPheThrArgTyrGlnLeuArgGlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerProIle
 GlyLysCysGlyGluProAsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIleAspValGlyCysThrAspLeuAsn
 5 GluAspLeuGlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 LysArgGluLysLeuGluTrpGluThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMetIleHisAlaAlaAspLysArgVal
 HisSerIleArgGluAlaTyrLeuProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 10 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsnValIleLysAsnGlyAspPheAsn
 AsnGlyLeuSerCysTrpAsnValLysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCysProGlyArgGlyTyrIleLeuArg
 ValThrAlaTyrLysGluGlyTyrGlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThrValThrCysAsnAspTyrThrVal
 15 AsnGlnGluGluTyrGlyGlyAlaTyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCysGluPheAsnArg
 GlyTyrArgAspTyrThrProLeuProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGlu
 Glu

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6.6.3 AMINO ACID SEQUENCE OF THE EG11735 CRYSTAL PROTEIN (SEQ ID NO:14)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
 GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
 25 TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
 ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
 LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 30 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspHisAlaValArgTrpTyrAsnThrGlyLeuGlu
 ArgValTrpGlyProAspSerArgAspTrpIleArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValSerLeuPheProAsnTyrAspSerArgThrTyrProIleArgThrValSerGlnLeuThr
 ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 GlySerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
 35 GluTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro

LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
 ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp
 SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 5 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpThr
 HisArgSerAlaThrProThrAsnThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGlyGlyAspIleLeuArgArgThrSer
 GlyGlyProPheAlaTyrThrIleValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGluArgIlePheAlaGlyGlnPheAsn
 10 LysThrMetAspThrGlyAspProLeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSerSerGlyAsnGluValTyrIleAsp
 ArgPheGluLeuIleProValThrAlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspValThrAspTyrHisIleAspGlnVal
 SerAsnLeuValAspCysLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 15 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsnPheLysGlyIleAsnArgGlnLeu
 AspArgGlyTrpArgGlySerThrAspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLysIleAspGluSerLysLeuLys
 AlaPheThrArgTyrGlnLeuArgGlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerProIle
 20 GlyLysCysGlyGluProAsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIleAspValGlyCysThrAspLeuAsn
 GluAspLeuGlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 LysArgGluLysLeuGluTrpGluThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 25 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMetIleHisAlaAlaAspLysArgVal
 HisSerIleArgGluAlaTyrLeuProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsnValIleLysAsnGlyAspPheAsn
 AsnGlyLeuSerCysTrpAsnValLysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCysProGlyArgGlyTyrIleLeuArg
 30 ValThrAlaTyrLysGluGlyTyrGlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThrValThrCysAsnAspTyrThrVal
 AsnGlnGluGluTyrGlyGlyAlaTyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCysGluPheAsnArg
 GlyTyrArgAspTyrThrProLeuProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 35 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGlu
 Glu

6.6.4 AMINO ACID SEQUENCE OF THE EG11092 CRYSTAL PROTEIN (SEQ ID NO:26)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
5 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
10 LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspHisAlaValArgTrpTyrAsnThrGlyLeuGlu
ArgValTrpGlyProAspSerArgAspTrpIleArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
LeuAspIleValSerLeuPheProAsnTyrAspSerArgThrTyrProIleArgThrValSerGlnLeuThr
ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
15 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
TyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp
20 SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpThr
HisArgSerAlaThrProThrAsnThrIleAspProGluArgIleThrGlnIleProLeuValLysAlaHis
ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGlyGlyAspIleLeuArgArgThrSer
GlyGlyProPheAlaTyrThrIleValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
25 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGluArgIlePheAlaGlyGlnPheAsn
LysThrMetAspThrGlyAspProLeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSerSerGlyAsnGluValTyrIleAsp
ArgPheGluLeuIleProValThrAlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspValThrAspTyrHisIleAspGlnVal
30 SerAsnLeuValAspCysLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsnPheLysGlyIleAsnArgGlnLeu
AspArgGlyTrpArgGlySerThrAspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLysIleAspGluSerLysLeuLys
AlaPheThrArgTyrGlnLeuArgGlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
35 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerProIle

GlyLysCysGlyGluProAsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIleAspValGlyCysThrAspLeuAsn
 GluAspLeuGlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 5 LysArgGluLysLeuGluTrpGluThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMetIleHisAlaAlaAspLysArgVal
 HisSerIleArgGluAlaTyrLeuProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsnValIleLysAsnGlyAspPheAsn
 AsnGlyLeuSerCysTrpAsnValLysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 10 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCysProGlyArgGlyTyrIleLeuArg
 ValThrAlaTyrLysGluGlyTyrGlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThrValThrCysAsnAspTyrThrVal
 AsnGlnGluGluTyrGlyGlyAlaTyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCysGluPheAsnArg
 15 GlyTyrArgAspTyrThrProLeuProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGlu
 Glu

6.6.5 AMINO ACID SEQUENCE OF THE EG11751 CRYSTAL PROTEIN (SEQ ID NO:28)

20 MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
 GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
 TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
 25 ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
 LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaValArgTrpTyrAsnThrGlyLeuGlu
 ArgValTrpGlyProAspSerArgAspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 30 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrProIleArgThrValSerGlnLeuThr
 ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
 TyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
 35 ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp

GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp
 SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpIle
 HisArgSerAlaGluPheAsnAsnIleIleAlaSerAspSerIleThrGlnIleProLeuValLysAlaHis
 5 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGlyGlyAspIleLeuArgArgThrSer
 GlyGlyProPheAlaTyrThrIleValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGluArgIlePheAlaGlyGlnPheAsn
 LysThrMetAspThrGlyAspProLeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSerSerGlyAsnGluValTyrIleAsp
 10 ArgPheGluLeuIleProValThrAlaThrPheGluAlaGluTyrAspLeuGluArgAlaGlnLysAlaVal
 AsnAlaLeuPheThrSerIleAsnGlnIleGlyIleLysThrAspValThrAspTyrHisIleAspGlnVal
 SerAsnLeuValAspCysLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspProAsnPheLysGlyIleAsnArgGlnLeu
 AspArgGlyTrpArgGlySerThrAspIleThrIleGlnArgGlyAspAspValPheLysGluAsnTyrVal
 15 ThrLeuProGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLysIleAspGluSerLysLeuLys
 AlaPheThrArgTyrGlnLeuArgGlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerProIle
 GlyLysCysGlyGluProAsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIleAspValGlyCysThrAspLeuAsn
 20 GluAspLeuGlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 LysArgGluLysLeuGluTrpGluThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMetIleHisAlaAlaAspLysArgVal
 HisSerIleArgGluAlaTyrLeuProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 25 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsnValIleLysAsnGlyAspPheAsn
 AsnGlyLeuSerCysTrpAsnValLysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCysProGlyArgGlyTyrIleLeuArg
 ValThrAlaTyrLysGluGlyTyrGlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThrValThrCysAsnAspTyrThrVal
 30 AsnGlnGluGluTyrGlyGlyAlaTyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCysGluPheAsnArg
 GlyTyrArgAspTyrThrProLeuProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGlu
 Glu
 35

6.6.6 AMINO ACID SEQUENCE OF THE EG11091 CRYSTAL PROTEIN (SEQ ID NO:30)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
5 TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
10 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaValArgTrpTyrAsnThrGlyLeuGlu
ArgValTrpGlyProAspSerArgAspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrProIleArgThrValSerGlnLeuThr
ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
15 TyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp
SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
20 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpIle
HisArgSerAlaThrLeuThrAsnThrIleAspProGluArgIleAsnGlnIleProLeuValLysGlyPhe
ArgValTrpGlyGlyThrSerValIleThrGlyProGlyPheThrGlyGlyAspIleLeuArgArgAsnThr
PheGlyAspPheValSerLeuGlnValAsnIleAsnSerProIleThrGlnArgTyrArgLeuArgPheArg
TyrAlaSerSerArgAspAlaArgValIleValLeuThrGlyAlaAlaSerThrGlyValGlyGlyGlnVal
25 SerValAsnMetProLeuGlnLysThrMetGluIleGlyGluAsnLeuThrSerArgThrPheArgTyrThr
AspPheSerAsnProPheSerPheArgAlaAsnProAspIleIleGlyIleSerGluGlnProLeuPheGly
AlaGlySerIleSerSerGlyGluLeuTyrIleAspLysIleGluIleIleLeuAlaAspAlaThrPheGlu
AlaGluSerAspLeuGluArgAlaGlnLysAlaValAsnAlaLeuPheThrSerSerAsnGlnIleGlyLeu
LysThrAspValThrAspTyrHisIleAspGlnValSerAsnLeuValAspCysLeuSerAspGluPheCys
30 LeuAspGluLysArgGluLeuSerGluLysValLysHisAlaLysArgLeuSerAspGluArgAsnLeuLeu
GlnAspProAsnPheArgGlyIleAsnArgGlnProAspArgGlyTrpArgGlySerThrAspIleThrIle
GlnGlyGlyAspAspValPheLysGluAsnTyrValThrLeuProGlyThrValAspGluCysTyrProThr
TyrLeuTyrGlnLysIleAspGluSerLysLeuLysAlaTyrThrArgTyrGluLeuArgGlyTyrIleGlu
AspSerGlnAspLeuGluIleTyrLeuIleArgTyrAsnAlaLysHisGluIleValAsnValProGlyThr
35 GlySerLeuTrpProLeuSerAlaGlnSerProIleGlyLysCysGlyGluProAsnArgCysAlaProHis

LeuGluTrpAsnProAspLeuAspCysSerCysArgAspGlyGluLysCysAlaHisHisSerHisHisPhe
 ThrLeuAspIleAspValGlyCysThrAspLeuAsnGluAspLeuGlyValTrpValIlePheLysIleLys
 ThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGluPheLeuGluGluLysProLeuLeuGlyGluAlaLeu
 AlaArgValLysArgAlaGluLysLysTrpArgAspLysArgGluLysLeuGlnLeuGluThrAsnIleVal
 5 TyrLysGluAlaLysGluSerValAspAlaLeuPheValAsnSerGlnTyrAspArgLeuGlnValAspThr
 AsnIleAlaMetIleHisAlaAlaAspLysArgValHisArgIleArgGluAlaTyrLeuProGluLeuSer
 ValIleProGlyValAsnAlaAlaIlePheGluGluLeuGluGlyArgIlePheThrAlaTyrSerLeuTyr
 AspAlaArgAsnValIleLysAsnGlyAspPheAsnAsnGlyLeuLeuCysTrpAsnValLysGlyHisVal
 AspValGluGluGlnAsnAsnHisArgSerValLeuValIleProGluTrpGluAlaGluValSerGlnGlu
 10 ValArgValCysProGlyArgGlyTyrIleLeuArgValThrAlaTyrLysGluGlyTyrGlyGluGlyCys
 ValThrIleHisGluIleGluAspAsnThrAspGluLeuLysPheSerAsnCysValGluGluGluValTyr
 ProAsnAsnThrValThrCysAsnAsnTyrThrGlyThrGlnGluGluTyrGluGlyThrTyrThrSerArg
 AsnGlnGlyTyrAspGluAlaTyrGlyAsnAsnProSerValProAlaAspTyrAlaSerValTyrGluGlu
 LysSerTyrThrAspGlyArgArgGluAsnProCysGluSerAsnArgGlyTyrGlyAspTyrThrProLeu
 15 ProAlaGlyTyrValThrLysAspLeuGluTyrPheProGluThrAspLysValTrpIleGluIleGlyGlu
 ThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGluGlu

6.6.7 AMINO ACID SEQUENCE OF THE EG11768 CRYSTAL PROTEIN (SEQ ID NO:34)

MetAspAsnAsnProAsnIleAsnGluCysIleProTyrAsnCysLeuSerAsnProGluValGluValLeu
 20 GlyGlyGluArgIleGluThrGlyTyrThrProIleAspIleSerLeuSerLeuThrGlnPheLeuLeuSer
 GluPheValProGlyAlaGlyPheValLeuGlyLeuValAspIleIleTrpGlyIlePheGlyProSerGln
 TrpAspAlaPheLeuValGlnIleGluGlnLeuIleAsnGlnArgIleGluGluPheAlaArgAsnGlnAla
 IleSerArgLeuGluGlyLeuSerAsnLeuTyrGlnIleTyrAlaGluSerPheArgGluTrpGluAlaAsp
 ProThrAsnProAlaLeuArgGluGluMetArgIleGlnPheAsnAspMetAsnSerAlaLeuThrThrAla
 25 IleProLeuPheAlaValGlnAsnTyrGlnValProLeuLeuSerValTyrValGlnAlaAlaAsnLeuHis
 LeuSerValLeuArgAspValSerValPheGlyGlnArgTrpGlyPheAspAlaAlaThrIleAsnSerArg
 TyrAsnAspLeuThrArgLeuIleGlyAsnTyrThrAspTyrAlaValArgTrpTyrAsnThrGlyLeuGlu
 ArgValTrpGlyProAspSerArgAspTrpValArgTyrAsnGlnPheArgArgGluLeuThrLeuThrVal
 LeuAspIleValAlaLeuPheProAsnTyrAspSerArgArgTyrProIleArgThrValSerGlnLeuThr
 30 ArgGluIleTyrThrAsnProValLeuGluAsnPheAspGlySerPheArgGlySerAlaGlnGlyIleGlu
 ArgSerIleArgSerProHisLeuMetAspIleLeuAsnSerIleThrIleTyrThrAspAlaHisArgGly
 TyrTyrTyrTrpSerGlyHisGlnIleMetAlaSerProValGlyPheSerGlyProGluPheThrPhePro
 LeuTyrGlyThrMetGlyAsnAlaAlaProGlnGlnArgIleValAlaGlnLeuGlyGlnGlyValTyrArg
 ThrLeuSerSerThrLeuTyrArgArgProPheAsnIleGlyIleAsnAsnGlnGlnLeuSerValLeuAsp
 35 GlyThrGluPheAlaTyrGlyThrSerSerAsnLeuProSerAlaValTyrArgLysSerGlyThrValAsp

SerLeuAspGluIleProProGlnAsnAsnAsnValProProArgGlnGlyPheSerHisArgLeuSerHis
 ValSerMetPheArgSerGlyPheSerAsnSerSerValSerIleIleArgAlaProMetPheSerTrpIle
 HisArgSerAlaGluPheAsnAsnIleIleAlaSerAspSerIleThrGlnIleProLeuValLysAlaHis
 ThrLeuGlnSerGlyThrThrValValArgGlyProGlyPheThrGlyGlyAspIleLeuArgArgThrSer
 5 GlyGlyProPheAlaTyrThrIleValAsnIleAsnGlyGlnLeuProGlnArgTyrArgAlaArgIleArg
 TyrAlaSerThrThrAsnLeuArgIleTyrValThrValAlaGlyGluArgIlePheAlaGlyGlnPheAsn
 LysThrMetAspThrGlyAspProLeuThrPheGlnSerPheSerTyrAlaThrIleAsnThrAlaPheThr
 PheProMetSerGlnSerSerPheThrValGlyAlaAspThrPheSerSerGlyAsnGluValTyrIleAsp
 ArgPheGluLeuIleProValThrAlaThrLeuGluAlaGluTyrAsnLeuGluArgAlaGlnLysAlaVal
 10 AsnAlaLeuPheThrSerThrAsnGlnLeuGlyLeuLysThrAsnValThrAspTyrHisIleAspGlnVal
 SerAsnLeuValThrTyrLeuSerAspGluPheCysLeuAspGluLysArgGluLeuSerGluLysValLys
 HisAlaLysArgLeuSerAspGluArgAsnLeuLeuGlnAspSerAsnPheLysAspIleAsnArgGlnPro
 GluArgGlyTrpGlyGlySerThrGlyIleThrIleGlnGlyGlyAspAspValPheLysGluAsnTyrVal
 ThrLeuSerGlyThrPheAspGluCysTyrProThrTyrLeuTyrGlnLysIleAspGluSerLysLeuLys
 15 AlaPheThrArgTyrGlnLeuArgGlyTyrIleGluAspSerGlnAspLeuGluIleTyrLeuIleArgTyr
 AsnAlaLysHisGluThrValAsnValProGlyThrGlySerLeuTrpProLeuSerAlaGlnSerProIle
 GlyLysCysGlyGluProAsnArgCysAlaProHisLeuGluTrpAsnProAspLeuAspCysSerCysArg
 AspGlyGluLysCysAlaHisHisSerHisHisPheSerLeuAspIleAspValGlyCysThrAspLeuAsn
 GluAspLeuGlyValTrpValIlePheLysIleLysThrGlnAspGlyHisAlaArgLeuGlyAsnLeuGlu
 20 PheLeuGluGluLysProLeuValGlyGluAlaLeuAlaArgValLysArgAlaGluLysLysTrpArgAsp
 LysArgGluLysLeuGluTrpGluThrAsnIleValTyrLysGluAlaLysGluSerValAspAlaLeuPhe
 ValAsnSerGlnTyrAspGlnLeuGlnAlaAspThrAsnIleAlaMetIleHisAlaAlaAspLysArgVal
 HisSerIleArgGluAlaTyrLeuProGluLeuSerValIleProGlyValAsnAlaAlaIlePheGluGlu
 LeuGluGlyArgIlePheThrAlaPheSerLeuTyrAspAlaArgAsnValIleLysAsnGlyAspPheAsn
 25 AsnGlyLeuSerCysTrpAsnValLysGlyHisValAspValGluGluGlnAsnAsnGlnArgSerValLeu
 ValValProGluTrpGluAlaGluValSerGlnGluValArgValCysProGlyArgGlyTyrIleLeuArg
 ValThrAlaTyrLysGluGlyTyrGlyGluGlyCysValThrIleHisGluIleGluAsnAsnThrAspGlu
 LeuLysPheSerAsnCysValGluGluGluIleTyrProAsnAsnThrValThrCysAsnAspTyrThrVal
 AsnGlnGluGluTyrGlyGlyAlaTyrThrSerArgAsnArgGlyTyrAsnGluAlaProSerValProAla
 30 AspTyrAlaSerValTyrGluGluLysSerTyrThrAspGlyArgArgGluAsnProCysGluPheAsnArg
 GlyTyrArgAspTyrThrProLeuProValGlyTyrValThrLysGluLeuGluTyrPheProGluThrAsp
 LysValTrpIleGluIleGlyGluThrGluGlyThrPheIleValAspSerValGluLeuLeuLeuMetGlu
 Glu

6.7 EXAMPLE 7 -- DNA SEQUENCES ENCODING THE NOVEL CRYSTAL PROTEINS

6.7.1 DNA SEQUENCE ENCODING THE EG11063 CRYSTAL PROTEIN (SEQ ID NO:9)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
5	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
10	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
15	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
20	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
25	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
30	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
35	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584

	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
5	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
10	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
15	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
20	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
25	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
30	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
35	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312

	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
5	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3531

6.7.2 DNA SEQUENCE ENCODING THE EG11074 CRYSTAL PROTEIN (SEQ ID NO:11)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
10	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
15	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
20	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
25	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
30	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
35	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344

	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
5	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
10	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA CTC GAG GCT GAA TAT AAT CTG GAA AGA GCG CAG AAG GCG GTG	1872
	AAT GCG CTG TTT ACG TCT ACA AAC CAA CTA GGG CTA AAA ACA AAT GTA	1920
	ACG GAT TAT CAT ATT GAT CAA GTG TCC AAT TTA GTT ACG TAT TTA TCG	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
15	CAT GCG AAG CGA CTC AGT GAT GAA CGC AAT TTA CTC CAA GAT TCA AAT	2064
	TTC AAA GAC ATT AAT AGG CAA CCA GAA CGT GGG TGG GGC GGA AGT ACA	2112
	GGG ATT ACC ATC CAA GGA GGG GAT GAC GTA TTT AAA GAA AAT TAC GTC	2160
	ACA CTA TCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
20	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
25	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
30	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
35	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072

	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
5	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
10	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3531

6.7.3 DNA SEQUENCE ENCODING THE EG11735 CRYSTAL PROTEIN (SEQ ID NO:13)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
15	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
20	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
25	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT CAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAG CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG ATA AGA TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT TCT CTA TTT CCG AAC TAT GAT AGT AGA ACG TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
30	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	GGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGA GGA GAA TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
35	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104

	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
5	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
10	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
15	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
20	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
25	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
30	GAT GGA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
35	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832

	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
5	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
10	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
15	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3531

6.7.4 DNA SEQUENCE ENCODING THE EG11092 CRYSTAL PROTEIN (SEQ ID NO:25)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
20	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
25	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
30	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT CAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAG CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG ATA AGA TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT TCT CTA TTT CCG AAC TAT GAT AGT AGA ACG TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
35	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864

	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
5	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
10	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
15	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
20	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
25	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
30	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
35	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592

	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
5	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
10	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
15	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
20	AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG	3534

6.7.5 DNA SEQUENCE ENCODING THE EG11751 CRYSTAL PROTEIN (SEQ ID NO:27)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
25	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
30	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
35	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624

	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
5	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
10	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
15	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCT GAA TTT AAT AAT	1392
	ATA ATT GCA TCG GAT AGT ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
20	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
25	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
30	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
35	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352

	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
5	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
10	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
15	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
20	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
25	AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG	3534

6.7.6 DNA SEQUENCE ENCODING THE EG11091 CRYSTAL PROTEIN (SEQ ID NO:29)

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
30	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
35	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384

	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
5	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
10	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
15	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
20	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCA ACT CTT ACA AAT	1392
	ACA ATT GAT CCA GAG AGA ATT AAT CAA ATA CCT TTA GTG AAA GGA TTT	1440
	AGA GTT TGG GGG GGC ACC TCT GTC ATT ACA GGA CCA GGA TTT ACA GGA	1488
	GGG GAT ATC CTT CGA AGA AAT ACC TTT GGT GAT TTT GTA TCT CTA CAA	1536
25	GTC AAT ATT AAT TCA CCA ATT ACC CAA AGA TAC CGT TTA AGA TTT CGT	1584
	TAC GCT TCC AGT AGG GAT GCA CGA GTT ATA GTA TTA ACA GGA GCG GCA	1632
	TCC ACA GGA GTG GGA GGC CAA GTT AGT GTA AAT ATG CCT CTT CAG AAA	1680
	ACT ATG GAA ATA GGG GAG AAC TTA ACA TCT AGA ACA TTT AGA TAT ACC	1728
	GAT TTT AGT AAT CCT TTT TCA TTT AGA GCT AAT CCA GAT ATA ATT GGG	1776
30	ATA AGT GAA CAA CCT CTA TTT GGT GCA GGT TCT ATT AGT AGC GGT GAA	1824
	CTT TAT ATA GAT AAA ATT GAA ATT ATT CTA GCA GAT GCA ACA TTT GAA	1872
	GCA GAA TCT GAT TTA GAA AGA GCA CAA AAG GCG GTG AAT GCC CTG TTT	1920
	ACT TCT TCC AAT CAA ATC GGG TTA AAA ACC GAT GTG ACG GAT TAT CAT	1968
	ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA GAT GAA TTT TGT	2016
35	CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA CAT GCG AAG CGA	2064
	CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC TTC AGA GGG ATC	2112

	AAT AGA CAA CCA GAC CGT GGC TGG AGA GGA AGT ACA GAT ATT ACC ATC	2160
	CAA GGA GGA GAT GAC GTA TTC AAA GAG AAT TAC GTC ACA CTA CCG GGT	2208
	ACC GTT GAT GAG TGC TAT CCA ACG TAT TTA TAT CAG AAA ATA GAT GAG	2256
	TCG AAA TTA AAA GCT TAT ACC CGT TAT GAA TTA AGA GGG TAT ATC GAA	2304
5	GAT AGT CAA GAC TTA GAA ATC TAT TTG ATC CGT TAC AAT GCA AAA CAC	2352
	GAA ATA GTA AAT GTG CCA GGC ACG GGT TCC TTA TGG CCG CTT TCA GCC	2400
	CAA AGT CCA ATC GGA AAG TGT GGA GAA CCG AAT CGA TGC GCG CCA CAC	2448
	CTT GAA TGG AAT CCT GAT CTA GAT TGT TCC TGC AGA GAC GGG GAA AAA	2496
	TGT GCA CAT CAT TCC CAT CAT TTC ACC TTG GAT ATT GAT GTT GGA TGT	2544
10	ACA GAC TTA AAT GAG GAC TTA GGT GTA TGG GTG ATA TTC AAG ATT AAG	2592
	ACG CAA GAT GGC CAT GCA AGA CTA GGG AAT CTA GAG TTT CTC GAA GAG	2640
	AAA CCA TTA TTA GGG GAA GCA CTA GCT CGT GTG AAA AGA GCG GAG AAG	2688
	AAG TGG AGA GAC AAA CGA GAG AAA CTG CAG TTG GAA ACA AAT ATT GTT	2736
	TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT GTA AAC TCT CAA	2784
15	TAT GAT AGA TTA CAA GTG GAT ACG AAC ATC GCA ATG ATT CAT GCG GCA	2832
	GAT AAA CGC GTT CAT AGA ATC CGG GAA GCG TAT CTG CCA GAG TTG TCT	2880
	GTG ATT CCA GGT GTC AAT GCG GCC ATT TTC GAA GAA TTA GAG GGA CGT	2928
	ATT TTT ACA GCG TAT TCC TTA TAT GAT GCG AGA AAT GTC ATT AAA AAT	2976
	GGC GAT TTC AAT AAT GGC TTA TTA TGC TGG AAC GTG AAA GGT CAT GTA	3024
20	GAT GTA GAA GAG CAA AAC AAC CAC CGT TCG GTC CTT GTT ATC CCA GAA	3072
	TGG GAG GCA GAA GTG TCA CAA GAG GTT CGT GTC TGT CCA GGT CGT GGC	3120
	TAT ATC CTT CGT GTC ACA GCA TAT AAA GAG GGA TAT GGA GAG GGC TGC	3168
	GTA ACG ATC CAT GAG ATC GAA GAC AAT ACA GAC GAA CTG AAA TTC AGC	3216
	AAC TGT GTA GAA GAG GAA GTA TAT CCA AAC AAC ACA GTA ACG TGT AAT	3264
25	AAT TAT ACT GGG ACT CAA GAA GAA TAT GAG GGT ACG TAC ACT TCT CGT	3312
	AAT CAA GGA TAT GAC GAA GCC TAT GGT AAT AAC CCT TCC GTA CCA GCT	3360
	GAT TAC GCT TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3408
	GAG AAT CCT TGT GAA TCT AAC AGA GGC TAT GGG GAT TAC ACA CCA CTA	3456
	CCG GCT GGT TAT GTA ACA AAG GAT TTA GAG TAC TTC CCA GAG ACC GAT	3504
30	AAG GTA TGG ATT GAG ATC GGA GAA ACA GAA GGA ACA TTC ATC GTG GAT	3552
	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3579

6.7.7 DNA SEQUENCE ENCODING THE EG11768 CRYSTAL PROTEIN (SEQ ID NO:33)

35	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
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	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
5	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
10	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
15	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
20	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
25	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCT GAA TTT AAT AAT	1392
	ATA ATT GCA TCG GAT AGT ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
30	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
35	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776

	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	GCA ACA CTC GAG GCT GAA TAT AAT CTG GAA AGA GCG CAG AAG GCG GTG	1872
	AAT GCG CTG TTT ACG TCT ACA AAC CAA CTA GGG CTA AAA ACA AAT GTA	1920
	ACG GAT TAT CAT ATT GAT CAA GTG TCC AAT TTA GTT ACG TAT TTA TCG	1968
5	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	CAT GCG AAG CGA CTC AGT GAT GAA CGC AAT TTA CTC CAA GAT TCA AAT	2064
	TTC AAA GAC ATT AAT AGG CAA CCA GAA CGT GGG TGG GGC GGA AGT ACA	2112
	GGG ATT ACC ATC CAA GGA GGG GAT GAC GTA TTT AAA GAA AAT TAC GTC	2160
	ACA CTA TCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
10	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
15	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
20	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
25	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
30	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
35	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504

7. REFERENCES

5 The following references, to the extent that they provide exemplary procedural or other details supplementary to those set forth herein, are specifically incorporated herein by reference.

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- U. S. Patent 4,683,195.
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- U. S. Patent 4,702,914.
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- U. S. Patent 4,940,835.
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- U. S. Patent 4,971,908.
- U. S. Patent 5,004,863.
- U. S. Patent 5,015,580.
- U. S. Patent 5,055,294.
- 20 U. S. Patent 5,128,130.
- U. S. Patent 5,176,995.
- U. S. Patent 5,349,124.
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8. SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Malvar, Thomas
Gilmer, Amy Jelen

(ii) TITLE OF INVENTION: BROAD-SPECTRUM DELTA-ENDOTOXINS

(iii) NUMBER OF SEQUENCES: 35

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: Arnold, White & Durkee
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(C) CITY: Houston
(D) STATE: Texas
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(F) ZIP: 77210

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER: US Unknown
(B) FILING DATE: Concurrently Herewith
(C) CLASSIFICATION: Unknown

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: US 08/754/490
(B) FILING DATE: 20-NOV-1996

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Kitchell, Barbara S.
(B) REGISTRATION NUMBER: 33,928
(C) REFERENCE/DOCKET NUMBER: MOBT:163

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: 512/418-3000
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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GGATAGCACT CATCAAAGGT ACC 23

5

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

10 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

15 GAAGATATCC AATTCGAACA GTTCC 27

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 28 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CATATTCTGC CTCGAGTGTT GCAGTAAC 28

30

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs

35 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

40 CCCGATCGGC CGCATGC 17

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17 base pairs

(B) TYPE: nucleic acid

50 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CATTGGAGCT CTCCATG 17

5

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16 base pairs

10 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

15 GCACTACGAT GTATCC 16

(2) INFORMATION FOR SEQ ID NO:7:

20 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CATCGTAGTG CAACTCTTAC 20

30

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

35 (A) LENGTH: 39 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

40 CCAAGAAAAT ACTAGAGCTC TTGTTAAAAA AGGTGTTCC 39

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3531 base pairs

50 (B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..3531

5 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu	
	1 5 10 15	
10	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
	Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly	
	20 25 30	
15	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
	Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser	
	35 40 45	
20	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile	
	50 55 60	
25	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile	
	65 70 75 80	
	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	
	85 90 95	
30	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
35	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
40	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	
45	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	
	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	
	165 170 175	
50	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	
	180 185 190	

	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	
	195 200 205	
5	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	
	210 215 220	
10	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225 230 235 240	
15	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	
	245 250 255	
	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260 265 270	
20	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275 280 285	
25	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290 295 300	
30	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	
	305 310 315 320	
35	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325 330 335	
40	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	
	340 345 350	
	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	
	355 360 365	
45	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	
	370 375 380	
50	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	
	385 390 395 400	

	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	
	405 410 415	
5	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	
	420 425 430	
10	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	
	435 440 445	
15	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn	
	450 455 460	
	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His	
	465 470 475 480	
20	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	
	485 490 495	
25	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	
	500 505 510	
30	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	
	515 520 525	
35	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	
	530 535 540	
40	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	
	545 550 555 560	
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	
	565 570 575	
45	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	
	580 585 590	
50	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	
	595 600 605	

	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val	
	610 615 620	
5	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val	
	625 630 635 640	
10	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser	
	645 650 655	
15	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys	
	660 665 670	
20	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn	
	675 680 685	
25	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr	
	690 695 700	
30	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val	
	705 710 715 720	
35	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln	
	725 730 735	
40	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg	
	740 745 750	
45	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr	
	755 760 765	
50	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	
	770 775 780	
55	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	
	785 790 795 800	
60	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg	
	805 810 815	

	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile	
	820 825 830	
5	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile	
	835 840 845	
10	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
15	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
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	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
25	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
30	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
35	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
40	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
45	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	
50	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
55	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
60	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	

Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30
 5 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60
 10 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80
 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
 85 90 95
 15 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
 100 105 110
 Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
 115 120 125
 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
 130 135 140
 25 Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
 145 150 155 160
 Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175
 30 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190
 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val
 195 200 205
 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220
 40 Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 225 230 235 240
 Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro
 245 250 255
 45 Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270
 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285
 50 Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300

Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320

5 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335

Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350

10 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 355 360 365

Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 15 370 375 380

Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400

20 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415

Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430

25 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile
 435 440 445

Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn
 30 450 455 460

Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His
 465 470 475 480

35 Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly
 485 490 495

Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile
 500 505 510

40 Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg
 515 520 525

Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu
 45 530 535 540

Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro
 545 550 555 560

50 Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr
 565 570 575

Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser
 580 585 590

Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr
 595 600 605
 5 Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val
 610 615 620
 Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val
 625 630 635 640
 10 Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser
 645 650 655
 Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys
 660 665 670
 15 His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn
 675 680 685
 20 Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr
 690 695 700
 Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val
 705 710 715 720
 25 Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln
 725 730 735
 Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg
 740 745 750
 30 Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr
 755 760 765
 35 Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp
 770 775 780
 Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg
 785 790 795 800
 40 Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg
 805 810 815
 Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile
 820 825 830
 45 Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile
 835 840 845
 50 Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu
 850 855 860
 Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys
 865 870 875 880

Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu
 885 890 895
 5 Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe
 900 905 910
 Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met
 915 920 925
 10 Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu
 930 935 940
 Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu
 15 945 950 955 960
 Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn
 965 970 975
 20 Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val
 980 985 990
 Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu
 995 1000 1005
 25 Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys
 1010 1015 1020
 Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr
 30 1025 1030 1035 1040
 Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu
 1045 1050 1055
 35 Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr
 1060 1065 1070
 Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala
 1075 1080 1085
 40 Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100
 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 45 1105 1110 1115 1120
 Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135
 50 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

Ser Val Glu Leu Leu Leu Met Glu Glu
1170 1175

5

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

10

- (A) LENGTH: 3531 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

15

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..3531

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

20	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu 1 5 10 15	48
25	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly 20 25 30	96
30	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser 35 40 45	144
35	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile 50 55 60	192
40	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile 65 70 75 80	240
45	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala 85 90 95	288
50	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu 100 105 110	336
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu 115 120 125	384
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala 130 135 140	432

	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	
5	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	
	165 170 175	
10	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	
	180 185 190	
15	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	
	195 200 205	
20	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	
	210 215 220	
	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225 230 235 240	
25	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	
	245 250 255	
30	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260 265 270	
35	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275 280 285	
40	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290 295 300	
45	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	
	305 310 315 320	
	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325 330 335	
50	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	
	340 345 350	

	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg 355 360 365	1104
5	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp 370 375 380	1152
10	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val 385 390 395 400	1200
15	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln 405 410 415	1248
20	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His 420 425 430	1296
25	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile 435 440 445	1344
30	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn 450 455 460	1392
35	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His 465 470 475 480	1440
40	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly 485 490 495	1488
45	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile 500 505 510	1536
50	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg 515 520 525	1584
55	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu 530 535 540	1632
60	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro 545 550 555 560	1680

	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	
	565 570 575	
5	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	
	580 585 590	
10	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	
	595 600 605	
15	GCA ACA CTC GAG GCT GAA TAT AAT CTG GAA AGA GCG CAG AAG GCG GTG	1872
	Ala Thr Leu Glu Ala Glu Tyr Asn Leu Glu Arg Ala Gln Lys Ala Val	
	610 615 620	
20	AAT GCG CTG TTT ACG TCT ACA AAC CAA CTA GGG CTA AAA ACA AAT GTA	1920
	Asn Ala Leu Phe Thr Ser Thr Asn Gln Leu Gly Leu Lys Thr Asn Val	
	625 630 635 640	
25	ACG GAT TAT CAT ATT GAT CAA GTG TCC AAT TTA GTT ACG TAT TTA TCG	1968
	Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Thr Tyr Leu Ser	
	645 650 655	
30	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys	
	660 665 670	
35	CAT GCG AAG CGA CTC AGT GAT GAA CGC AAT TTA CTC CAA GAT TCA AAT	2064
	His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Ser Asn	
	675 680 685	
40	TTC AAA GAC ATT AAT AGG CAA CCA GAA CGT GGG TGG GGC GGA AGT ACA	2112
	Phe Lys Asp Ile Asn Arg Gln Pro Glu Arg Gly Trp Gly Gly Ser Thr	
	690 695 700	
45	GGG ATT ACC ATC CAA GGA GGG GAT GAC GTA TTT AAA GAA AAT TAC GTC	2160
	Gly Ile Thr Ile Gln Gly Gly Asp Asp Val Phe Lys Glu Asn Tyr Val	
	705 710 715 720	
50	ACA CTA TCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	Thr Leu Ser Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln	
	725 730 735	
55	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg	
	740 745 750	
60	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr	
	755 760 765	

	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	
	770 775 780	
5	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	
	785 790 795 800	
10	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg	
	805 810 815	
15	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile	
	820 825 830	
20	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile	
	835 840 845	
25	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
30	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
35	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
40	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
45	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
50	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
55	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
60	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	

	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
5	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
10	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	
15	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025 1030 1035 1040	
20	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
	1045 1050 1055	
25	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	Leu Lys Phe Ser Asn Cys Val Glu Glu Ile Tyr Pro Asn Asn Thr	
	1060 1065 1070	
30	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala	
	1075 1080 1085	
35	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala	
	1090 1095 1100	
40	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg	
	1105 1110 1115 1120	
45	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu	
	1125 1130 1135	
50	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp	
	1140 1145 1150	
55	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp	
	1155 1160 1165	
60	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3531
	Ser Val Glu Leu Leu Leu Met Glu Glu	
	1170 1175	

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 1177 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
1 5 10 15
15 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
20 25 30
Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
35 40 45
20 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
50 55 60
Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
25 65 70 75 80
Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
85 90 95
30 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
100 105 110
Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
115 120 125
35 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
130 135 140
Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
40 145 150 155 160
Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
165 170 175
45 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
180 185 190
Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val
195 200 205
50 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
210 215 220

	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	225	230	235	240
5	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	245	250	255	
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	260	265	270	
10	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	275	280	285	
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	290	295	300	
15	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	305	310	315	320
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	325	330	335	
20	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	340	345	350	
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	355	360	365	
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	370	375	380	
30	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	385	390	395	400
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	405	410	415	
35	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	420	425	430	
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	435	440	445	
	Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn	450	455	460	
45	Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His	465	470	475	480
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	485	490	495	
50	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	500	505	510	

	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	
	515	520 525
5	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	
	530	535 540
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	
	545	550 555 560
10	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	
		565 570 575
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	
		580 585 590
15	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	
		595 600 605
20	Ala Thr Leu Glu Ala Glu Tyr Asn Leu Glu Arg Ala Gln Lys Ala Val	
		610 615 620
	Asn Ala Leu Phe Thr Ser Thr Asn Gln Leu Gly Leu Lys Thr Asn Val	
		625 630 635 640
25	Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Thr Tyr Leu Ser	
		645 650 655
	Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys	
		660 665 670
30	His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Ser Asn	
		675 680 685
	Phe Lys Asp Ile Asn Arg Gln Pro Glu Arg Gly Trp Gly Gly Ser Thr	
		690 695 700
35	Gly Ile Thr Ile Gln Gly Gly Asp Asp Val Phe Lys Glu Asn Tyr Val	
		705 710 715 720
40	Thr Leu Ser Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln	
		725 730 735
	Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg	
		740 745 750
45	Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr	
		755 760 765
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	
		770 775 780
50	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	
		785 790 795 800

	Cys	Ala	Pro	His	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg	
					805					810					815		
5	Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile	
				820					825					830			
	Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile	
			835					840					845				
10	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	
		850					855					860					
	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	
15		865				870					875				880		
	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu	
				885					890					895			
20	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	
			900						905					910			
	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met	
			915				920						925				
25	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu	
		930					935					940					
	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	
30		945				950				955					960		
	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	
				965					970					975			
35	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val	
			980					985						990			
	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu	
			995					1000					1005				
40	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	
		1010					1015					1020					
	Pro	Gly	Arg	Gly	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	Tyr	
45		1025				1030				1035					1040		
	Gly	Glu	Gly	Cys	Val	Thr	Ile	His	Glu	Ile	Glu	Asn	Asn	Thr	Asp	Glu	
				1045					1050					1055			
50	Leu	Lys	Phe	Ser	Asn	Cys	Val	Glu	Glu	Glu	Ile	Tyr	Pro	Asn	Asn	Thr	
			1060						1065					1070			
	Val	Thr	Cys	Asn	Asp	Tyr	Thr	Val	Asn	Gln	Glu	Glu	Tyr	Gly	Gly	Ala	
			1075					1080						1085			

Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100

5 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1105 1110 1115 1120

Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135

10 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150

Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

15 Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

20 (2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3531 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

25 (ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 1..3531

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

35 ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA 48
 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15

40 AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT 96
 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30

45 TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT 144
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45

GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA 192
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60

50 TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT 240
 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80

	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	
	85 90 95	
5	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
10	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
15	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	
20	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	
25	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	
	165 170 175	
30	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	
	180 185 190	
35	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT CAT GCT GTA	624
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val	
	195 200 205	
40	CGC TGG TAC AAT ACG GGA TTA GAG CGT GTA TGG GGA CCG GAT TCT AGA	672
	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	
	210 215 220	
45	GAT TGG ATA AGA TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225 230 235 240	
50	TTA GAT ATC GTT TCT CTA TTT CCG AAC TAT GAT AGT AGA ACG TAT CCA	768
	Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro	
	245 250 255	
55	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260 265 270	
60	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275 280 285	

	GGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	Gly Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290 295 300	
5	ATC TAT ACG GAT GCT CAT AGA GGA GAA TAT TAT TGG TCA GGG CAT CAA	960
	Ile Tyr Thr Asp Ala His Arg Gly Glu Tyr Tyr Trp Ser Gly His Gln	
	305 310 315 320	
10	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325 330 335	
15	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	
	340 345 350	
20	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	
	355 360 365	
25	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	
	370 375 380	
30	GGA ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	
	385 390 395 400	
35	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	
	405 410 415	
40	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	
	420 425 430	
45	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	
	435 440 445	
50	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn	
	450 455 460	
55	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His	
	465 470 475 480	
60	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	
	485 490 495	

	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	
	500 505 510	
5	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	
	515 520 525	
10	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	
	530 535 540	
15	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	
	545 550 555 560	
	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	
	565 570 575	
20	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	
	580 585 590	
25	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	
	595 600 605	
30	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val	
	610 615 620	
35	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val	
	625 630 635 640	
40	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser	
	645 650 655	
	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys	
	660 665 670	
45	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn	
	675 680 685	
50	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr	
	690 695 700	

	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val	
	705 710 715 720	
5	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln	
	725 730 735	
10	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg	
	740 745 750	
15	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr	
	755 760 765	
20	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	
	770 775 780	
25	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	
	785 790 795 800	
30	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg	
	805 810 815	
35	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile	
	820 825 830	
40	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile	
	835 840 845	
45	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
50	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
55	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
60	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
65	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	

	915	920	925	
5	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu 930 935 940	2832		
10	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu 945 950 955 960	2880		
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn 965 970 975	2928		
15	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val 980 985 990	2976		
20	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu 995 1000 1005	3024		
25	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys 1010 1015 1020	3072		
30	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr 1025 1030 1035 1040	3120		
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu 1045 1050 1055	3168		
35	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr 1060 1065 1070	3216		
40	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala 1075 1080 1085	3264		
45	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala 1090 1095 1100	3312		
50	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg 1105 1110 1115 1120	3360		
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu 1125 1130 1135	3408		

	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp	
	1140 1145 1150	
5	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp	
	1155 1160 1165	
10	AGC GTG GAA TTA CTC CTT ATG GAG GAA	3531
	Ser Val Glu Leu Leu Leu Met Glu Glu	
	1170 1175	

(2) INFORMATION FOR SEQ ID NO:14:

15

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1177 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

20

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

25	Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu	
	1 5 10 15	
	Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly	
	20 25 30	
30	Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser	
	35 40 45	
	Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile	
35	50 55 60	
	Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile	
	65 70 75 80	
40	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	
	85 90 95	
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
45	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
50	130 135 140	
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	

Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
 165 170 175
 5 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
 180 185 190
 Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val
 195 200 205
 10 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
 210 215 220
 Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
 225 230 235 240
 15 Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro
 245 250 255
 20 Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val
 260 265 270
 Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu
 275 280 285
 25 Gly Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr
 290 295 300
 Ile Tyr Thr Asp Ala His Arg Gly Glu Tyr Tyr Trp Ser Gly His Gln
 305 310 315 320
 30 Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro
 325 330 335
 35 Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala
 340 345 350
 Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg
 355 360 365
 40 Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp
 370 375 380
 Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val
 385 390 395 400
 45 Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln
 405 410 415
 50 Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His
 420 425 430
 Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile
 435 440 445

	Arg	Ala	Pro	Met	Phe	Ser	Trp	Thr	His	Arg	Ser	Ala	Thr	Pro	Thr	Asn	
	450						455					460					
5	Thr	Ile	Asp	Pro	Glu	Arg	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
	465					470					475					480	
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485					490					495		
10	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505						510		
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
				515				520					525				
15	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	
	530						535					540					
20	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
	545					550					555					560	
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
				565						570					575		
25	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	
				580					585					590			
	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	
			595					600					605				
30	Ala	Thr	Phe	Glu	Ala	Glu	Tyr	Asp	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val	
	610						615					620					
35	Asn	Ala	Leu	Phe	Thr	Ser	Ile	Asn	Gln	Ile	Gly	Ile	Lys	Thr	Asp	Val	
	625					630					635					640	
	Thr	Asp	Tyr	His	Ile	Asp	Gln	Val	Ser	Asn	Leu	Val	Asp	Cys	Leu	Ser	
				645					650					655			
40	Asp	Glu	Phe	Cys	Leu	Asp	Glu	Lys	Arg	Glu	Leu	Ser	Glu	Lys	Val	Lys	
				660					665					670			
	His	Ala	Lys	Arg	Leu	Ser	Asp	Glu	Arg	Asn	Leu	Leu	Gln	Asp	Pro	Asn	
			675					680					685				
45	Phe	Lys	Gly	Ile	Asn	Arg	Gln	Leu	Asp	Arg	Gly	Trp	Arg	Gly	Ser	Thr	
	690						695					700					
50	Asp	Ile	Thr	Ile	Gln	Arg	Gly	Asp	Asp	Val	Phe	Lys	Glu	Asn	Tyr	Val	
	705				710					715					720		
	Thr	Leu	Pro	Gly	Thr	Phe	Asp	Glu	Cys	Tyr	Pro	Thr	Tyr	Leu	Tyr	Gln	
				725						730					735		

Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg
 740 745 750
 5 Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr
 755 760 765
 Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp
 770 775 780
 10 Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg
 785 790 795 800
 Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg
 805 810 815
 15 Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile
 820 825 830
 Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile
 835 840 845
 20 Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu
 850 855 860
 25 Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys
 865 870 875 880
 Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu
 885 890 895
 30 Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe
 900 905 910
 Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met
 915 920 925
 35 Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu
 930 935 940
 40 Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu
 945 950 955 960
 Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn
 965 970 975
 45 Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val
 980 985 990
 Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu
 995 1000 1005
 50 Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys
 1010 1015 1020

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

TTTAGTCATC GATTAAATCA

20

5

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

15

ATAATAAGAG CTCCAATGTT

20

(2) INFORMATION FOR SEQ ID NO:18:

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

TACATCGTAG TGCAACTCTT

20

30

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

35

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

40

TCATGGAGAG CTCCTATGTT

20

45

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

50

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

TTAACAAGAG CTCCTATGTT

20

5

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

10

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

15

ACTACCAGGT ACCTTTGATG

20

(2) INFORMATION FOR SEQ ID NO:22:

20

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

ACTACCGGGT ACCTTTGATA

20

30

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

35

(A) LENGTH: 18 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

40

ATTTGAGTAA TACTATCC

18

45

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

50

(A) LENGTH: 19 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

ATTACTCAAA TACCATTGG

19

5

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:

10

- (A) LENGTH: 3534 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ix) FEATURE:

15

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..3531

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

20	ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA	48
	Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu	
	1 5 10 15	
	AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT	96
25	Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly	
	20 25 30	
	TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT	144
30	Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser	
	35 40 45	
	GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA	192
	Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile	
35	50 55 60	
	TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT	240
	Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile	
	65 70 75 80	
40	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	
	85 90 95	
	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
45	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
50	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	

5	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val 145 150 155 160	480
10	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser 165 170 175	528
15	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg 180 185 190	576
20	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT CAT GCT GTA Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val 195 200 205	624
25	CGC TGG TAC AAT ACG GGA TTA GAG CGT GTA TGG GGA CCG GAT TCT AGA Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg 210 215 220	672
30	GAT TGG ATA AGA TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val 225 230 235 240	720
35	TTA GAT ATC GTT TCT CTA TTT CCG AAC TAT GAT AGT AGA ACG TAT CCA Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro 245 250 255	768
40	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val 260 265 270	816
45	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu 275 280 285	864
50	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr 290 295 300	912
55	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln 305 310 315 320	960
60	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro 325 330 335	1008
65	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala 340 345 350	1056

	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	
	355 360 365	
5	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	
	370 375 380	
10	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	
	385 390 395 400	
15	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	
	405 410 415	
20	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	
	420 425 430	
25	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	
	435 440 445	
30	AGA GCT CCA ATG TTT TCT TGG ACG CAC CGT AGT GCA ACC CCT ACA AAT	1392
	Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn	
	450 455 460	
35	ACA ATT GAT CCG GAG AGG ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His	
	465 470 475 480	
40	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	
	485 490 495	
45	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	
	500 505 510	
50	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	
	515 520 525	
55	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	
	530 535 540	
60	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	
	545 550 555 560	

	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr 565 570 575	1728
5	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser 580 585 590	1776
10	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr 595 600 605	1824
15	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val 610 615 620	1872
20	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val 625 630 635 640	1920
25	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser 645 650 655	1968
30	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys 660 665 670	2016
35	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn 675 680 685	2064
40	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr 690 695 700	2112
45	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val 705 710 715 720	2160
50	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln 725 730 735	2208
55	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg 740 745 750	2256
60	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr 755 760 765	2304

	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	
	770 775 780	
5	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	
	785 790 795 800	
10	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg	
	805 810 815	
15	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile	
	820 825 830	
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile	
	835 840 845	
20	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
25	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
30	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
35	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
40	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	
	915 920 925	
45	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG	2832
	Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu	
	930 935 940	
50	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA	2880
	Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu	
	945 950 955 960	
50	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT	2928
	Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn	
	965 970 975	

	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG	2976
	Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val	
	980 985 990	
5	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT	3024
	Lys Gly His Val Asp Val Glu Gln Asn Asn Gln Arg Ser Val Leu	
	995 1000 1005	
10	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT	3072
	Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys	
	1010 1015 1020	
15	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT	3120
	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025 1030 1035 1040	
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA	3168
	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
	1045 1050 1055	
20	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG	3216
	Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr	
	1060 1065 1070	
25	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG	3264
	Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala	
	1075 1080 1085	
30	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT	3312
	Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala	
	1090 1095 1100	
35	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA	3360
	Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg	
	1105 1110 1115 1120	
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA	3408
	Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu	
	1125 1130 1135	
40	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp	
	1140 1145 1150	
45	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp	
	1155 1160 1165	
50	AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG	3534
	Ser Val Glu Leu Leu Leu Met Glu Glu	
	1170 1175	

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 1177 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
1 5 10 15
15 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
20 25 30
Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
35 40 45
20 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
50 55 60
Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
25 65 70 75 80
Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
85 90 95
30 Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
100 105 110
Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
115 120 125
35 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
130 135 140
Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
40 145 150 155 160
Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
165 170 175
45 Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
180 185 190
Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp His Ala Val
195 200 205
50 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
210 215 220

	Asp Trp Ile Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225	235 240
5	Leu Asp Ile Val Ser Leu Phe Pro Asn Tyr Asp Ser Arg Thr Tyr Pro	
	245	250 255
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260	265 270
10	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275	280 285
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290	295 300
15	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	
	305	310 315 320
20	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325	330 335
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	
	340	345 350
25	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	
	355	360 365
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	
	370	375 380
30	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	
	385	390 395 400
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	
35	405	410 415
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	
	420	425 430
40	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	
	435	440 445
	Arg Ala Pro Met Phe Ser Trp Thr His Arg Ser Ala Thr Pro Thr Asn	
	450	455 460
45	Thr Ile Asp Pro Glu Arg Ile Thr Gln Ile Pro Leu Val Lys Ala His	
	465	470 475 480
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	
50	485	490 495
	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	
	500	505 510

	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	515	520	525
5	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	530	535	540
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	545	550	555
10	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	565	570	575
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	580	585	590
15	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	595	600	605
	Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val	610	615	620
20	Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val	625	630	635
	Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser	645	650	655
	Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys	660	665	670
30	His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn	675	680	685
	Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr	690	695	700
35	Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val	705	710	715
	Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln	725	730	735
	Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg	740	745	750
45	Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr	755	760	765
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	770	775	780
50	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	785	790	795
				800

	Cys	Ala	Pro	His	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg	
					805					810					815		
5	Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile	
				820					825					830			
	Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile	
			835					840					845				
10	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	
		850				855						860					
	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	
		865				870					875					880	
15	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu	
				885					890					895			
	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	
20				900					905					910			
	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met	
		915					920						925				
25	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu	
		930				935						940					
	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	
		945				950				955					960		
30	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	
				965					970					975			
	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val	
35			980					985						990			
	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu	
		995					1000					1005					
40	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	
		1010				1015						1020					
	Pro	Gly	Arg	Gly	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	Tyr	
		1025			1030					1035					1040		
45	Gly	Glu	Gly	Cys	Val	Thr	Ile	His	Glu	Ile	Glu	Asn	Asn	Thr	Asp	Glu	
				1045					1050					1055			
	Leu	Lys	Phe	Ser	Asn	Cys	Val	Glu	Glu	Glu	Ile	Tyr	Pro	Asn	Asn	Thr	
50				1060				1065					1070				
	Val	Thr	Cys	Asn	Asp	Tyr	Thr	Val	Asn	Gln	Glu	Glu	Tyr	Gly	Gly	Ala	
		1075					1080						1085				

Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
 1090 1095 1100

5 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1105 1110 1115 1120

Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
 1125 1130 1135

10 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
 1140 1145 1150

Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1155 1160 1165

15 Ser Val Glu Leu Leu Leu Met Glu Glu
 1170 1175

20 (2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 3534 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

25 (ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION: 1..3531

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

35 ATG GAT AAC AAT CCG AAC ATC AAT GAA TGC ATT CCT TAT AAT TGT TTA 48
 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
 1 5 10 15

40 AGT AAC CCT GAA GTA GAA GTA TTA GGT GGA GAA AGA ATA GAA ACT GGT 96
 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
 20 25 30

45 TAC ACC CCA ATC GAT ATT TCC TTG TCG CTA ACG CAA TTT CTT TTG AGT 144
 Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
 35 40 45

GAA TTT GTT CCC GGT GCT GGA TTT GTG TTA GGA CTA GTT GAT ATA ATA 192
 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
 50 55 60

50 TGG GGA ATT TTT GGT CCC TCT CAA TGG GAC GCA TTT CTT GTA CAA ATT 240
 Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
 65 70 75 80

	GAA CAG TTA ATT AAC CAA AGA ATA GAA GAA TTC GCT AGG AAC CAA GCC	288
	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	
	85 90 95	
5	ATT TCT AGA TTA GAA GGA CTA AGC AAT CTT TAT CAA ATT TAC GCA GAA	336
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
10	TCT TTT AGA GAG TGG GAA GCA GAT CCT ACT AAT CCA GCA TTA AGA GAA	384
	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
15	GAG ATG CGT ATT CAA TTC AAT GAC ATG AAC AGT GCC CTT ACA ACC GCT	432
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
	130 135 140	
20	ATT CCT CTT TTT GCA GTT CAA AAT TAT CAA GTT CCT CTT TTA TCA GTA	480
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	
25	TAT GTT CAA GCT GCA AAT TTA CAT TTA TCA GTT TTG AGA GAT GTT TCA	528
	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	
	165 170 175	
30	GTG TTT GGA CAA AGG TGG GGA TTT GAT GCC GCG ACT ATC AAT AGT CGT	576
	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	
	180 185 190	
35	TAT AAT GAT TTA ACT AGG CTT ATT GGC AAC TAT ACA GAT TAT GCT GTA	624
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	
	195 200 205	
40	CGC TGG TAC AAT ACG GGA TTA GAA CGT GTA TGG GGA CCG GAT TCT AGA	672
	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	
	210 215 220	
45	GAT TGG GTA AGG TAT AAT CAA TTT AGA AGA GAA TTA ACA CTA ACT GTA	720
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	
	225 230 235 240	
50	TTA GAT ATC GTT GCT CTG TTC CCG AAT TAT GAT AGT AGA AGA TAT CCA	768
	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	
	245 250 255	
55	ATT CGA ACA GTT TCC CAA TTA ACA AGA GAA ATT TAT ACA AAC CCA GTA	816
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	
	260 265 270	
60	TTA GAA AAT TTT GAT GGT AGT TTT CGA GGC TCG GCT CAG GGC ATA GAA	864
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	
	275 280 285	

	AGA AGT ATT AGG AGT CCA CAT TTG ATG GAT ATA CTT AAC AGT ATA ACC	912
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	
	290 295 300	
5	ATC TAT ACG GAT GCT CAT AGG GGT TAT TAT TAT TGG TCA GGG CAT CAA	960
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	
	305 310 315 320	
10	ATA ATG GCT TCT CCT GTA GGG TTT TCG GGG CCA GAA TTC ACT TTT CCG	1008
	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	
	325 330 335	
15	CTA TAT GGA ACT ATG GGA AAT GCA GCT CCA CAA CAA CGT ATT GTT GCT	1056
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	
	340 345 350	
20	CAA CTA GGT CAG GGC GTG TAT AGA ACA TTA TCG TCC ACT TTA TAT AGA	1104
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	
	355 360 365	
25	AGA CCT TTT AAT ATA GGG ATA AAT AAT CAA CAA CTA TCT GTT CTT GAC	1152
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	
	370 375 380	
30	GGG ACA GAA TTT GCT TAT GGA ACC TCC TCA AAT TTG CCA TCC GCT GTA	1200
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	
	385 390 395 400	
35	TAC AGA AAA AGC GGA ACG GTA GAT TCG CTG GAT GAA ATA CCG CCA CAG	1248
	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	
	405 410 415	
40	AAT AAC AAC GTG CCA CCT AGG CAA GGA TTT AGT CAT CGA TTA AGC CAT	1296
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	
	420 425 430	
45	GTT TCA ATG TTT CGT TCA GGC TTT AGT AAT AGT AGT GTA AGT ATA ATA	1344
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	
	435 440 445	
50	AGA GCT CCT ATG TTC TCT TGG ATA CAT CGT AGT GCT GAA TTT AAT AAT	1392
	Arg Ala Pro Met Phe Ser Trp Ile His Arg Ser Ala Glu Phe Asn Asn	
	450 455 460	
55	ATA ATT GCA TCG GAT AGT ATT ACT CAA ATA CCA TTG GTA AAA GCA CAT	1440
	Ile Ile Ala Ser Asp Ser Ile Thr Gln Ile Pro Leu Val Lys Ala His	
	465 470 475 480	
60	ACA CTT CAG TCA GGT ACT ACT GTT GTA AGA GGG CCC GGG TTT ACG GGA	1488
	Thr Leu Gln Ser Gly Thr Thr Val Val Arg Gly Pro Gly Phe Thr Gly	
	485 490 495	

	GGA GAT ATT CTT CGA CGA ACA AGT GGA GGA CCA TTT GCT TAT ACT ATT	1536
	Gly Asp Ile Leu Arg Arg Thr Ser Gly Gly Pro Phe Ala Tyr Thr Ile	
	500 505 510	
5	GTT AAT ATA AAT GGG CAA TTA CCC CAA AGG TAT CGT GCA AGA ATA CGC	1584
	Val Asn Ile Asn Gly Gln Leu Pro Gln Arg Tyr Arg Ala Arg Ile Arg	
	515 520 525	
10	TAT GCC TCT ACT ACA AAT CTA AGA ATT TAC GTA ACG GTT GCA GGT GAA	1632
	Tyr Ala Ser Thr Thr Asn Leu Arg Ile Tyr Val Thr Val Ala Gly Glu	
	530 535 540	
15	CGG ATT TTT GCT GGT CAA TTT AAC AAA ACA ATG GAT ACC GGT GAC CCA	1680
	Arg Ile Phe Ala Gly Gln Phe Asn Lys Thr Met Asp Thr Gly Asp Pro	
	545 550 555 560	
20	TTA ACA TTC CAA TCT TTT AGT TAC GCA ACT ATT AAT ACA GCT TTT ACA	1728
	Leu Thr Phe Gln Ser Phe Ser Tyr Ala Thr Ile Asn Thr Ala Phe Thr	
	565 570 575	
25	TTC CCA ATG AGC CAG AGT AGT TTC ACA GTA GGT GCT GAT ACT TTT AGT	1776
	Phe Pro Met Ser Gln Ser Ser Phe Thr Val Gly Ala Asp Thr Phe Ser	
	580 585 590	
30	TCA GGG AAT GAA GTT TAT ATA GAC AGA TTT GAA TTG ATT CCA GTT ACT	1824
	Ser Gly Asn Glu Val Tyr Ile Asp Arg Phe Glu Leu Ile Pro Val Thr	
	595 600 605	
35	GCA ACA TTT GAA GCA GAA TAT GAT TTA GAA AGA GCA CAA AAG GCG GTG	1872
	Ala Thr Phe Glu Ala Glu Tyr Asp Leu Glu Arg Ala Gln Lys Ala Val	
	610 615 620	
40	AAT GCG CTG TTT ACT TCT ATA AAC CAA ATA GGG ATA AAA ACA GAT GTG	1920
	Asn Ala Leu Phe Thr Ser Ile Asn Gln Ile Gly Ile Lys Thr Asp Val	
	625 630 635 640	
45	ACG GAT TAT CAT ATT GAT CAA GTA TCC AAT TTA GTG GAT TGT TTA TCA	1968
	Thr Asp Tyr His Ile Asp Gln Val Ser Asn Leu Val Asp Cys Leu Ser	
	645 650 655	
50	GAT GAA TTT TGT CTG GAT GAA AAG CGA GAA TTG TCC GAG AAA GTC AAA	2016
	Asp Glu Phe Cys Leu Asp Glu Lys Arg Glu Leu Ser Glu Lys Val Lys	
	660 665 670	
55	CAT GCG AAG CGA CTC AGT GAT GAG CGG AAT TTA CTT CAA GAT CCA AAC	2064
	His Ala Lys Arg Leu Ser Asp Glu Arg Asn Leu Leu Gln Asp Pro Asn	
	675 680 685	
60	TTC AAA GGC ATC AAT AGG CAA CTA GAC CGT GGT TGG AGA GGA AGT ACG	2112
	Phe Lys Gly Ile Asn Arg Gln Leu Asp Arg Gly Trp Arg Gly Ser Thr	
	690 695 700	

	GAT ATT ACC ATC CAA AGA GGA GAT GAC GTA TTC AAA GAA AAT TAT GTC	2160
	Asp Ile Thr Ile Gln Arg Gly Asp Asp Val Phe Lys Glu Asn Tyr Val	
	705 710 715 720	
5	ACA CTA CCA GGT ACC TTT GAT GAG TGC TAT CCA ACA TAT TTG TAT CAA	2208
	Thr Leu Pro Gly Thr Phe Asp Glu Cys Tyr Pro Thr Tyr Leu Tyr Gln	
	725 730 735	
10	AAA ATC GAT GAA TCA AAA TTA AAA GCC TTT ACC CGT TAT CAA TTA AGA	2256
	Lys Ile Asp Glu Ser Lys Leu Lys Ala Phe Thr Arg Tyr Gln Leu Arg	
	740 745 750	
15	GGG TAT ATC GAA GAT AGT CAA GAC TTA GAA ATC TAT TTA ATT CGC TAC	2304
	Gly Tyr Ile Glu Asp Ser Gln Asp Leu Glu Ile Tyr Leu Ile Arg Tyr	
	755 760 765	
	AAT GCA AAA CAT GAA ACA GTA AAT GTG CCA GGT ACG GGT TCC TTA TGG	2352
	Asn Ala Lys His Glu Thr Val Asn Val Pro Gly Thr Gly Ser Leu Trp	
	770 775 780	
20	CCG CTT TCA GCC CAA AGT CCA ATC GGA AAG TGT GGA GAG CCG AAT CGA	2400
	Pro Leu Ser Ala Gln Ser Pro Ile Gly Lys Cys Gly Glu Pro Asn Arg	
	785 790 795 800	
25	TGC GCG CCA CAC CTT GAA TGG AAT CCT GAC TTA GAT TGT TCG TGT AGG	2448
	Cys Ala Pro His Leu Glu Trp Asn Pro Asp Leu Asp Cys Ser Cys Arg	
	805 810 815	
30	GAT GGA GAA AAG TGT GCC CAT CAT TCG CAT CAT TTC TCC TTA GAC ATT	2496
	Asp Gly Glu Lys Cys Ala His His Ser His His Phe Ser Leu Asp Ile	
	820 825 830	
	GAT GTA GGA TGT ACA GAC TTA AAT GAG GAC CTA GGT GTA TGG GTG ATC	2544
	Asp Val Gly Cys Thr Asp Leu Asn Glu Asp Leu Gly Val Trp Val Ile	
	835 840 845	
35	TTT AAG ATT AAG ACG CAA GAT GGG CAC GCA AGA CTA GGG AAT CTA GAG	2592
	Phe Lys Ile Lys Thr Gln Asp Gly His Ala Arg Leu Gly Asn Leu Glu	
	850 855 860	
40	TTT CTC GAA GAG AAA CCA TTA GTA GGA GAA GCG CTA GCT CGT GTG AAA	2640
	Phe Leu Glu Glu Lys Pro Leu Val Gly Glu Ala Leu Ala Arg Val Lys	
	865 870 875 880	
45	AGA GCG GAG AAA AAA TGG AGA GAC AAA CGT GAA AAA TTG GAA TGG GAA	2688
	Arg Ala Glu Lys Lys Trp Arg Asp Lys Arg Glu Lys Leu Glu Trp Glu	
	885 890 895	
50	ACA AAT ATC GTT TAT AAA GAG GCA AAA GAA TCT GTA GAT GCT TTA TTT	2736
	Thr Asn Ile Val Tyr Lys Glu Ala Lys Glu Ser Val Asp Ala Leu Phe	
	900 905 910	
	GTA AAC TCT CAA TAT GAT CAA TTA CAA GCG GAT ACG AAT ATT GCC ATG	2784
	Val Asn Ser Gln Tyr Asp Gln Leu Gln Ala Asp Thr Asn Ile Ala Met	

	915	920	925	
5	ATT CAT GCG GCA GAT AAA CGT GTT CAT AGC ATT CGA GAA GCT TAT CTG Ile His Ala Ala Asp Lys Arg Val His Ser Ile Arg Glu Ala Tyr Leu 930 935 940	2832		
10	CCT GAG CTG TCT GTG ATT CCG GGT GTC AAT GCG GCT ATT TTT GAA GAA Pro Glu Leu Ser Val Ile Pro Gly Val Asn Ala Ala Ile Phe Glu Glu 945 950 955 960	2880		
	TTA GAA GGG CGT ATT TTC ACT GCA TTC TCC CTA TAT GAT GCG AGA AAT Leu Glu Gly Arg Ile Phe Thr Ala Phe Ser Leu Tyr Asp Ala Arg Asn 965 970 975	2928		
15	GTC ATT AAA AAT GGT GAT TTT AAT AAT GGC TTA TCC TGC TGG AAC GTG Val Ile Lys Asn Gly Asp Phe Asn Asn Gly Leu Ser Cys Trp Asn Val 980 985 990	2976		
20	AAA GGG CAT GTA GAT GTA GAA GAA CAA AAC AAC CAA CGT TCG GTC CTT Lys Gly His Val Asp Val Glu Glu Gln Asn Asn Gln Arg Ser Val Leu 995 1000 1005	3024		
25	GTT GTT CCG GAA TGG GAA GCA GAA GTG TCA CAA GAA GTT CGT GTC TGT Val Val Pro Glu Trp Glu Ala Glu Val Ser Gln Glu Val Arg Val Cys 1010 1015 1020	3072		
30	CCG GGT CGT GGC TAT ATC CTT CGT GTC ACA GCG TAC AAG GAG GGA TAT Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr 1025 1030 1035 1040	3120		
	GGA GAA GGT TGC GTA ACC ATT CAT GAG ATC GAG AAC AAT ACA GAC GAA Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu 1045 1050 1055	3168		
35	CTG AAG TTT AGC AAC TGC GTA GAA GAG GAA ATC TAT CCA AAT AAC ACG Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr 1060 1065 1070	3216		
40	GTA ACG TGT AAT GAT TAT ACT GTA AAT CAA GAA GAA TAC GGA GGT GCG Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala 1075 1080 1085	3264		
45	TAC ACT TCT CGT AAT CGA GGA TAT AAC GAA GCT CCT TCC GTA CCA GCT Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala 1090 1095 1100	3312		
50	GAT TAT GCG TCA GTC TAT GAA GAA AAA TCG TAT ACA GAT GGA CGA AGA Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg 1105 1110 1115 1120	3360		
	GAG AAT CCT TGT GAA TTT AAC AGA GGG TAT AGG GAT TAC ACG CCA CTA Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu 1125 1130 1135	3408		

	CCA GTT GGT TAT GTG ACA AAA GAA TTA GAA TAC TTC CCA GAA ACC GAT	3456
	Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp	
	1140 1145 1150	
5	AAG GTA TGG ATT GAG ATT GGA GAA ACG GAA GGA ACA TTT ATC GTG GAC	3504
	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp	
	1155 1160 1165	
10	AGC GTG GAA TTA CTC CTT ATG GAG GAA TAG	3534
	Ser Val Glu Leu Leu Leu Met Glu Glu	
	1170 1175	

(2) INFORMATION FOR SEQ ID NO:28:

15 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1177 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

25	Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu	
	1 5 10 15	
	Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly	
	20 25 30	
30	Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser	
	35 40 45	
	Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile	
35	50 55 60	
	Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile	
	65 70 75 80	
40	Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala	
	85 90 95	
	Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu	
	100 105 110	
45	Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu	
	115 120 125	
	Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala	
50	130 135 140	
	Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val	
	145 150 155 160	

	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	165	170	175
5	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	180	185	190
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	195	200	205
10	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	210	215	220
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	225	230	235
15	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	245	250	255
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	260	265	270
20	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	275	280	285
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	290	295	300
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	305	310	315
30	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	325	330	335
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	340	345	350
35	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	355	360	365
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	370	375	380
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	385	390	395
45	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	405	410	415
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	420	425	430
50	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	435	440	445

	Arg	Ala	Pro	Met	Phe	Ser	Trp	Ile	His	Arg	Ser	Ala	Glu	Phe	Asn	Asn	
	450						455					460					
5	Ile	Ile	Ala	Ser	Asp	Ser	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
	465					470				475					480		
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485					490					495		
10	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505						510		
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
			515					520					525				
15	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	
		530					535					540					
	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
20		545				550				555					560		
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
				565					570						575		
25	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	
				580					585					590			
	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	
			595					600					605				
30	Ala	Thr	Phe	Glu	Ala	Glu	Tyr	Asp	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val	
		610					615					620					
	Asn	Ala	Leu	Phe	Thr	Ser	Ile	Asn	Gln	Ile	Gly	Ile	Lys	Thr	Asp	Val	
35		625				630					635					640	
	Thr	Asp	Tyr	His	Ile	Asp	Gln	Val	Ser	Asn	Leu	Val	Asp	Cys	Leu	Ser	
				645					650					655			
40	Asp	Glu	Phe	Cys	Leu	Asp	Glu	Lys	Arg	Glu	Leu	Ser	Glu	Lys	Val	Lys	
				660				665						670			
	His	Ala	Lys	Arg	Leu	Ser	Asp	Glu	Arg	Asn	Leu	Leu	Gln	Asp	Pro	Asn	
				675				680						685			
45	Phe	Lys	Gly	Ile	Asn	Arg	Gln	Leu	Asp	Arg	Gly	Trp	Arg	Gly	Ser	Thr	
		690					695					700					
	Asp	Ile	Thr	Ile	Gln	Arg	Gly	Asp	Asp	Val	Phe	Lys	Glu	Asn	Tyr	Val	
50		705				710					715					720	
	Thr	Leu	Pro	Gly	Thr	Phe	Asp	Glu	Cys	Tyr	Pro	Thr	Tyr	Leu	Tyr	Gln	
				725						730					735		

	Lys	Ile	Asp	Glu	Ser	Lys	Leu	Lys	Ala	Phe	Thr	Arg	Tyr	Gln	Leu	Arg	
				740					745					750			
5	Gly	Tyr	Ile	Glu	Asp	Ser	Gln	Asp	Leu	Glu	Ile	Tyr	Leu	Ile	Arg	Tyr	
				755				760					765				
	Asn	Ala	Lys	His	Glu	Thr	Val	Asn	Val	Pro	Gly	Thr	Gly	Ser	Leu	Trp	
				770			775					780					
10	Pro	Leu	Ser	Ala	Gln	Ser	Pro	Ile	Gly	Lys	Cys	Gly	Glu	Pro	Asn	Arg	
							790				795					800	
	Cys	Ala	Pro	His	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg	
					805					810					815		
15	Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile	
				820					825					830			
	Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile	
20				835				840					845				
	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	
				850			855					860					
25	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	
						870					875					880	
	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu	
					885					890					895		
30	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	
				900					905					910			
	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met	
35				915				920					925				
	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu	
				930			935					940					
40	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	
						950					955					960	
	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	
					965					970					975		
45	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val	
				980					985					990			
	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu	
50				995				1000					1005				
	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	
				1010			1015					1020					

Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr
1025 1030 1035 1040

5 Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu
1045 1050 1055

Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr
1060 1065 1070

10 Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala
1075 1080 1085

Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala
1090 1095 1100

15 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
1105 1110 1115 1120

Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu
1125 1130 1135

20 Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp
1140 1145 1150

25 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
1155 1160 1165

Ser Val Glu Leu Leu Leu Met Glu Glu
1170 1175

30

(2) INFORMATION FOR SEQ ID NO:29:

35 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3579 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

40 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

ATGGATAACA ATCCGAACAT CAATGAATGC ATTCCTTATA ATTGTTTAAG TAACCCTGAA	60
GTAGAAGTAT TAGGTGGAGA AAGAATAGAA ACTGGTTACA CCCCAATCGA TATTTCTTG	120
45 TCGCTAACGC AATTTCTTTT GAGTGAATTT GTTCCCGGTG CTGGATTGT GTTAGGACTA	180
GTTGATATAA TATGGGGAAT TTTTGGTCCC TCTCAATGGG ACGCATTTCT TGTACAAATT	240
50 GAACAGTTAA TTAACCAAAG AATAGAAGAA TTCGCTAGGA ACCAAGCCAT TTCTAGATTA	300
GAAGGACTAA GCAATCTTTA TCAAATTTAC GCAGAATCTT TTAGAGAGTG GGAAGCAGAT	360
CCTACTAATC CAGCATTAAAG AGAAGAGATG CGTATTCAAT TCAATGACAT GAACAGTGCC	420

	CTTACAACCG CTATTCCTCT TTTGTCAGTT CAAAATTATC AAGTTCCTCT TTTATCAGTA	480
	TATGTTCAAG CTGCAAATTT ACATTTATCA GTTTTGAGAG ATGTTTCAGT GTTTGGACAA	540
5	AGGTGGGGAT TTGATGCCGC GACTATCAAT AGTCGTTATA ATGATTTAAC TAGGCTTATT	600
	GGCAACTATA CAGATTATGC TGTACGCTGG TACAATACGG GATTAGAACG TGTATGGGGA	660
10	CCGGATTCTA GAGATTGGGT AAGGTATAAT CAATTTAGAA GAGAATTAAC ACTAACTGTA	720
	TTAGATATCG TTGCTCTGTT CCCGAATTAT GATAGTAGAA GATATCCAAT TCGAACAGTT	780
	TCCCAATTAA CAAGAGAAAT TTATACAAAC CCAGTATTAG AAAATTTTGA TGGTAGTTTT	840
15	CGAGGCTCGG CTCAGGGCAT AGAAAGAAGT ATTAGGAGTC CACATTTGAT GGATATACTT	900
	AACAGTATAA CCATCTATAC GGATGCTCAT AGGGGTTATT ATTATTGGTC AGGGCATCAA	960
20	ATAATGGCTT CTCCTGTAGG GTTTTCGGGG CCAGAATTCA CTTTCCGCT ATATGGAAC	1020
	ATGGGAAATG CAGCTCCACA ACAACGTATT GTTGCTCAAC TAGGTCAGGG CGTGTATAGA	1080
	ACATTATCGT CCACTTTATA TAGAAGACCT TTTAATATAG GGATAAATAA TCAACAATA	1140
25	TCTGTTCTTG ACGGGACAGA ATTTGCTTAT GGAACCTCCT CAAATTTGCC ATCCGCTGTA	1200
	TACAGAAAAA GCGGAACGGT AGATTCGCTG GATGAAATAC CGCCACAGAA TAACAACGTG	1260
30	CCACCTAGGC AAGGATTTAG TCATCGATTA AGCCATGTTT CAATGTTTCG TTCAGGCTTT	1320
	AGTAATAGTA GTGTAAGTAT AATAAGAGCT CCTATGTTCT CTTGGATACA TCGTAGTGCA	1380
	ACTCTTACAA ATACAATTGA TCCAGAGAGA ATTAATCAAA TACCTTTAGT GAAAGGATTT	1440
35	AGAGTTTGGG GGGGCACCTC TGTCATTACA GGACCAGGAT TTACAGGAGG GGATATCCTT	1500
	CGAAGAAATA CCTTTGGTGA TTTTGTATCT CTACAAGTCA ATATTAATTC ACCAATTACC	1560
40	CAAAGATACC GTTTAAGATT TCGTTACGCT TCCAGTAGGG ATGCACGAGT TATAGTATTA	1620
	ACAGGAGCGG CATCCACAGG AGTGGGAGGC CAAGTTAGTG TAAATATGCC TCTTCAGAAA	1680
	ACTATGGAAA TAGGGGAGAA CTTAACATCT AGAACATTTA GATATACCGA TTTTAGTAAT	1740
45	CCTTTTTTCAT TTAGAGCTAA TCCAGATATA ATTGGGATAA GTGAACAACC TCTATTGGT	1800
	GCAGGTTCTA TTAGTAGCGG TGAACCTTAT ATAGATAAAA TTGAAATTAT TCTAGCAGAT	1860
50	GCAACATTTG AAGCAGAATC TGATTTAGAA AGAGCACAAA AGGCGGTGAA TGCCCTGTTT	1920
	ACTTCTTCCA ATCAAATCGG GTTAAAAACC GATGTGACGG ATTATCATAT TGATCAAGTA	1980
	TCCAATTTAG TGGATTGTTT ATCAGATGAA TTTTGTCTGG ATGAAAAGCG AGAATTGTCC	2040

	GAGAAAGTCA AACATGCGAA GCGACTCAGT GATGAGCGGA ATTTACTTCA AGATCCAAAC	2100
	TTCAGAGGGA TCAATAGACA ACCAGACCGT GGCTGGAGAG GAAGTACAGA TATTACCATC	2160
5	CAAGGAGGAG ATGACGTATT CAAAGAGAAT TACGTCACAC TACCGGGTAC CGTTGATGAG	2220
	TGCTATCCAA CGTATTTATA TCAGAAAATA GATGAGTCGA AATTAAGAGC TTATACCCGT	2280
10	TATGAATTAA GAGGGTATAT CGAAGATAGT CAAGACTTAG AAATCTATTT GATCCGTTAC	2340
	AATGCAAAAC ACGAAATAGT AAATGTGCCA GGCACGGGTT CCTTATGGCC GCTTTCAGCC	2400
	CAAAGTCCAA TCGGAAAGTG TGGAGAACCG AATCGATGCG CGCCACACCT TGAATGGAAT	2460
15	CCTGATCTAG ATTGTTTCCTG CAGAGACGGG GAAAAATGTG CACATCATTC CCATCATTTTC	2520
	ACCTTGGATA TTGATGTTGG ATGTACAGAC TTAAATGAGG ACTTAGGTGT ATGGGTGATA	2580
20	TTCAAGATTA AGACGCAAGA TGGCCATGCA AGACTAGGGA ATCTAGAGTT TCTCGAAGAG	2640
	AAACCATTAT TAGGGGAAGC ACTAGCTCGT GTGAAAAGAG CGGAGAAGAA GTGGAGAGAC	2700
	AAACGAGAGA AACTGCAGTT GGAAACAAAT ATTGTTTATA AAGAGGCAAA AGAATCTGTA	2760
25	GATGCTTTAT TTGTAAACTC TCAATATGAT AGATTACAAG TGGATACGAA CATCGCAATG	2820
	ATTCATGCGG CAGATAAACG CGTTCATAGA ATCCGGGAAG CGTATCTGCC AGAGTTGTCT	2880
30	GTGATTCCAG GTGTCAATGC GGCCATTTTC GAAGAATTAG AGGGACGTAT TTTTACAGCG	2940
	TATTCCTTAT ATGATGCGAG AAATGTCATT AAAAATGGCG ATTTCAATAA TGGCTTATTA	3000
	TGCTGGAACG TGAAAGGTCA TGTAGATGTA GAAGAGCAAA ACAACCACCG TTCGGTCCTT	3060
35	GTTATCCCAG AATGGGAGGC AGAAGTGTC AAGAGGTTT GTGTCTGTCC AGGTGCTGGC	3120
	TATATCCTTC GTGTCACAGC ATATAAAGAG GGATATGGAG AGGGCTGCGT AACGATCCAT	3180
40	GAGATCGAAG ACAATACAGA CGAACTGAAA TTCAGCAACT GTGTAGAAGA GGAAGTATAT	3240
	CCAAACAACA CAGTAACGTG TAATAATTAT ACTGGGACTC AAGAAGAATA TGAGGGTACG	3300
	TACACTTCTC GTAATCAAGG ATATGACGAA GCCTATGGTA ATAACCCTTC CGTACCAGCT	3360
45	GATTACGCTT CAGTCTATGA AGAAAAATCG TATACAGATG GACGAAGAGA GAATCCTTGT	3420
	GAATCTAACA GAGGCTATGG GGATTACACA CCACTACCGG CTGGTTATGT AACAAAGGAT	3480
50	TTAGAGTACT TCCCAGAGAC CGATAAGGTA TGGATTGAGA TCGGAGAAAC AGAAGGAACA	3540
	TTCATCGTGG ATAGCGTGGA ATTACTCCTT ATGGAGGAA	3579

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 1193 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS:
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

10 Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
1 5 10 15

15 Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
20 25 30

Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
35 40 45

20 Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
50 55 60

Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
65 70 75 80

25 Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
85 90 95

Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
100 105 110

Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
115 120 125

35 Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
130 135 140

Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
145 150 155 160

40 Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser
165 170 175

Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg
180 185 190

Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val
195 200 205

50 Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg
210 215 220

Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val
225 230 235 240

	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	245	250	255
5	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	260	265	270
	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	275	280	285
10	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	290	295	300
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	305	310	315
15	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	325	330	335
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	340	345	350
	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	355	360	365
25	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	370	375	380
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	385	390	395
30	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	405	410	415
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	420	425	430
	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	435	440	445
40	Arg Ala Pro Met Phe Ser Trp Ile His Arg Ser Ala Thr Leu Thr Asn	450	455	460
	Thr Ile Asp Pro Glu Arg Ile Asn Gln Ile Pro Leu Val Lys Gly Phe	465	470	475
45	Arg Val Trp Gly Gly Thr Ser Val Ile Thr Gly Pro Gly Phe Thr Gly	485	490	495
	Gly Asp Ile Leu Arg Arg Asn Thr Phe Gly Asp Phe Val Ser Leu Gln	500	505	510
50	Val Asn Ile Asn Ser Pro Ile Thr Gln Arg Tyr Arg Leu Arg Phe Arg	515	520	525

	Tyr	Ala	Ser	Ser	Arg	Asp	Ala	Arg	Val	Ile	Val	Leu	Thr	Gly	Ala	Ala	
	530						535					540					
5	Ser	Thr	Gly	Val	Gly	Gly	Gln	Val	Ser	Val	Asn	Met	Pro	Leu	Gln	Lys	
	545						550				555					560	
	Thr	Met	Glu	Ile	Gly	Glu	Asn	Leu	Thr	Ser	Arg	Thr	Phe	Arg	Tyr	Thr	
					565					570						575	
10	Asp	Phe	Ser	Asn	Pro	Phe	Ser	Phe	Arg	Ala	Asn	Pro	Asp	Ile	Ile	Gly	
				580						585					590		
	Ile	Ser	Glu	Gln	Pro	Leu	Phe	Gly	Ala	Gly	Ser	Ile	Ser	Ser	Gly	Glu	
15			595					600					605				
	Leu	Tyr	Ile	Asp	Lys	Ile	Glu	Ile	Ile	Leu	Ala	Asp	Ala	Thr	Phe	Glu	
	610						615					620					
20	Ala	Glu	Ser	Asp	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val	Asn	Ala	Leu	Phe	
	625					630					635					640	
	Thr	Ser	Ser	Asn	Gln	Ile	Gly	Leu	Lys	Thr	Asp	Val	Thr	Asp	Tyr	His	
				645						650						655	
25	Ile	Asp	Gln	Val	Ser	Asn	Leu	Val	Asp	Cys	Leu	Ser	Asp	Glu	Phe	Cys	
				660					665						670		
	Leu	Asp	Glu	Lys	Arg	Glu	Leu	Ser	Glu	Lys	Val	Lys	His	Ala	Lys	Arg	
30			675					680					685				
	Leu	Ser	Asp	Glu	Arg	Asn	Leu	Leu	Gln	Asp	Pro	Asn	Phe	Arg	Gly	Ile	
	690						695					700					
35	Asn	Arg	Gln	Pro	Asp	Arg	Gly	Trp	Arg	Gly	Ser	Thr	Asp	Ile	Thr	Ile	
	705					710					715					720	
	Gln	Gly	Gly	Asp	Asp	Val	Phe	Lys	Glu	Asn	Tyr	Val	Thr	Leu	Pro	Gly	
				725						730						735	
40	Thr	Val	Asp	Glu	Cys	Tyr	Pro	Thr	Tyr	Leu	Tyr	Gln	Lys	Ile	Asp	Glu	
				740					745						750		
	Ser	Lys	Leu	Lys	Ala	Tyr	Thr	Arg	Tyr	Glu	Leu	Arg	Gly	Tyr	Ile	Glu	
45			755					760					765				
	Asp	Ser	Gln	Asp	Leu	Glu	Ile	Tyr	Leu	Ile	Arg	Tyr	Asn	Ala	Lys	His	
	770						775					780					
50	Glu	Ile	Val	Asn	Val	Pro	Gly	Thr	Gly	Ser	Leu	Trp	Pro	Leu	Ser	Ala	
	785					790					795					800	
	Gln	Ser	Pro	Ile	Gly	Lys	Cys	Gly	Glu	Pro	Asn	Arg	Cys	Ala	Pro	His	
				805						810						815	

	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg	Asp	Gly	Glu	Lys	
				820					825					830			
5	Cys	Ala	His	His	Ser	His	His	Phe	Thr	Leu	Asp	Ile	Asp	Val	Gly	Cys	
			835					840					845				
	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile	Phe	Lys	Ile	Lys	
		850					855					860					
10	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	Phe	Leu	Glu	Glu	
	865					870				875						880	
	Lys	Pro	Leu	Leu	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	Arg	Ala	Glu	Lys	
15				885						890					895		
	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Gln	Leu	Glu	Thr	Asn	Ile	Val	
			900						905					910			
20	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	Val	Asn	Ser	Gln	
		915						920					925				
	Tyr	Asp	Arg	Leu	Gln	Val	Asp	Thr	Asn	Ile	Ala	Met	Ile	His	Ala	Ala	
		930					935					940					
25	Asp	Lys	Arg	Val	His	Arg	Ile	Arg	Glu	Ala	Tyr	Leu	Pro	Glu	Leu	Ser	
	945				950						955					960	
	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	Leu	Glu	Gly	Arg	
30				965					970						975		
	Ile	Phe	Thr	Ala	Tyr	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	Val	Ile	Lys	Asn	
			980						985					990			
35	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Leu	Cys	Trp	Asn	Val	Lys	Gly	His	Val	
		995					1000						1005				
	Asp	Val	Glu	Glu	Gln	Asn	Asn	His	Arg	Ser	Val	Leu	Val	Ile	Pro	Glu	
	1010					1015						1020					
40	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	Pro	Gly	Arg	Gly	
	1025				1030					1035						1040	
	Tyr	Ile	Leu	Arg	Val	Thr	Ala	Tyr	Lys	Glu	Gly	Tyr	Gly	Glu	Gly	Cys	
45				1045					1050					1055			
	Val	Thr	Ile	His	Glu	Ile	Glu	Asp	Asn	Thr	Asp	Glu	Leu	Lys	Phe	Ser	
			1060					1065						1070			
50	Asn	Cys	Val	Glu	Glu	Glu	Val	Tyr	Pro	Asn	Asn	Thr	Val	Thr	Cys	Asn	
		1075						1080					1085				
	Asn	Tyr	Thr	Gly	Thr	Gln	Glu	Glu	Tyr	Glu	Gly	Thr	Tyr	Thr	Ser	Arg	
		1090				1095						1100					

Asn Gln Gly Tyr Asp Glu Ala Tyr Gly Asn Asn Pro Ser Val Pro Ala
 1105 1110 1115 1120
 5 Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg
 1125 1130 1135
 Glu Asn Pro Cys Glu Ser Asn Arg Gly Tyr Gly Asp Tyr Thr Pro Leu
 1140 1145 1150
 10 Pro Ala Gly Tyr Val Thr Lys Asp Leu Glu Tyr Phe Pro Glu Thr Asp
 1155 1160 1165
 Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp
 1170 1175 1180
 15 Ser Val Glu Leu Leu Leu Met Glu Glu
 1185 1190

20

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 16 base pairs
 25 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

30

CGTTGCTCTG TTCCCG

16

35 (2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 20 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 40 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

45 TCAAATACCA TTGGTAAAAG

20

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3534 base pairs
 50 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

	ATGGATAACA ATCCGAACAT CAATGAATGC ATTCCTTATA ATTGTTTAAG TAACCCTGAA	60
5	GTAGAAGTAT TAGGTGGAGA AAGAATAGAA ACTGGTTACA CCCCAATCGA TATTCCTTG	120
	TCGCTAACGC AATTTCTTTT GAGTGAATTT GTTCCCGGTG CTGGATTTGT GTTAGGACTA	180
	GTTGATATAA TATGGGGAAT TTTTGGTCCC TCTCAATGGG ACGCATTTCT TGTACAAATT	240
10	GAACAGTTAA TTAACCAAAG AATAGAAGAA TTCGCTAGGA ACCAAGCCAT TTCTAGATTA	300
	GAAGGACTAA GCAATCTTTA TCAAATTTAC GCAGAATCTT TTAGAGAGTG GGAAGCAGAT	360
15	CCTACTAATC CAGCATTAAG AGAAGAGATG CGTATTCAAT TCAATGACAT GAACAGTGCC	420
	CTTACAACCG CTATTCCTCT TTTTGCAGTT CAAAATTATC AAGTTCCTCT TTTATCAGTA	480
	TATGTTCAAG CTGCAAATTT ACATTTATCA GTTTTGAGAG ATGTTTCAGT GTTTGGACAA	540
20	AGGTGGGGAT TTGATGCCGC GACTATCAAT AGTCGTTATA ATGATTTAAC TAGGCTTATT	600
	GGCAACTATA CAGATTATGC TGTACGCTGG TACAATACGG GATTAGAACG TGTATGGGGA	660
25	CCGGATTCTA GAGATTGGGT AAGGTATAAT CAATTTAGAA GAGAATTAAC ACTAACTGTA	720
	TTAGATATCG TTGCTCTGTT CCCGAATTAT GATAGTAGAA GATATCCAAT TCGAACAGTT	780
	TCCCAATTAA CAAGAGAAAT TTATACAAAC CCAGTATTAG AAAATTTTGA TGGTAGTTTT	840
30	CGAGGCTCGG CTCAGGGCAT AGAAAGAAGT ATTAGGAGTC CACATTTGAT GGATATACTT	900
	AACAGTATAA CCATCTATAC GGATGCTCAT AGGGGTTATT ATTATTGGTC AGGGCATCAA	960
35	ATAATGGCTT CTCCTGTAGG GTTTTCGGGG CCAGAATTCA CTTTCCGCT ATATGGAAC	1020
	ATGGGAAATG CAGCTCCACA ACAACGTATT GTTGCTCAAC TAGGTCAGGG CGTGTATAGA	1080
	ACATTATCGT CCACTTTATA TAGAAGACCT TTAAATATAG GGATAAATAA TCAACAATA	1140
40	TCTGTTCTTG ACGGGACAGA ATTTGCTTAT GGAACCTCCT CAAATTTGCC ATCCGCTGTA	1200
	TACAGAAAAA GCGGAACGGT AGATTGCTG GATGAAATAC CGCCACAGAA TAACAACGTG	1260
45	CCACCTAGGC AAGGATTTAG TCATCGATTA AGCCATGTTT CAATGTTTCG TTCAGGCTTT	1320
	AGTAATAGTA GTGTAAGTAT AATAAGAGCT CCTATGTTCT CTTGGATACA TCGTAGTGCT	1380
	GAATTTAATA ATATAATTGC ATCGGATAGT ATTACTCAA TACCATTGGT AAAAGCACAT	1440
50	ACACTTCAGT CAGGTACTAC TGTGTGAAGA GGGCCCGGGT TTACGGGAGG AGATATTCTT	1500
	CGACGAACAA GTGGAGGACC ATTTGCTTAT ACTATTGTTA ATATAAATGG GCAATTACCC	1560

	CAAAGGTATC GTGCAAGAAT ACGCTATGCC TCTACTACAA ATCTAAGAAT TTACGTAACG	1620
	GTTGCAGGTG AACGGATTTT TGCTGGTCAA TTTAACAAAA CAATGGATAC CGGTGACCCA	1680
5	TTAACATTCC AATCTTTTAG TTACGCAACT ATTAATACAG CTTTACATT CCCAATGAGC	1740
	CAGAGTAGTT TCACAGTAGG TGCTGATACT TTTAGTTCAG GGAATGAAGT TTATATAGAC	1800
10	AGATTTGAAT TGATTCCAGT TACTGCAACA CTCGAGGCTG AATATAATCT GGAAAGAGCG	1860
	CAGAAGGCGG TGAATGCGCT GTTTACGTCT ACAAACCAAC TAGGGCTAAA AACAAATGTA	1920
	ACGGATTATC ATATTGATCA AGTGTCCAAT TTAGTTACGT ATTTATCGGA TGAATTTTGT	1980
15	CTGGATGAAA AGCGAGAATT GTCCGAGAAA GTCAAACATG CGAAGCGACT CAGTGATGAA	2040
	CGCAATTTAC TCCAAGATTG AAATTTCAAA GACATTAATA GGCAACCAGA ACGTGGGTGG	2100
20	GGCGGAAGTA CAGGGATTAC CATCCAAGGA GGGGATGACG TATTTAAAGA AAATTACGTC	2160
	ACACTATCAG GTACCTTTGA TGAGTGCTAT CCAACATATT TGTATCAAAA AATCGATGAA	2220
	TCAAAATTAA AAGCCTTTAC CCGTTATCAA TTAAGAGGGT ATATCGAAGA TAGTCAAGAC	2280
25	TTAGAAATCT ATTTAATTCG CTACAATGCA AAACATGAAA CAGTAAATGT GCCAGGTACG	2340
	GGTTCCTTAT GGCCGCTTTC AGCCCAAAGT CCAATCGGAA AGTGTGGAGA GCCGAATCGA	2400
30	TGCGCGCCAC ACCTTGAATG GAATCCTGAC TTAGATTGTT CGTGTAGGGA TGGAGAAAAG	2460
	TGTGCCCATC ATTCGCATCA TTTCTCCTTA GACATTGATG TAGGATGTAC AGACTTAAAT	2520
	GAGGACCTAG GTGTATGGGT GATCTTTAAG ATTAAGACGC AAGATGGGCA CGCAAGACTA	2580
35	GGGAATCTAG AGTTTCTCGA AGAGAAACCA TTAGTAGGAG AAGCGCTAGC TCGTGTGAAA	2640
	AGAGCGGAGA AAAAATGGAG AGACAAACGT GAAAAATTGG AATGGGAAAC AAATATCGTT	2700
40	TATAAGAGG CAAAAGAATC TGTAGATGCT TTATTTGTAA ACTCTCAATA TGATCAATTA	2760
	CAAGCGGATA CGAATATTGC CATGATTCAT GCGGCAGATA AACGTGTTCA TAGCATTCTGA	2820
	GAAGCTTATC TGCCTGAGCT GTCTGTGATT CCGGGTGTCA ATGCGGCTAT TTTTGAAGAA	2880
45	TTAGAAGGGC GTATTTTCAC TGCATTCTCC CTATATGATG CGAGAAATGT CATTAAAAAT	2940
	GGTGATTTTA ATAATGGCTT ATCCTGCTGG AACGTGAAAG GGCATGTAGA TGTAGAAGAA	3000
50	CAAAACAACC AACGTTCCGT CCTTGTTGTT CCGGAATGGG AAGCAGAAGT GTCACAAGAA	3060
	GTTCGTGTCT GTCCGGGTCG TGGCTATATC CTTCTGTGTA CAGCGTACAA GGAGGGATAT	3120
	GGAGAAGGTT GCGTAACCAT TCATGAGATC GAGAACAATA CAGACGAACT GAAGTTTAGC	3180

AACTGCGTAG AAGAGGAAAT CTATCCAAAT AACACGGTAA CGTGTAATGA TTATACTGTA 3240
AATCAAGAAG AATACGGAGG TCGGTACACT TCTCGTAATC GAGGATATAA CGAAGCTCCT 3300
5 TCCGTACCAG CTGATTATGC GTCAGTCTAT GAAGAAAAAT CGTATACAGA TGGACGAAGA 3360
GAGAATCCTT GTGAATTTAA CAGAGGGTAT AGGGATTACA CGCCACTACC AGTTGGTTAT 3420
GTGACAAAAG AATTAGAATA CTTCCAGAA ACCGATAAGG TATGGATTGA GATTGGAGAA 3480
10 ACGGAAGGAA CATTTATCGT GGACAGCGTG GAATTACTCC TTATGGAGGA ATAG 3534

(2) INFORMATION FOR SEQ ID NO:34:

15 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1177 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS:
20 (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Met Asp Asn Asn Pro Asn Ile Asn Glu Cys Ile Pro Tyr Asn Cys Leu
25 1 5 10 15
Ser Asn Pro Glu Val Glu Val Leu Gly Gly Glu Arg Ile Glu Thr Gly
20 25 30
Tyr Thr Pro Ile Asp Ile Ser Leu Ser Leu Thr Gln Phe Leu Leu Ser
30 35 40 45
Glu Phe Val Pro Gly Ala Gly Phe Val Leu Gly Leu Val Asp Ile Ile
35 50 55 60
Trp Gly Ile Phe Gly Pro Ser Gln Trp Asp Ala Phe Leu Val Gln Ile
65 70 75 80
Glu Gln Leu Ile Asn Gln Arg Ile Glu Glu Phe Ala Arg Asn Gln Ala
40 85 90 95
Ile Ser Arg Leu Glu Gly Leu Ser Asn Leu Tyr Gln Ile Tyr Ala Glu
100 105 110
Ser Phe Arg Glu Trp Glu Ala Asp Pro Thr Asn Pro Ala Leu Arg Glu
45 115 120 125
Glu Met Arg Ile Gln Phe Asn Asp Met Asn Ser Ala Leu Thr Thr Ala
50 130 135 140
Ile Pro Leu Phe Ala Val Gln Asn Tyr Gln Val Pro Leu Leu Ser Val
145 150 155 160

	Tyr Val Gln Ala Ala Asn Leu His Leu Ser Val Leu Arg Asp Val Ser	165	170	175
5	Val Phe Gly Gln Arg Trp Gly Phe Asp Ala Ala Thr Ile Asn Ser Arg	180	185	190
	Tyr Asn Asp Leu Thr Arg Leu Ile Gly Asn Tyr Thr Asp Tyr Ala Val	195	200	205
10	Arg Trp Tyr Asn Thr Gly Leu Glu Arg Val Trp Gly Pro Asp Ser Arg	210	215	220
	Asp Trp Val Arg Tyr Asn Gln Phe Arg Arg Glu Leu Thr Leu Thr Val	225	230	235
15	Leu Asp Ile Val Ala Leu Phe Pro Asn Tyr Asp Ser Arg Arg Tyr Pro	245	250	255
	Ile Arg Thr Val Ser Gln Leu Thr Arg Glu Ile Tyr Thr Asn Pro Val	260	265	270
20	Leu Glu Asn Phe Asp Gly Ser Phe Arg Gly Ser Ala Gln Gly Ile Glu	275	280	285
	Arg Ser Ile Arg Ser Pro His Leu Met Asp Ile Leu Asn Ser Ile Thr	290	295	300
	Ile Tyr Thr Asp Ala His Arg Gly Tyr Tyr Tyr Trp Ser Gly His Gln	305	310	315
30	Ile Met Ala Ser Pro Val Gly Phe Ser Gly Pro Glu Phe Thr Phe Pro	325	330	335
	Leu Tyr Gly Thr Met Gly Asn Ala Ala Pro Gln Gln Arg Ile Val Ala	340	345	350
35	Gln Leu Gly Gln Gly Val Tyr Arg Thr Leu Ser Ser Thr Leu Tyr Arg	355	360	365
	Arg Pro Phe Asn Ile Gly Ile Asn Asn Gln Gln Leu Ser Val Leu Asp	370	375	380
	Gly Thr Glu Phe Ala Tyr Gly Thr Ser Ser Asn Leu Pro Ser Ala Val	385	390	395
45	Tyr Arg Lys Ser Gly Thr Val Asp Ser Leu Asp Glu Ile Pro Pro Gln	405	410	415
	Asn Asn Asn Val Pro Pro Arg Gln Gly Phe Ser His Arg Leu Ser His	420	425	430
50	Val Ser Met Phe Arg Ser Gly Phe Ser Asn Ser Ser Val Ser Ile Ile	435	440	445

	Arg	Ala	Pro	Met	Phe	Ser	Trp	Ile	His	Arg	Ser	Ala	Glu	Phe	Asn	Asn	
	450						455					460					
5	Ile	Ile	Ala	Ser	Asp	Ser	Ile	Thr	Gln	Ile	Pro	Leu	Val	Lys	Ala	His	
	465					470					475					480	
	Thr	Leu	Gln	Ser	Gly	Thr	Thr	Val	Val	Arg	Gly	Pro	Gly	Phe	Thr	Gly	
					485					490						495	
10	Gly	Asp	Ile	Leu	Arg	Arg	Thr	Ser	Gly	Gly	Pro	Phe	Ala	Tyr	Thr	Ile	
				500					505						510		
	Val	Asn	Ile	Asn	Gly	Gln	Leu	Pro	Gln	Arg	Tyr	Arg	Ala	Arg	Ile	Arg	
				515				520						525			
15	Tyr	Ala	Ser	Thr	Thr	Asn	Leu	Arg	Ile	Tyr	Val	Thr	Val	Ala	Gly	Glu	
	530						535							540			
20	Arg	Ile	Phe	Ala	Gly	Gln	Phe	Asn	Lys	Thr	Met	Asp	Thr	Gly	Asp	Pro	
	545					550					555					560	
	Leu	Thr	Phe	Gln	Ser	Phe	Ser	Tyr	Ala	Thr	Ile	Asn	Thr	Ala	Phe	Thr	
					565					570					575		
25	Phe	Pro	Met	Ser	Gln	Ser	Ser	Phe	Thr	Val	Gly	Ala	Asp	Thr	Phe	Ser	
				580					585					590			
	Ser	Gly	Asn	Glu	Val	Tyr	Ile	Asp	Arg	Phe	Glu	Leu	Ile	Pro	Val	Thr	
			595					600					605				
30	Ala	Thr	Leu	Glu	Ala	Glu	Tyr	Asn	Leu	Glu	Arg	Ala	Gln	Lys	Ala	Val	
	610						615					620					
	Asn	Ala	Leu	Phe	Thr	Ser	Thr	Asn	Gln	Leu	Gly	Leu	Lys	Thr	Asn	Val	
35	625					630					635					640	
	Thr	Asp	Tyr	His	Ile	Asp	Gln	Val	Ser	Asn	Leu	Val	Thr	Tyr	Leu	Ser	
				645						650					655		
40	Asp	Glu	Phe	Cys	Leu	Asp	Glu	Lys	Arg	Glu	Leu	Ser	Glu	Lys	Val	Lys	
				660					665					670			
	His	Ala	Lys	Arg	Leu	Ser	Asp	Glu	Arg	Asn	Leu	Leu	Gln	Asp	Ser	Asn	
				675				680						685			
45	Phe	Lys	Asp	Ile	Asn	Arg	Gln	Pro	Glu	Arg	Gly	Trp	Gly	Gly	Ser	Thr	
	690						695					700					
	Gly	Ile	Thr	Ile	Gln	Gly	Gly	Asp	Asp	Val	Phe	Lys	Glu	Asn	Tyr	Val	
50	705					710					715					720	
	Thr	Leu	Ser	Gly	Thr	Phe	Asp	Glu	Cys	Tyr	Pro	Thr	Tyr	Leu	Tyr	Gln	
					725					730						735	

	Lys	Ile	Asp	Glu	Ser	Lys	Leu	Lys	Ala	Phe	Thr	Arg	Tyr	Gln	Leu	Arg	
				740					745					750			
5	Gly	Tyr	Ile	Glu	Asp	Ser	Gln	Asp	Leu	Glu	Ile	Tyr	Leu	Ile	Arg	Tyr	
				755				760					765				
	Asn	Ala	Lys	His	Glu	Thr	Val	Asn	Val	Pro	Gly	Thr	Gly	Ser	Leu	Trp	
		770					775					780					
10	Pro	Leu	Ser	Ala	Gln	Ser	Pro	Ile	Gly	Lys	Cys	Gly	Glu	Pro	Asn	Arg	
	785					790					795					800	
	Cys	Ala	Pro	His	Leu	Glu	Trp	Asn	Pro	Asp	Leu	Asp	Cys	Ser	Cys	Arg	
				805						810					815		
15	Asp	Gly	Glu	Lys	Cys	Ala	His	His	Ser	His	His	Phe	Ser	Leu	Asp	Ile	
				820					825					830			
	Asp	Val	Gly	Cys	Thr	Asp	Leu	Asn	Glu	Asp	Leu	Gly	Val	Trp	Val	Ile	
20			835					840					845				
	Phe	Lys	Ile	Lys	Thr	Gln	Asp	Gly	His	Ala	Arg	Leu	Gly	Asn	Leu	Glu	
		850					855					860					
25	Phe	Leu	Glu	Glu	Lys	Pro	Leu	Val	Gly	Glu	Ala	Leu	Ala	Arg	Val	Lys	
	865					870					875					880	
	Arg	Ala	Glu	Lys	Lys	Trp	Arg	Asp	Lys	Arg	Glu	Lys	Leu	Glu	Trp	Glu	
				885						890					895		
30	Thr	Asn	Ile	Val	Tyr	Lys	Glu	Ala	Lys	Glu	Ser	Val	Asp	Ala	Leu	Phe	
			900						905					910			
	Val	Asn	Ser	Gln	Tyr	Asp	Gln	Leu	Gln	Ala	Asp	Thr	Asn	Ile	Ala	Met	
35			915					920					925				
	Ile	His	Ala	Ala	Asp	Lys	Arg	Val	His	Ser	Ile	Arg	Glu	Ala	Tyr	Leu	
		930					935					940					
40	Pro	Glu	Leu	Ser	Val	Ile	Pro	Gly	Val	Asn	Ala	Ala	Ile	Phe	Glu	Glu	
	945					950					955					960	
	Leu	Glu	Gly	Arg	Ile	Phe	Thr	Ala	Phe	Ser	Leu	Tyr	Asp	Ala	Arg	Asn	
				965						970					975		
45	Val	Ile	Lys	Asn	Gly	Asp	Phe	Asn	Asn	Gly	Leu	Ser	Cys	Trp	Asn	Val	
			980						985					990			
	Lys	Gly	His	Val	Asp	Val	Glu	Glu	Gln	Asn	Asn	Gln	Arg	Ser	Val	Leu	
50			995					1000					1005				
	Val	Val	Pro	Glu	Trp	Glu	Ala	Glu	Val	Ser	Gln	Glu	Val	Arg	Val	Cys	
			1010				1015					1020					

	Pro Gly Arg Gly Tyr Ile Leu Arg Val Thr Ala Tyr Lys Glu Gly Tyr	
	1025	1030 1035 1040
5	Gly Glu Gly Cys Val Thr Ile His Glu Ile Glu Asn Asn Thr Asp Glu	
		1045 1050 1055
	Leu Lys Phe Ser Asn Cys Val Glu Glu Glu Ile Tyr Pro Asn Asn Thr	
		1060 1065 1070
10	Val Thr Cys Asn Asp Tyr Thr Val Asn Gln Glu Glu Tyr Gly Gly Ala	
		1075 1080 1085
	Tyr Thr Ser Arg Asn Arg Gly Tyr Asn Glu Ala Pro Ser Val Pro Ala	
		1090 1095 1100
15	Asp Tyr Ala Ser Val Tyr Glu Glu Lys Ser Tyr Thr Asp Gly Arg Arg	
		1105 1110 1115 1120
	Glu Asn Pro Cys Glu Phe Asn Arg Gly Tyr Arg Asp Tyr Thr Pro Leu	
20		1125 1130 1135
	Pro Val Gly Tyr Val Thr Lys Glu Leu Glu Tyr Phe Pro Glu Thr Asp	
		1140 1145 1150
25	Lys Val Trp Ile Glu Ile Gly Glu Thr Glu Gly Thr Phe Ile Val Asp	
		1155 1160 1165
	Ser Val Glu Leu Leu Leu Met Glu Glu	
30		1170 1175

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

TGCAACACTC GAGGCTGAAT

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All of the compositions and methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.